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Address to:

Box Patent Application
Assistant Commissioner for Patents
Washington, D.C. 20231

Attorney's Docket No. REDC-1511 USA

First Named Inventor DOMINIQUE P. BRIDON

UTILITY PATENT APPLICATION TRANSMITTAL
(under 37 CFR 1.53(b))

SIR:

Transmitted herewith for filing is the patent application entitled:

**LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL INFECTION
CERTIFICATION UNDER 37 CFR § 1.10**

I hereby certify that this New Application and the documents referred to as enclosed herein are being deposited with the United States Postal Service on this date September 7, 2000, in an envelope bearing "Express Mail Post Office To Addressee" Mailing Label Number EL254111583US addressed to: Box Patent Application, Assistant Commissioner for Patents, Washington, D.C. 20231.

LANA T. BRENNER

(Name of person mailing paper)

[Signature]
(Signature)

Enclosed are:

1. ☒ Transmittal Form (two copies required)
2. The papers required for filing date under CFR § 1.53(b):
 - i. 170 Pages of specification (including claims and abstract);
 - ii. Sheets of drawings. formal informal
 - iii. 37 Pages of Sequence Listing
3. Declaration or oath
 - a. ☒ Newly executed COMBINED DECLARATION AND POWER OF ATTORNEY (original or copy)
4. Microfiche Computer Program (Appendix, see 37 CFR 1.96)
5. ☒ Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)
 - i. ☒ Computer Readable Copy
 - ii. ☒ Paper Copy (identical to computer copy)
 - iii. ☒ Pursuant to 37 C.F.R. § 1.821(f), the undersigned has reviewed the paper copy and the computer readable copy of the Sequence Listing and determined the information recorded in computer readable form is identical to the written Sequence Listing.

ACCOMPANYING APPLICATION PARTS

6. ☒ An assignment of the invention to CONJUCHEM, INC. is attached (including Form PTO-1595).
 - i. 37 CFR 3.73(b) Statement (when there is an assignee)
7. ☒ POWER OF ATTORNEY (Combined Declaration and Power of Attorney)
8. An Information Disclosure Statement (IDS) is enclosed, including a PTO-1449 and copies of references.
9. Preliminary Amendment.
10. ☒ Return Receipt Postcard (MPEP 503 -- should be specifically itemized)
11. Other

12. AMENDMENT AND PRIORITY CLAIM

☒ Please insert before the Field of Invention at page 1, line 5, the following:

--This application claims the benefit under 35 U.S.C. §119(e) of United States provisional patent application No. 60/153,406 filed September 10, 1999, which is hereby incorporated by reference in its entirety--

13. FEE CALCULATION

a. ☐ Amendment changing number of claims or deleting multiple dependencies is enclosed.

CLAIMS AS FILED

	Number Filed	Number Extra	Rate	Basic Fee (\$690)
Total Claims	30 - 20	* 10	x \$18.00	\$180.00
Independent Claims	5 - 3	* 2	x \$78.00	\$156.00
<input type="checkbox"/> Multiple dependent claim(s), if any			\$260.00	

*If less than zero, enter "0".

Filing Fee Calculation \$1026.00

50% Filing Fee Reduction (if applicable) \$ 513.00

14. Small Entity Status

- a. ☒ A small entity statement is enclosed.
b. ☐ A small entity statement was filed in the prior nonprovisional application and such status is still proper and desired.
c. ☐ is no longer claimed.

15. Other Fees

- ☒ Recording Assignment [\$40.00] \$ 40.00
☐ Other fees
☐ Specify _____ \$

Total Fees Enclosed \$ 553.00

16. Payment of Fees

- ☒ Check(s) in the amount of \$553.00 (INCLUDES ASSIGNMENT FEE) enclosed.
☐ Charge Account No. 12-1420 in the amount of \$____.
A duplicate of this transmittal is attached.

17. All correspondence regarding this application should be forwarded to the undersigned attorney:

Michael R. Ward, Esq.
Limbach & Limbach L.L.P.
2001 Ferry Building
San Francisco, CA 94111
Telephone: 415/433-4150
Facsimile: 415/433-8716

18. Authorization to Charge Additional Fees

- ☒ The Commissioner is hereby authorized to charge any additional fees (or credit any overpayment) associated with this communication and which may be required under 37 CFR § 1.16 or § 1.17 to Account No. 12-1420. A duplicate of this transmittal is attached.

LIMBACH & LIMBACH L.L.P.

September 7, 2000
(Date)

Attorney Docket No. REDC-1511 USA

By: Michael R. Ward
Michael R. Ward, Esq.
Registration No. 38,651
Attorney(s) or Agent(s) of Record

Applicant or Patentee: DOMINIQUE P. BRIDON ET AL.

Appln. or Patent No.:

Attorney's
Docket No.: REDC-1511 USA

Filed or Issued: HEREWITH

For: LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL INFECTION

Express Mail: EL254111583US

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY
STATUS (37 CFR 1.9(f) and 1.27(c)) - SMALL BUSINESS CONCERN**

I hereby declare that I am

☐ the owner of the small business concern identified below:

☒ an official of the small business concern empowered to act on behalf of the concern identified below:

NAME OF CONCERN CONJUCHEM, INC.

ADDRESS OF CONCERN 225 President Kennedy Avenue West, Third Floor, Suite 3950, Montreal, Quebec H2X 3Y8,
Canada

I hereby declare that the above identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention, entitled LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL INFECTION by inventor(s) DOMINIQUE P. BRIDON ET AL. described in

☒ the specification filed herewith with title as listed above.

☐ application no. _____, filed _____.

☐ patent no. _____, issued _____.

If the rights held by the above identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed below* and no rights to the invention are held by any person, other than the inventor, who could not qualify as an independent inventor under 37 CFR 1.9(c) if that person made the invention, or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

*NOTE: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27).

NAME _____

ADDRESS _____

☐ Individual ☐ Small Business Concern ☐ Nonprofit Organization

NAME _____

ADDRESS _____

☐ Individual ☐ Small Business Concern ☐ Nonprofit Organization

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time or paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING Dominique P. Bridon

TITLE OF PERSON OTHER THAN OWNER Vice President of Research

ADDRESS OF PERSON SIGNING 225 President Kennedy Avenue West, Third Floor, Suite 3950, Montreal, Quebec H2X 3Y8, Canada

SIGNATURE: _____

DATE: 15 Aug 2000

LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL INFECTION

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FIELD OF THE INVENTION

This invention relates to modified peptides that are inhibitors of viral activity and/or exhibit antifusogenic properties. In particular, this invention relates to modified peptide inhibitors of human immunodeficiency virus (HIV), respiratory syncytial virus (RSV), human parainfluenza virus (HPV), measles virus (MeV), and simian immunodeficiency virus (SIV) with long duration of action for the treatment of the respective viral infections. The invention also relates to conjugates of the modified peptides and endogenous carriers, particularly conjugates of the modified peptides and various mobile blood components, particularly mobile endogenous proteins.

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BACKGROUND OF THE INVENTION

Membrane fusion events, while commonplace in normal cell biological processes, are also involved in a variety of disease states, including, for example the entry of enveloped viruses into cells. Peptides are known that inhibit or otherwise disrupt membrane fusion-associated events, including, for example, inhibiting retroviral transmission to uninfected cells. As an example, the synthetic peptides DP-107 and DP-178 derived from separate domains within the human immunodeficiency virus type 1 ("HIV-1") transmembrane ("TM") glycoprotein gp41, are potent inhibitors of HIV-1 infection and HIV induced cell-cell fusion.

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Lambert, et al., "Peptides from Conserved Regions of Paramyxovirus Fusion (F) Proteins are Potent Inhibitors of Viral Fusion," Proc. Natl. Acad. Science U.S.A., March 5, 1996, Vol. 93 (5), pp. 2186-91, discloses that the synthetic peptides DP-107 and DP-178 (T-20), derived from separate domains within the human immunodeficiency virus type 1 (HIV-1) transmembrane (TM) protein, gp41, are potent inhibitors of HIV-1 infection and fusion. Using a

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computer searching strategy (computerized antiviral searching technology, C.A.S.T.) based on the predicted secondary structure of DP-107 and DP-178 (T-20), Lambert, et al. identified conserved heptad repeat domains analogous to the DP-107 and DP-178 regions of HIV-1 gp41 within the glycoproteins of other fusogenic viruses. Antiviral peptides derived from three representative paramyxoviruses, respiratory syncytial virus (RSV), human parainfluenza virus type 3 (HPIV-3), and measles virus (MV) blocked homologous virus-mediated syncytium formation and exhibited EC_{50} values in the range 0.015-0.250 μ M. Moreover, these peptides were highly selective for the virus of origin.

U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459 incorporated herein in their entirety, likewise disclose that the 36 amino acid peptide DP178 corresponding to amino acids 638 to 673 of gp41 from the HIV-1 isolate LAI (HIV-1_{LAI}), and the 38 amino acid peptide DP107 corresponding to amino acids 558-595 of gp41 from the HIV-1_{LAI}, both exhibit potent anti-HIV-1 activity.

While many of the anti-viral or anti-fusogenic peptides described in the art exhibit potent anti-viral and/or anti-fusogenic activity, these peptides suffer from short plasma half-lives *in vivo*, primarily due to rapid serum clearance and peptidase and protease activity. This in turn greatly reduces the effective anti-viral activity of the peptides. There is therefore a need for a method of prolonging the half-life of existing anti-viral and/or anti-fusogenic peptides and providing for longer duration of action of these peptides *in vivo*.

SUMMARY OF THE INVENTION

The present invention meets these and other needs and is directed to modified peptides having anti-viral activity and/or anti-fusogenic activity. These modified peptides provide for an increased stability *in vivo* and a reduced susceptibility to peptidase or protease degradation. These modified peptides thereby minimize, e.g., the need for more frequent, or even continual,

administration of the peptides. The products of varying embodiments of the present invention can be used, e.g., as a prophylactic against and/or treatment for infection of a number of viruses, including human immunodeficiency virus (HIV), human respiratory syncytial virus (RSV), human parainfluenza virus (HPV), measles virus (MeV) and simian immunodeficiency virus (SIV). Modification of other peptides involved in viral transfection (e.g., Hepatitis, Epstein Barr and other related viruses) is also within the scope of the invention.

This invention relates to chemically reactive modifications of peptides exhibiting anti-viral and/or anti-fusogenic activity such that the modified peptides can react with available functionalities on blood components to form stable covalent bonds. In one embodiment of the invention, the modified peptides comprise a reactive group which is reactive with amino groups, hydroxyl groups, or thiol groups on blood components to form stable covalent bonds. In another embodiment of the invention, the reactive group can be a maleimide which is reactive with a thiol group on a blood protein, including a mobile blood protein such as albumin.

In particular, the invention relates to such chemically reactive modifications of DP107 and DP178 peptides and analogs thereof, including peptides comprised of amino acid sequences from other (non-HIV) viruses that correspond to the gp41 region of HIV from which DP107 and DP178 are derived and that exhibit anti-viral or anti-fusogenic activity. More particularly, these peptides can exhibit anti-viral activity against, among others, human respiratory syncytial virus (RSV), human parainfluenza virus (HPV), measles virus (MeV) and simian immunodeficiency virus (SIV). The invention also relates to such chemically reactive modifications of the peptides of SEQ ID NO:1 to SEQ ID NO:86.

The invention also relates to compositions for use in the prevention and/or treatment of viral infection comprising a peptide that exhibits anti-viral activity modified with a reactive group as described. More particularly, the invention relates to such compositions for use in the prevention and/or treatment of AIDS,

human respiratory syncytial virus (RSV), human parainfluenza virus (HPV), measles virus (MeV) and simian immunodeficiency virus (SIV).

BRIEF DESCRIPTION OF THE TABLES

5 The invention will be better understood by reference to the Tables, in which:

Table 1 lists the commonly occurring amino acids together with their one letter and three letter abbreviations, and common protecting groups.

Table 2 shows DP178 carboxy truncations.

10 Table 3 shows DP178 amino truncations.

Table 4 shows DP107 carboxy truncations.

Table 5 shows DP107 amino truncations.

Table 6 shows HIV-2_{NIH} DP178 analog carboxy truncations.

Table 7 shows HIV-2_{NIH} DP178 analog amino truncations.

15 Table 8 shows RSV F2 region DP107 analog carboxy truncations.

Table 9 shows RSV F2 region DP107 analog amino truncations.

Table 10 shows RSV F1 region DP178 analog carboxy truncations.

Table 11 shows RSV F1 region DP178 analog amino truncations.

Table 12 shows HPV3 F1 region DP 178 analog carboxy truncations.

20 Table 13 shows HPV3 F1 region DP 178 analog amino truncations.

Table 14 shows HPV3 F1 region DP107 analog carboxy truncations.

Table 15 shows HPV3 F1 region DP107 analog amino truncations.

Table 16 shows representative anti-RSV peptides.

Table 17 shows representative anti-HPV3 peptides.

25 Table 18 shows representative anti-SIV peptides.

Table 19 shows representative anti-MeV peptides.

BRIEF DESCRIPTION OF SEQUENCE LISTING

30 The invention will be better understood by reference to the Sequence Listing, in which:

SEQ ID NO:1 shows the peptide sequence of DP178.

SEQ ID NO:2 shows the peptide sequence of DP107

SEQ ID NO:3-9 show peptide sequences of certain DP178 analogs.

5 SEQ ID NO:10-30 show the peptide sequences of RSV F1 region and F2 region corresponding to DP178 and DP107, and representative anti-RSV peptides;

SEQ ID NO:31-62 show the peptide sequences of HPIV3 F1 region corresponding to DP178 and DP107, and representative anti-HPIV3 peptides;

10 SEQ ID NO:63-73 show peptide sequences of SIV corresponding to DP178 and representative anti-SIV peptides; and

SEQ ID NO:74-78 show peptide sequences of MeV corresponding to DP178 and representative anti-MeV peptides.

DETAILED DESCRIPTION OF THE INVENTION

15 To ensure a complete understanding of the invention the following definitions are provided:

20 **Anti-viral peptides:** As used herein, anti-viral peptides shall refer to peptides that inhibit viral infection of cells, by, for example, inhibiting cell-cell fusion or free virus infection. The route of infection may involve membrane fusion, as occurs in the case of enveloped viruses, or some other fusion event involving viral and cellular structures. Peptides that inhibit viral infection by a particular virus may be referenced with respect to that particular virus, e.g., anti-HIV peptide, anti-RSV peptide, etc.

25 **Antifusogenic peptides:** Antifusogenic peptides are peptides demonstrating an ability to inhibit or reduce the level of membrane fusion events between two or more entities, e.g., virus-cell or cell-cell, relative to the level of membrane fusion that occurs in the absence of the peptide.

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HIV and anti-HIV peptides: The human immunodeficiency virus (HIV), which is responsible for acquired immune deficiency syndrome (AIDS), is a member of the lentivirus family of retroviruses. There are two prevalent types of HIV, HIV-1 and HIV-2, with various strain of each having been identified. HIV targets CD-4+ cells, and viral entry depends on binding of the HIV protein gp41 to CD-4+ cell surface receptors. Anti-HIV peptides refer to peptides that exhibit anti-viral activity against HIV, including inhibiting CD-4+ cell infection by free virus and/or inhibiting HIV-induced syncytia formation between infected and uninfected CD-4+ cells.

SIV and anti-SIV peptides: Simian immunodeficiency viruses (SIV) are lentiviruses that cause acquired immunodeficiency syndrome (AIDS)-like illnesses in susceptible monkeys. Anti-SIV peptides are peptides that exhibit anti-viral activity against SIV, including inhibiting of infection of cells by the SIV virus and inhibiting syncytia formation between infected and uninfected cells.

RSV and anti-RSV peptides: Respiratory syncytial virus (RSV) is a respiratory pathogen, especially dangerous in infants and small children where it can cause bronchiolitis (inflammation of the small air passages) and pneumonia. RSVs are negative sense, single stranded RNA viruses and are members of the *Paramyxoviridae* family of viruses. The route of infection of RSV is typically through the mucous membranes by the respiratory tract, i.e., nose, throat, windpipe and bronchi and bronchioles. Anti-RSV peptides are peptides that exhibit anti-viral activity against RSV, including inhibiting mucous membrane cell infection by free RSV virus and syncytia formation between infection and uninfected cells.

HPV and anti-HPV peptides: Human parainfluenza virus (HPIV or HPV), like RSV, is another leading cause of respiratory tract disease, and like RSVs, are negative sense, single stranded RNA viruses that are members of the *Paramyxoviridae* family of viruses. There are four recognized serotypes of

HPIV -- HPIV-1, HPIV-2, HPIV-3 and HPIV-4. HPIV-1 is the leading cause of croup in children, and both HPIV-1 and HPIV-2 cause upper and lower respiratory tract illnesses. HPIV-3 is more often associated with bronchiolitis and pneumonia. Anti-HPV peptides are peptides that exhibit anti-viral activity against HPV, including inhibiting infection by free HPV virus and syncytia formation between infected and uninfected cells.

MeV and anti-Mev peptides: Measles virus (VM or MeV) is an enveloped negative, single-stranded RNA virus belonging to the *Paramyxoviridae* family of viruses. Like RSV and HPV, MeV causes respiratory disease, and also produces an immuno-suppression responsible for additional, opportunistic infections. In some cases, MeV can establish infection of the brain leading to severe neurological complications. Anti-MeV peptides are peptides that exhibit anti-viral activity against MeV, including inhibiting infection by free MeV virus and syncytia formation between infected and uninfected cells.

DP-178 and DP178 analogs: Unless otherwise indicated explicitly or by context, DP-178 means the 36 amino acid DP-178 peptide corresponding to amino acid residues 638-673 of the gp41 glycoprotein of HIV-1 isolate LAI (HIV_{LAI}) and having the sequence:

YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF (SEQ ID NO:1)

as well as truncations, deletions and/or insertions thereof. Truncations of the DP178 peptide may comprise peptides of between 3-36 amino acids. Deletions consist of the removal of one or more amino acid residues from the DP178 peptide, and may involve the removal of a single contiguous portion of the peptide sequence or multiple portions. Insertions may comprise single amino acid residues or stretches of residues and may be made at the carboxy or amino terminal end of the DP178 peptide or at a position internal to the peptide.

DP178 peptide analogs are peptides whose amino acid sequences are comprised of the amino acid sequences of peptide regions of viruses other than HIV-1_{LAI} that correspond to the gp41 region from which DP178 was derived, as well as an truncations, deletions or insertions thereof. Such other viruses may include, but are not limited to, other HIV isolates such as HIV-2_{NIH2}, respiratory syncytial virus (RSV), human parainfluenza virus (HPV), simian immunodeficiency virus (SIV), and measles virus (MeV). DP178 analogs also refer to those peptide sequences identified or recognized by the ALLMOTI5, 107x178x4 and PLZIP search motifs described in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459 and incorporated herein, having structural and/or amino acid motif similarity to DP178. DP178 analogs further refer to peptides described as "DP178-like" as that term is defined in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459.

DP-107 and DP107 analogs: Unless otherwise indicated explicitly or by context, DP-107 means the 38 amino acid DP-107 peptide corresponding to amino acid residues 558-595 of the gp41 protein of HIV-1 isolate LAI (HIV_{LAI}) and having the sequence:

NNLLRAIEAQQHLLQLTVWQIKQLQARILAVEERYLKDQ (SEQ ID NO:2)

as well as truncations, deletions and/or insertions thereof. Truncations of the DP107 peptide may comprise peptides of between 3-38 amino acids. Deletions consist of the removal of one or more amino acid residues from the DP107 peptide, and may involve the removal of a single contiguous portion of the peptide sequence or multiple portions. Insertions may comprise single amino acid residues or stretches of residues and may be made at the carboxy or amino terminal end of the DP107 peptide or at a position internal to the peptide.

DP107 peptide analogs are peptides whose amino acid sequences are comprised of the amino acid sequences of peptide regions of viruses other than HIV-1_{LAI} that correspond to the gp41 region from which DP107 was derived, as well as truncations, deletions and/or insertions thereof. Such other viruses may include, but are not limited to, other HIV isolates such as HIV-2_{NIH2}, respiratory syncytial virus (RSV), human parainfluenza virus (HPV), simian immunodeficiency virus (SIV), and measles virus (MeV). DP107 analogs also refer to those peptide sequences identified or recognized by the ALLMOTI5, 107x178x4 and PLZIP search motifs described in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459 and incorporated herein, having structural and/or amino acid motif similarity to DP107. DP107 analogs further refer to peptides described as "DP107-like" as that term is defined in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459.

Reactive Groups: Reactive groups are chemical groups capable of forming a covalent bond. Such reactive groups are coupled or bonded to a DP-107 or DP-178 peptide or analogs thereof or other anti-viral or anti-fusogenic peptide of interest. Reactive groups will generally be stable in an aqueous environment and will usually be carboxy, phosphoryl, or convenient acyl group, either as an ester or a mixed anhydride, or an imidate, thereby capable of forming a covalent bond with functionalities such as an amino group, a hydroxy or a thiol at the target site on mobile blood components. For the most part, the esters will involve phenolic compounds, or be thiol esters, alkyl esters, phosphate esters, or the like.

Functionalities: Functionalities are groups on blood components to which reactive groups on modified anti-viral peptides react to form covalent bonds. Functionalities include hydroxyl groups for bonding to ester reactive entities; thiol groups for bonding to maleimides, imidates and thioester groups; amino groups for bonding to carboxy, phosphoryl or acyl groups and carboxyl groups for bonding to amino groups.

Blood Components: Blood components may be either fixed or mobile.

Fixed blood components are non-mobile blood components and include tissues, membrane receptors, interstitial proteins, fibrin proteins, collagens, platelets, endothelial cells, epithelial cells and their associated membrane and membraneous receptors, somatic body cells, skeletal and smooth muscle cells, neuronal components, osteocytes and osteoclasts and all body tissues especially those associated with the circulatory and lymphatic systems. Mobile blood components are blood components that do not have a fixed situs for any extended period of time, generally not exceeding 5, more usually one minute. These blood components are not membrane-associated and are present in the blood for extended periods of time and are present in a minimum concentration of at least 0.1 µg/ml. Mobile blood components include serum albumin, transferrin, ferritin and immunoglobulins such as IgM and IgG. The half-life of mobile blood components is at least about 12 hours.

Protective Groups: Protective groups are chemical moieties utilized to protect peptide derivatives from reacting with themselves. Various protective groups are disclosed herein and in U.S. 5,493,007, which is hereby incorporated by reference. Such protective groups include acetyl, fluorenylmethyloxycarbonyl (Fmoc), t-butyloxycarbonyl (Boc), benzyloxycarbonyl (CBZ), and the like. The specific protected amino acids are depicted in Table 1.

TABLE 1

NATURAL AMINO ACIDS AND THEIR ABBREVIATIONS

Name	3-Letter Abbreviation	1-Letter Abbreviation	Modified Amino Acids
Alanine	Ala	A	Fmoc-Ala-OH
Arginine	Arg	R	Fmoc-Arg(Pbf)-OH
Asparagine	Asn	N	Fmoc-Asn(Trt)-OH
Aspartic acid	Asp	D	Asp(tBu)-OH
Cysteine	Cys	C	Fmoc-Cys(Trt)
Glutamic acid	Glu	E	Fmoc-Glu(tBu)-OH
Glutamine	Gln	Q	Fmoc-Gln(Trt)-OH
Glycine	Gly	G	Fmoc-Gly-OH
Histidine	His	H	Fmoc-His(Trt)-OH
Isoleucine	Ile	I	Fmoc-Ile-OH
Leucine	Leu	L	Fmoc-Leu-OH
Lysine	Lys	Z	Boc-Lys(Aloc)-OH
Lysine	Lys	X	Fmoc-Lys(Aloc)-OH
Lysine	Lys	K	Fmoc-Lys(Mtt)-OH
Methionine	Met	M	Fmoc-Met-OH
Phenylalanine	Phe	F	Fmoc-Phe-OH
Proline	Pro	P	Fmoc-Pro-OH
Serine	Ser	S	Fmoc-Ser(tBu)-OH
Threonine	Thr	T	Fmoc-Thr(tBu)-OH
Tryptophan	Trp	W	Fmoc-Trp(Boc)-OH
Tyrosine	Tyr	Y	Boc-Tyr(tBu)-OH
Valine	Val	V	Fmoc-Val-OH

Linking Groups: Linking (spacer) groups are chemical moieties that link

- 5 or connect reactive entities to antiviral or antifusogenic peptides. Linking groups may comprise one or more alkyl moieties, alkoxy moiety, alkenyl moiety, alkynyl moiety or amino moiety substituted by alkyl moieties, cycloalkyl moiety, polycyclic moiety, aryl moiety, polyaryl moieties, substituted aryl moieties, heterocyclic moieties, and substituted heterocyclic moieties. Linking groups may
- 10 also comprise poly ethoxy amino acids, such as AEA ((2-amino) ethoxy acetic acid) or a preferred linking group AEEA ([2-(2-amino) ethoxy]] ethoxy acetic acid.

Sensitive Functional Groups – A sensitive functional group is a group of atoms that represents a potential reaction site on an antiviral and/or antifusogenic peptide. If present, a sensitive functional group may be chosen as the attachment point for the linker-reactive group modification. Sensitive functional groups include but are not limited to carboxyl, amino, thiol, and hydroxyl groups.

Modified Peptides – A modified peptide is an antiviral and/or antifusogenic peptide that has been modified by attaching a reactive group. The reactive group may be attached to the peptide either via a linking group, or optionally without using a linking group. It is also contemplated that one or more additional amino acids may be added to the peptide to facilitate the attachment of the reactive entity. Modified peptides may be administered *in vivo* such that conjugation with blood components occurs *in vivo*, or they may be first conjugated to blood components *in vitro* and the resulting conjugated peptide (as defined below) administered *in vivo*.

Conjugated Peptides – A conjugated peptide is a modified peptide that has been conjugated to a blood component via a covalent bond formed between the reactive group of the modified peptide and the functionalities of the blood component, with or without a linking group. As used throughout this application, the term “conjugated peptide” can be made more specific to refer to particular conjugated peptides, for example “conjugated DP178” or “conjugated DP107.”

Taking into account these definitions, the present invention takes advantage of the properties of existing anti-viral and antifusogenic peptides. The viruses that may be inhibited by the peptides include, but are not limited to all strains of viruses listed, e.g., in U.S. Patent Nos. 6,013,263, 6,017,536 and 6,020,459 at Tables V-VII and IX-XIV therein. These viruses include, e.g., human retroviruses, including HIV-1, HIV-2, and human T-lymphocyte viruses (HTLV-I and HTLV-II), and non-human retroviruses, including bovine leukosis virus, feline sarcoma virus,

feline leukemia virus, simian immunodeficiency virus (SIV), simian sarcoma virus, simian leukemia, and sheep progress pneumonia virus. Non-retroviral viruses may also be inhibited by the peptides of the present invention, including human respiratory syncytial virus (RSV), canine distemper virus, Newcastle Disease virus, human parainfluenza virus (HPIV), influenza viruses, measles viruses (MeV), Epstein-Barr viruses, hepatitis B viruses, and simian Mason-Pfizer viruses. Non-enveloped viruses may also be inhibited by the peptides of the present invention, and include, but are not limited to, picornaviruses such as polio viruses, hepatitis A virus, enteroviruses, echoviruses, coxsackie viruses, papovaviruses such as papilloma virus, parvoviruses, adenoviruses, and reoviruses.

As an example, the mechanism of action of HIV fusion peptides has been described as discussed in the background section of this application and antiviral and antifusogenic properties of the peptides have been well established. A synthetic peptide corresponding to the carboxyl-terminal ectodomain sequence (for instance, amino acid residues 643-678 of HIV-1 class B, of the LAI strain or residues 638-673 from similar strain as well as residues 558-595) has been shown to inhibit virus-mediated cell-cell fusion completely at low concentration. The fusion peptide competes with the leucine zipper region of the native viral gp41 thus resulting in the interference of the fusion/infection of the virus into the cell.

The focus of the present invention is to modify a selected anti-viral and/or antifusogenic peptide with the DAC (Drug Activity Complex) technology to confer to this peptide improved bio-availability, extended half-life and better distribution through selective conjugation of the peptide onto a protein carrier but without modifying the peptide's anti-viral properties. The carrier of choice (but not limited to) for this invention would be albumin conjugated through its free thiol by an anti-viral and/or antifusogenic peptide modified with a maleimide moiety.

Several peptide sequences have been described in the literature as highly potent for the prevention of HIV-1 fusion/infection. As examples, peptide DP178 binds to a conformation of gp41 that is relevant for fusion. Thus in one embodiment of the invention, DP178 and DP178-like peptides are modified.

Likewise, other embodiments of the invention include modification of DP107 and DP107-like peptide for use against HIV, as well as peptides analogous to DP107 and DP178 that are found in RSV, HPV, MeV and SIV viruses.

5 1. **DP178 and DP107**

A. DP178 Peptides

 The DP178 peptide corresponds to amino acid residues 638 to 673 of the transmembrane protein gp41 from the HIV-1_{LAI} isolate, and has the 36 amino acid
10 sequence (reading from amino to carboxy terminus):

 NH₂-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-COOH (SEQ
 ID NO:1)

15 In addition to the full-length DP178 36-mer, the peptides of this invention include truncations of the DP178 peptide comprising peptides of between 3 and 36 amino acid residues (i.e., peptides ranging in size from a tripeptide to a 36-mer polypeptide), These truncated peptides are shown in Tables 2 and 3.

20 In addition amino acid substitutions of the DP178 peptide are also within the scope of the invention. HIV-1 and HIV-2 enveloped proteins are structurally distinct, but there exists a striking amino acid conservation within the DP178-corresponding regions of HIV-1 and HIV-2. The amino acid conservation is of a periodic nature, suggesting some conservation of structure and/or function.

25 Therefore, one possible class of amino acid substitutions would include those amino acid changes which are predicted to stabilize the structure of the DP178 peptides of the invention. Utilizing the DP178 and DP178 analog sequences described herein, the skilled artisan can readily compile DP178 consensus sequences and ascertain from these, conserved amino acid residues which would
30 represent preferred amino acid substitutions.

The amino acid substitutions may be of a conserved or non-conserved nature. Conserved amino acid substitutions consist of replacing one or more amino acids of the DP178 peptide sequence with amino acids of similar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to aspartic acid (D) amino acid substitution. Non-conserved substitutions consist of replacing one or more amino acids of the DP178 peptide sequence with amino acids possessing dissimilar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to valine (V) substitution.

Amino acid insertions of DP178 may consist of single amino acid residues or stretches of residues. The insertions may be made at the carboxy or amino terminal end of the DP178 or DP178 truncated peptides, as well as at a position internal to the peptide.

Such insertions will generally range from 2 to 15 amino acids in length. It is contemplated that insertions made at either the carboxy or amino terminus of the peptide of interest may be of a broader size range, with about 2 to about 50 amino acids being preferred. One or more such insertions may be introduced into DP178 or DP178 truncations, as long as such insertions result in peptides which may still be recognized by the 107x178x4, ALLMOTI5 or PLZIP search motifs described above.

Preferred amino or carboxy terminal insertions are peptides ranging from about 2 to about 50 amino acid residues in length, corresponding to gp41 protein regions either amino to or carboxy to the actual DP178 gp41 amino acid sequence, respectively. Thus, a preferred amino terminal or carboxy terminal amino acid insertion would contain gp41 amino acid sequences found immediately amino to or carboxy to the DP178 region of the gp41 protein.

Deletions of DP178 or DP178 truncations are also within the scope of this invention. Such deletions consist of the removal of one or more amino acids from the DP178 or DP178-like peptide sequence, with the lower limit length of the resulting peptide sequence being 4 to 6 amino acids.

Such deletions may involve a single contiguous or greater than one discrete portion of the peptide sequences. One or more such deletions may be introduced into DP178 or DP178 truncations, as long as such deletions result in peptides which may still be recognized by the 107x178x4, ALLMOTI5 or PLZIP search motifs described above.

B. DP107 Peptides

DP107 is a 38 amino acid peptide which exhibits potent antiviral activity, and corresponds to residues 558 to 595 of HIV-1_{LAI} isolate transmembrane (TM) gp41 glycoprotein, as shown here:

NH₂-NNLLRAIEAQQHLLQLTVWQIKQLQARILAVEERYLKDQ-COOH
(SEQ ID NO:2)

In addition to the full-length DP107 38-mer, the DP107 peptides include truncations of the DP107 peptide comprising peptides of between 3 and 38 amino acid residues (i.e., peptides ranging in size from a tripeptide to a 38-mer polypeptide). These peptides are shown in Tables 4 and 5, below.

In addition, amino acid substitutions of the DP178 peptide are also within the scope of the invention. As for DP178, there also exists a striking amino acid conservation within the DP107-corresponding regions of HIV-1 and HIV-2, again of a periodic nature, suggesting conservation of structure and/or function. Therefore, one possible class of amino acid substitutions includes those amino acid

changes predicted to stabilize the structure of the DP107 peptides of the invention. Utilizing the DP107 and DP107 analog sequences described herein, the skilled artisan can readily compile DP107 consensus sequences and ascertain from these, conserved amino acid residues which would represent preferred amino acid substitutions.

5

The amino acid substitutions may be of a conserved or non-conserved nature. Conserved amino acid substitutions consist of replacing one or more amino acids of the DP107 peptide sequence with amino acids of similar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to aspartic acid (D) amino acid substitution. Non-conserved substitutions consist of replacing one or more amino acids of the DP107 peptide sequence with amino acids possessing dissimilar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to valine (V) substitution.

10

15

Amino acid insertions may consist of single amino acid residues or stretches of residues. The insertions may be made at the carboxy or amino terminal end of the DP107 or DP107 truncated peptides, as well as at a position internal to the peptide.

20

Such insertions will generally range from 2 to 15 amino acids in length. It is contemplated that insertions made at either the carboxy or amino terminus of the peptide of interest may be of a broader size range, with about 2 to about 50 amino acids being preferred. One or more such insertions may be introduced into DP107 or DP107 truncations, as long as such insertions result in peptides which may still be recognized by the 107x178x4, ALLMOTI5 or PLZIP search motifs described above.

25

30

Preferred amino or carboxy terminal insertions are peptides ranging from about 2 to about 50 amino acid residues in length, corresponding to gp41 protein

native sequences are found, or may exhibit an ability to modulate intracellular processes involving coiled-coil peptide structures.

A. DP178 analogs

5 DP178 analogs are peptides whose amino acid sequences are comprised of the amino acid sequences of peptide regions of, for example, other (i.e., other than HIV-1) viruses that correspond to the gp41 peptide region from which DP178 was derived. Such viruses may include, but are not limited to, other HIV-1 isolates and HIV-2 isolates.

10 DP178 analogs derived from the corresponding gp41 peptide region of other (i.e., non HIV-1LAI) HIV-1 isolates may include, for example, peptide sequences as shown below.

15 NH₂-YTNTIYTLLEESQNQQEKNEQELLELDKWASLWNWF-COOH (SEQ ID NO:3)
NH₂-YTGIIYNLLEESQNQQEKNEQELLELDKWANLWNWF-COOH (SEQ ID NO:4)
20 NH₂-YTSLIYSLLEKSQIQQEKNEQELLELDKWASLWNWF-COOH (SEQ ID NO:5)

The peptides of SEQ ID NO:3, SEQ ID NO:4 and SEQ ID NO:5 are derived from HIV-1_{SF2}, HIV-1_{RF}, and HIV-1_{MN}, respectively. Other DP178 analogs include those derived from HIV-2, including the peptides of SEQ ID NO:6 and
25 SEQ ID NO:7, which are derived from HIV-2_{ROD} and HIV-2_{NIHZ}, respectively. Still other useful analogs include the peptides of SEQ ID NO:8 and SEQ ID NO:9, which have been demonstrated to exhibit anti-viral activity.

30 In the present invention, it is preferred that the DP178 analogs represent peptides whose amino acid sequences correspond to the DP178 region of the gp41

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protein, it is also contemplated that the peptides disclosed herein may, additionally, include amino sequences, ranging from about 2 to about 50 amino acid residues in length, corresponding to gp41 protein regions either amino to or carboxy to the actual DP178 amino acid sequence.

5

Table 6 and Table 7 show some possible truncations of the HIV-2_{NIH2} DP178 analog, which may comprise peptides of between 3 and 36 amino acid residues (i.e., peptides ranging in size from a tripeptide to a 36-mer polypeptide). Peptide sequences in these tables are listed from amino (left) to carboxy (right) terminus.

10

B. Additional DP178 Analogs and DP107 Analogs

DP178 and DP107 analogs are recognized or identified, for example, by utilizing one or more of the 107x178x4, ALLMOTI5 or PLZIP computer-assisted search strategies described above. The search strategy identifies additional peptide regions which are predicted to have structural and/or amino acid sequence features similar to those of DP107 and/or DP178.

15

The search strategies are described fully in the example presented in Section 9 of US Patent Nos. 6,013,263, 6,017,536 and 6,020,459. While this search strategy is based, in part, on a primary amino acid motif deduced from DP107 and DP178, it is not based solely on searching for primary amino acid sequence homologies, as such protein sequence homologies exist within, but not between major groups of viruses. For example, primary amino acid sequence homology is high within the TM protein of different strains of HIV-1 or within the TM protein of different isolates of simian immunodeficiency virus (SIV).

20

25

The computer search strategy disclosed in US Patent Nos. 6,013,263, 6,017,536 and 6,020,459 successfully identified regions of proteins similar to

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DP107 or DP178. This search strategy was designed to be used with a commercially-available sequence database package, preferably PC/Gene.

In US Patent Nos. 6,013,263, 6,017,536 and 6,020,459, a series of search motifs, the 107x178x4, ALLMOTI5 and PLZIP motifs, were designed and engineered to range in stringency from strict to broad, with 107x178x4 being preferred. The sequences identified via such search motifs, such as those listed in Tables V-XIV, of US Patent Nos. 6,013,263, 6,017,536 and 6,020,459 and included in this application by incorporation by reference, potentially exhibit antifusogenic, such as antiviral, activity, may additionally be useful in the identification of antifusogenic, such as antiviral, compounds.

3. Other Anti-Viral Peptides

A. Anti-RSV Peptides

Anti-RSV peptides include DP178 and/or DP107 analogs identified from corresponding peptide sequences in RSV which have further been identified to inhibit viral infection by RSV. Such peptides of interest include the peptides of Table 16 and peptides of SEQ ID NO:10 to SEQ ID NO:30. Of particular interest are the following peptides:

YTSVITIELSNIKENKCNCAKVKLIKQELDKYK (SEQ ID NO:14)
TSVITIELSNIKENKCNCAKVKLIKQELDKYKN (SEQ ID NO:15)
VITIELSNIKENKCNCAKVKLIKQELDKYKNAV (SEQ ID NO:16)

IALSTNKAVVSLNNGVSVLTSKVLDLKNYIDK (SEQ ID NO:29)

The peptide of SEQ ID NO:10 is derived from the F2 region of RSV and was identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP107 and DP178 peptides (i.e., "DP107/178 like").

The peptides of SEQ ID NO:14 to SEQ ID NO:16 each have amino acid sequences contained within the peptide of SEQ ID NO:10 and each has been shown to exhibit anti-RSV activity, in particular, inhibiting fusion and syncytia

formation between RSV-infected and uninfected Hep-2 cells at concentrations of less than 50 µg/ml.

5 The peptide of SEQ ID NO:11 is derived from the F1 region of RSV and was identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP107 (i.e., "DP107-like"). The peptide of SEQ ID NO:29 contains amino acid sequences contained within the peptide of SEQ ID NO:10 and likewise has been shown to exhibit anti-RSV activity, in particular, inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells at concentrations of less than 50 µg/ml.

15 **B. Anti-HPIV Peptides**

Anti-HPIV peptides include DP178 and/or DP107 analogs identified from corresponding peptide sequences in HPIV and which have further been identified to inhibit viral infection by HPIV. Such peptides of interest include the peptides of Table 17 and SEQ ID NO:31 to SEQ ID NO:62. Of particular interest are the following peptides:

20 VEAQQARSDIEKLKEAIRD TNKAVQSVQSSIGNLI (SEQ ID NO:52)
RSDIEKLKEAIRD TNKAVQSVQSSIGNLIVAIKSV (SEQ ID NO:58)

25 NSVALDPIDISIELNKA KSDLEESKEWIRRSNQKL (SEQ ID NO:35)
ALDPIDISIELNKA KSDLEESKEWIRRSNQKLDSI (SEQ ID NO:38)
LDPIDISIELNKA KSDLEESKEWIRRSNQKLDSIG (SEQ ID NO:39)
DPIDISIELNKA KSDLEESKEWIRRSNQKLDSIGN (SEQ ID NO:40)
PIDISIELNKA KSDLEESKEWIRRSNQKLDSIGNW (SEQ ID NO:41)
30 IDISIELNKA KSDLEESKEWIRRSNQKLDSIGNWH (SEQ ID NO:42)

The peptide of SEQ ID NO:31 is derived from the F1 region of HPIV-3 and was identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP107 (i.e., "DP107-like"). The peptides of SEQ

ID NO:52 and SEQ ID NO:58 each have amino acid sequences contained within the peptide of SEQ ID NO:30 and each has been shown to exhibit anti-HPIV-3 activity, in particular, inhibiting fusion and syncytia formation between HPIV-3-infected Hep2 cells and uninfected CV-1W cells at concentrations of less than 1 µg/ml.

The peptide of SEQ ID NO:32 is also derived from the F1 region of HPIV-3 and was identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP178 (i.e., "DP178-like"). The peptides of SEQ ID NO:35 and SEQ ID NO:38 to SEQ ID NO:42 each have amino acid sequences contained within the peptide of SEQ ID NO:32 and each also has been shown to exhibit anti-HPIV-3 activity, in particular, inhibiting fusion and syncytia formation between HPIV-3-infected Hep2 cells and uninfected CV-1W cells at concentrations of less than 1 µg/ml.

C. Anti-MeV Peptides

Anti-MeV peptides are DP178 and/or DP107 analogs identified from corresponding peptide sequences in measles virus (MeV) which have further been identified to inhibit viral infection by the measles virus. Such peptides of particular interest include the peptides of Table 19 and peptides of SEQ ID NO:74 to SEQ ID NO:86. Of particular interest are the peptides listed below.

HRIDLGPPISLERLDVGTNLGNIAIAKLEAKELLE (SEQ ID NO:77)
IDLGPPISLERLDVGTNLGNIAIAKLEAKELLESS (SEQ ID NO:79)
LGPPISLERLDVGTNLGNIAIAKLEAKELLESSDQ (SEQ ID NO:81)
PISLERLDVGTNLGNIAIAKLEAKELLESSDQILR (SEQ ID NO:84)

Sequences derived from measles virus were identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to

DP178 (i.e., "DP178-like"). The peptides of SEQ ID NO:77, SEQ ID NO:79, SEQ ID NO:81 and SEQ ID NO:83 each have amino acid sequences so identified, and each has been shown to exhibit anti-MeV activity, in particular, inhibiting fusion and syncytia formation between MeV-infected Hep2 and uninfected Vero cells at concentrations of less than 1 µg/ml.

D. Anti-SIV Peptides

Anti-SIV peptides are DP178 and/or DP107 analogs identified from corresponding peptide sequences in SIV which have further been identified to inhibit viral infection by SIV. Such peptides of interest include the peptides of Table 18 and peptides of SEQ ID NO:63 to SEQ ID NO:73. Of particular interest are the following peptides:

WQEWERKVD	FLEENITALL	EEAQIQQ	EKNMYELQK	(SEQ ID NO:64)
QEWERKVD	FLEENITALL	EEAQIQQ	EKNMYELQKL	(SEQ ID NO:65)
EWERKVD	FLEENITALL	EEAQIQQ	EKNMYELQKLN	(SEQ ID NO:66)
WERKVD	FLEENITALL	EEAQIQQ	EKNMYELQKLNS	(SEQ ID NO:67)
ERKVD	FLEENITALL	EEAQIQQ	EKNMYELQKLNSW	(SEQ ID NO:68)
RKVD	FLEENITALL	EEAQIQQ	EKNMYELQKLNSW	(SEQ ID NO:69)
KVD	FLEENITALL	EEAQIQQ	EKNMYELQKLNSWDV	(SEQ ID NO:70)
V	FLEENITALL	EEAQIQQ	EKNMYELQKLNSWDVF	(SEQ ID NO:71)
DF	FLEENITALL	EEAQIQQ	EKNMYELQKLNSWDVFG	(SEQ ID NO:72)
FLEENITALL	EEAQIQQ	EKNMYELQKLNSWDVFGN	(SEQ ID NO:73)	

Sequences derived from SIV transmembrane fusion protein were identified in U.S. Patent Nos. 6,103,236 and 6,020,459 using the search motifs described as corresponding to DP178 (i.e., "DP178-like"). The peptides of SEQ ID NO:64 to SEQ ID NO:73 each have amino acid sequences so identified, and each has been shown to exhibit potent anti-SIV activity as crude peptides.

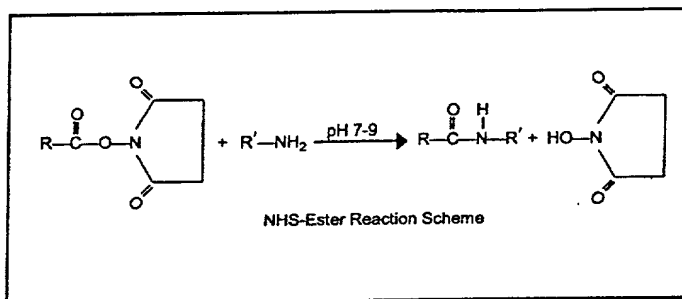
4. Modification of Anti-Viral and Antifusogenic Peptides

The invention contemplates modifying peptides that exhibit anti-viral and/or antifusogenic activity, including such modifications of DP-107 and DP-178 and analogs thereof. Such modified peptides can react with the available reactive

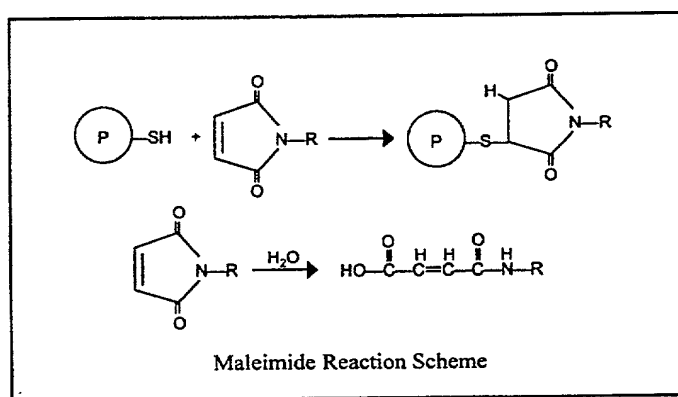
functionalities on blood components via covalent linkages. The invention also relates to such modifications, such combinations with blood components, and methods for their use. These methods include extending the effective therapeutic life of the conjugated anti-viral peptides derivatives as compared to administration of the unconjugated peptides to a patient. The modified peptides are of a type designated as a DAC (Drug Affinity Complex) which comprises the anti-viral peptide molecule and a linking group together with a chemically reactive group capable of reaction with a reactive functionality of a mobile blood protein. By reaction with the blood component or protein the modified peptide, or DAC, may be delivered via the blood to appropriate sites or receptors.

To form covalent bonds with functionalities on the protein, one may use as a reactive group a wide variety of active carboxyl groups, particularly esters, where the hydroxyl moiety is physiologically acceptable at the levels required to modify the peptide. While a number of different hydroxyl groups may be employed in these reactive groups, the most convenient would be N-hydroxysuccinimide or (NHS), N-hydroxy-sulfosuccinimide (sulfo-NHS). In preferred embodiments of this invention, the functionality on the protein will be a thiol group and the reactive group will be a maleimido-containing group such as gamma-maleimide-butyralamide (GMBA) or maleimidopropionic acid (MPA)

Primary amines are the principal targets for NHS esters. Accessible α -amine groups present on the N-termini of proteins react with NHS esters. However, α -amino groups on a protein may not be desirable or available for the NHS coupling. While five amino acids have nitrogen in their side chains, only the ϵ -amine of lysine reacts significantly with NHS esters. An amide bond is formed when the NHS ester conjugation reaction reacts with primary amines releasing N-hydroxysuccinimide as demonstrated in the schematic below.



In the preferred embodiments of this invention, the functional group on this protein will be a thiol group and the chemically reactive group will be a maleimido-containing group such as MPA or GMBA (gamma-maleimide-butylalamide). The maleimido group is most selective for sulfhydryl groups on peptides when the pH of the reaction mixture is kept between 6.5 and 7.4. At pH 7.0, the rate of reaction of maleimido groups with sulfhydryls is 1000-fold faster than with amines. A stable thioether linkage between the maleimido group and the sulfhydryl is formed which cannot be cleaved under physiological conditions, as demonstrated in the following schematic.



A. Specific Labeling.

Preferably, the modified peptides of this invention are designed to specifically react with thiol groups on mobile blood proteins. Such reaction is preferably established by covalent bonding of the peptide modified with a

maleimide link (e.g. prepared from GMBS, MPA or other maleimides) to a thiol group on a mobile blood protein such as serum albumin or IgG.

Under certain circumstances, specific labeling with maleimides offers several advantages over non-specific labeling of mobile proteins with groups such as NHS and sulfo-NHS. Thiol groups are less abundant *in vivo* than amino groups. Therefore, the maleimide-modified peptides of this invention, i.e., maleimide peptides, will covalently bond to fewer proteins. For example, in albumin (the most abundant blood protein) there is only a single thiol group. Thus, peptide-maleimide-albumin conjugates will tend to comprise approximately a 1:1 molar ratio of peptide to albumin. In addition to albumin, IgG molecules (class II) also have free thiols. Since IgG molecules and serum albumin make up the majority of the soluble protein in blood they also make up the majority of the free thiol groups in blood that are available to covalently bond to maleimide-modified peptides.

Further, even among free thiol-containing blood proteins, including IgGs, specific labeling with maleimides leads to the preferential formation of peptide-maleimide-albumin conjugates, due to the unique characteristics of albumin itself. The single free thiol group of albumin, highly conserved among species, is located at amino acid residue 34 (Cys³⁴). It has been demonstrated recently that the Cys³⁴ of albumin has increased reactivity relative to free thiols on other free thiol-containing proteins. This is due in part to the very low pK value of 5.5 for the Cys³⁴ of albumin. This is much lower than typical pK values for cysteine residues in general, which are typically about 8. Due to this low pK, under normal physiological conditions Cys³⁴ of albumin is predominantly in the ionized form, which dramatically increases its reactivity. In addition to the low pK value of Cys³⁴, another factor which enhances the reactivity of Cys³⁴ is its location, which is in a crevice close to the surface of one loop of region V of albumin. This location makes Cys³⁴ very available to ligands of all kinds, and is an important factor in Cys³⁴'s biological role as free radical trap and free thiol scavenger. These properties make Cys³⁴ highly reactive with maleimide-peptides, and the reaction

rate acceleration can be as much as 1000-fold relative to rates of reaction of maleimide-peptides with other free-thiol containing proteins.

Another advantage of peptide-maleimide-albumin conjugates is the reproducibility associated with the 1:1 loading of peptide to albumin specifically at Cys³⁴. Other techniques, such as glutaraldehyde, DCC, EDC and other chemical activations of, e.g, free amines, lack this selectivity. For example, albumin contains 52 lysine residues, 25-30 of which are located on the surface of albumin and therefore accessible for conjugation. Activating these lysine residues, or alternatively modifying peptides to couple through these lysine residues, results in a heterogenous population of conjugates. Even if 1:1 molar ratios of peptide to albumin are employed, the yield will consist of multiple conjugation products, some containing 0, 1, 2 or more peptides per albumin, and each having peptides randomly coupled at any one or more of the 25-30 available lysine sites. Given the numerous possible combinations, characterization of the exact composition and nature of each conjugate batch becomes difficult, and batch-to-batch reproducibility is all but impossible, making such conjugates less desirable as a therapeutic. Additionally, while it would seem that conjugation through lysine residues of albumin would at least have the advantage of delivering more therapeutic agent per albumin molecule, studies have shown that a 1:1 ratio of therapeutic agent to albumin is preferred. In an article by Stehle, et al., "The Loading Rate Determines Tumor Targeting properties of Methotrexate-Albumin Conjugates in Rats," Anti-Cancer Drugs, Vol. 8, pp. 677-685 (1988), incorporated herein in its entirety, the authors report that a 1:1 ratio of the anti-cancer methotrexate to albumin conjugated via glutaraldehyde gave the most promising results. These conjugates were preferentially taken up by tumor cells, whereas conjugates bearing 5:1 to 20:1 methotrexate molecules had altered HPLC profiles and were quickly taken up by the liver *in vivo*. It is postulated that at these higher ratios, conformational changes to albumin diminish its effectiveness as a therapeutic carrier.

Through controlled administration of maleimide-peptides *in vivo*, one can control the specific labeling of albumin and IgG *in vivo*. In typical administrations, 80-90% of the administered maleimide-peptides will label albumin and less than 5% will label IgG. Trace labeling of free thiols such as glutathione will also occur. Such specific labeling is preferred for *in vivo* use as it permits an accurate calculation of the estimated half-life of the administered agent.

In addition to providing controlled specific *in vivo* labeling, maleimide-peptides can provide specific labeling of serum albumin and IgG *ex vivo*. Such *ex vivo* labeling involves the addition of maleimide-peptides to blood, serum or saline solution containing serum albumin and/or IgG. Once conjugation has occurred *ex vivo* with the maleimide-peptides, the blood, serum or saline solution can be readministered to the patient's blood for *in vivo* treatment.

In contrast to NHS-peptides, maleimide-peptides are generally quite stable in the presence of aqueous solutions and in the presence of free amines. Since maleimide-peptides will only react with free thiols, protective groups are generally not necessary to prevent the maleimide-peptides from reacting with itself. In addition, the increased stability of the modified peptide permits the use of further purification steps such as HPLC to prepare highly purified products suitable for *in vivo* use. Lastly, the increased chemical stability provides a product with a longer shelf life.

B. Non-Specific Labeling.

The anti-viral peptides of the invention may also be modified for non-specific labeling of blood components. Bonds to amino groups will also be employed, particularly with the formation of amide bonds for non-specific labeling. To form such bonds, one may use as a chemically reactive group a wide variety of active carboxyl groups, particularly esters, where the hydroxyl moiety is physiologically acceptable at the levels required. While a number of different hydroxyl groups may be employed in these linking agents, the most convenient would be N-hydroxysuccinimide (NHS) and N-hydroxy-sulfosuccinimide (sulfo-

NHS).

Other linking agents which may be utilized are described in U.S. Patent 5,612,034, which is hereby incorporated herein.

5 The various sites with which the chemically reactive group of the modified peptides may react *in vivo* include cells, particularly red blood cells (erythrocytes) and platelets, and proteins, such as immunoglobulins, including IgG and IgM, serum albumin, ferritin, steroid binding proteins, transferrin, thyroxin binding protein, α -2-macroglobulin, and the like. Those receptors with which the modified peptides react, which are not long-lived, will generally be eliminated
10 from the human host within about three days. The proteins indicated above (including the proteins of the cells) will remain at least three days, and may remain five days or more (usually not exceeding 60 days, more usually not exceeding 30 days) particularly as to the half life, based on the concentration in the blood.

15 For the most part, reaction will be with mobile components in the blood, particularly blood proteins and cells, more particularly blood proteins and erythrocytes. By "mobile" is intended that the component does not have a fixed situs for any extended period of time, generally not exceeding 5 minutes, more usually one minute, although some of the blood component may be relatively stationary for extended periods of time. Initially, there will be a relatively
20 heterogeneous population of functionalized proteins and cells. However, for the most part, the population within a few days will vary substantially from the initial population, depending upon the half-life of the functionalized proteins in the blood stream. Therefore, usually within about three days or more, IgG will become the predominant functionalized protein in the blood stream.

25 Usually, by day 5 post-administration, IgG, serum albumin and erythrocytes will be at least about 60 mole %, usually at least about 75 mole %, of the conjugated components in blood, with IgG, IgM (to a substantially lesser extent) and serum albumin being at least about 50 mole %, usually at least about 75 mole %, more usually at least about 80 mole %, of the non-cellular conjugated
30 components.

The desired conjugates of non-specific modified peptides to blood components may be prepared *in vivo* by administration of the modified peptides to the patient, which may be a human or other mammal. The administration may be done in the form of a bolus or introduced slowly over time by infusion using metered flow or the like.

If desired, the subject conjugates may also be prepared *ex vivo* by combining blood with modified peptides of the present invention, allowing covalent bonding of the modified peptides to reactive functionalities on blood components and then returning or administering the conjugated blood to the host. Moreover, the above may also be accomplished by first purifying an individual blood component or limited number of components, such as red blood cells, immunoglobulins, serum albumin, or the like, and combining the component or components *ex vivo* with the chemically reactive modified peptides. The functionalized blood or blood component may then be returned to the host to provide *in vivo* the subject therapeutically effective conjugates. The blood also may be treated to prevent coagulation during handling *ex vivo*.

5. Synthesis of Modified Anti-Viral and Anti-Fusogenic Peptides

A. Peptide Synthesis

Anti-viral and/or anti-fusogenic peptides according to the present invention may be synthesized by standard methods of solid phase peptide chemistry known to those of ordinary skill in the art. For example, peptides may be synthesized by solid phase chemistry techniques following the procedures described by Steward and Young (Steward, J. M. and Young, J. D., Solid Phase Peptide Synthesis, 2nd Ed., Pierce Chemical Company, Rockford, Ill., (1984) using an Applied Biosystem synthesizer. Similarly, multiple peptide fragments may be synthesized then linked together to form larger peptides. These synthetic peptides can also be made with amino acid substitutions at specific locations.

For solid phase peptide synthesis, a summary of the many techniques may be found in J. M. Stewart and J. D. Young, Solid Phase Peptide Synthesis, W. H.

Freeman Co. (San Francisco), 1963 and J. Meienhofer, Hormonal Proteins and Peptides, vol. 2, p. 46, Academic Press (New York), 1973. For classical solution synthesis see G. Schroder and K. Lupke, The Peptides, Vol. 1, Academic Press (New York). In general, these methods comprise the sequential addition of one or
5 more amino acids or suitably protected amino acids to a growing peptide chain. Normally, either the amino or carboxyl group of the first amino acid is protected by a suitable protecting group. The protected or derivatized amino acid is then either attached to an inert solid support or utilized in solution by adding the next amino acid in the sequence having the complimentary (amino or carboxyl) group
10 suitably protected and under conditions suitable for forming the amide linkage. The protecting group is then removed from this newly added amino acid residue and the next amino acid (suitably protected) is added, and so forth.

After all the desired amino acids have been linked in the proper sequence, any remaining protecting groups (and any solid support) are removed sequentially
15 or concurrently to afford the final polypeptide. By simple modification of this general procedure, it is possible to add more than one amino acid at a time to a growing chain, for example, by coupling (under conditions which do not racemize chiral centers) a protected tripeptide with a properly protected dipeptide to form, after deprotection, a pentapeptide.

A particularly preferred method of preparing compounds of the present invention involves solid phase peptide synthesis wherein the amino acid α -N-terminal is protected by an acid or base sensitive group. Such protecting groups should have the properties of being stable to the conditions of peptide linkage formation while being readily removable without destruction of the growing
20 peptide chain or racemization of any of the chiral centers contained therein. Suitable protecting groups are 9-fluorenylmethyloxycarbonyl (Fmoc), t-butylloxycarbonyl (Boc), benzyloxycarbonyl (Cbz), biphenylisopropylloxycarbonyl, t-amylloxycarbonyl, isobornylloxycarbonyl, α , α -dimethyl-3,5-dimethoxybenzyloxycarbonyl, o-nitrophenylsulfonyl, 2-cyano-t-butylloxycarbonyl,
25 and the like. The 9-fluorenyl-methyloxycarbonyl (Fmoc) protecting group is
30

particularly preferred for the synthesis of the peptides of the present invention. Other preferred side chain protecting groups are, for side chain amino groups like lysine and arginine, 2,2,5,7,8-pentamethylchroman-6-sulfonyl (pmc), nitro, p-toluenesulfonyl, 4-methoxybenzene-sulfonyl, Cbz, Boc, and
5 adamantyloxycarbonyl; for tyrosine, benzyl, o-bromobenzyloxycarbonyl, 2,6-dichlorobenzyl, isopropyl, t-butyl (t-Bu), cyclohexyl, cyclopropyl and acetyl (Ac); for serine, t-butyl, benzyl and tetrahydropyranyl; for histidine, trityl, benzyl, Cbz, p-toluenesulfonyl and 2,4-dinitrophenyl; for tryptophan, formyl; for aspartic acid and glutamic acid, benzyl and t-butyl and for cysteine, triphenylmethyl (trityl).

10 In the solid phase peptide synthesis method, the α -C-terminal amino acid is attached to a suitable solid support or resin. Suitable solid supports useful for the above synthesis are those materials which are inert to the reagents and reaction conditions of the stepwise condensation-deprotection reactions, as well as being insoluble in the media used. The preferred solid support for synthesis of α -C-
15 terminal carboxy peptides is 4-hydroxymethylphenoxymethyl-copoly(styrene-1% divinylbenzene). The preferred solid support for α -C-terminal amide peptides is the 4-(2',4'-dimethoxyphenyl-Fmoc-aminomethyl)phenoxyacetamidoethyl resin available from Applied Biosystems (Foster City, Calif.). The α -C-terminal amino acid is coupled to the resin by means of N,N'-dicyclohexylcarbodiimide (DCC),
20 N,N'-diisopropylcarbodiimide (DIC) or O-benzotriazol-1-yl-N,N,N',N'-tetramethyluronium-hexafluorophosphate (HBTU), with or without 4-dimethylaminopyridine (DMAP), 1-hydroxybenzotriazole (HOBT), benzotriazol-1-yloxy-tris(dimethylamino)phosphonium-hexafluorophosphate (BOP) or bis(2-oxo-3-oxazolidinyl)phosphine chloride (BOPCl), mediated coupling for from
25 about 1 to about 24 hours at a temperature of between 10° and 50°C in a solvent such as dichloromethane or DMF.

When the solid support is 4-(2',4'-dimethoxyphenyl-Fmoc-aminomethyl)phenoxy-acetamidoethyl resin, the Fmoc group is cleaved with a secondary amine, preferably piperidine, prior to coupling with the α -C-terminal
30 amino acid as described above. The preferred method for coupling to the

deprotected 4-(2',4'-dimethoxyphenyl-Fmoc-aminomethyl)phenoxy-
acetamidoethyl resin is O-benzotriazol-1-yl-N,N,N',N'-
tetramethyluroniumhexafluoro-phosphate (HBTU, 1 equiv.) and 1-
hydroxybenzotriazole (HOBT, 1 equiv.) in DMF. The coupling of successive
5 protected amino acids can be carried out in an automatic polypeptide synthesizer
as is well known in the art. In a preferred embodiment, the α -N-terminal amino
acids of the growing peptide chain are protected with Fmoc. The removal of the
Fmoc protecting group from the α -N-terminal side of the growing peptide is
accomplished by treatment with a secondary amine, preferably piperidine. Each
10 protected amino acid is then introduced in about 3-fold molar excess, and the
coupling is preferably carried out in DMF. The coupling agent is normally O-
benzotriazol-1-yl-N,N,N',N'-tetramethyluroniumhexafluorophosphate (HBTU, 1
equiv.) and 1-hydroxybenzotriazole (HOBT, 1 equiv.).

At the end of the solid phase synthesis, the polypeptide is removed from
15 the resin and deprotected, either in successively or in a single operation. Removal
of the polypeptide and deprotection can be accomplished in a single operation by
treating the resin-bound polypeptide with a cleavage reagent comprising
thioanisole, water, ethanedithiol and trifluoroacetic acid. In cases wherein the α -
C-terminal of the polypeptide is an alkylamide, the resin is cleaved by aminolysis
20 with an alkylamine. Alternatively, the peptide may be removed by
transesterification, e.g. with methanol, followed by aminolysis or by direct
transamidation. The protected peptide may be purified at this point or taken to the
next step directly. The removal of the side chain protecting groups is accomplished
using the cleavage cocktail described above. The fully deprotected peptide is
25 purified by a sequence of chromatographic steps employing any or all of the
following types: ion exchange on a weakly basic resin (acetate form); hydrophobic
adsorption chromatography on underivitized polystyrene-divinylbenzene (for
example, Amberlite XAD); silica gel adsorption chromatography; ion exchange
chromatography on carboxymethylcellulose; partition chromatography, e.g. on
30 Sephadex G-25, LH-20 or countercurrent distribution; high performance liquid

chromatography (HPLC), especially reverse-phase HPLC on octyl- or octadecylsilyl-silica bonded phase column packing.

Molecular weights of these ITPs are determined using Fast Atom Bombardment (FAB) Mass Spectroscopy.

5

(1) N-Terminal Protective Groups

As discussed above, the term "N-protecting group" refers to those groups intended to protect the α -N-terminal of an amino acid or peptide or to otherwise protect the amino group of an amino acid or peptide against undesirable reactions during synthetic procedures. Commonly used N-protecting groups are disclosed in Greene, "Protective Groups In Organic Synthesis," (John Wiley & Sons, New York (1981)), which is hereby incorporated by reference. Additionally, protecting groups can be used as pro-drugs which are readily cleaved *in vivo*, for example, by enzymatic hydrolysis, to release the biologically active parent. α -N-protecting groups comprise loweralkanoyl groups such as formyl, acetyl ("Ac"), propionyl, pivaloyl, t-butylacetyl and the like; other acyl groups include 2-chloroacetyl, 2-bromoacetyl, trifluoroacetyl, trichloroacetyl, phthalyl, o-nitrophenoxyacetyl, -chlorobutyryl, benzoyl, 4-chlorobenzoyl, 4-bromobenzoyl, 4-nitrobenzoyl and the like; sulfonyl groups such as benzenesulfonyl, p-toluenesulfonyl and the like; carbamate forming groups such as benzyloxycarbonyl, p-chlorobenzyloxycarbonyl, p-methoxybenzyloxycarbonyl, p-nitrobenzyloxycarbonyl, 2-nitrobenzyloxycarbonyl, p-bromobenzyloxycarbonyl, 3,4-dimethoxybenzyloxycarbonyl, 3,5-dimethoxybenzyloxycarbonyl, 2,4-dimethoxybenzyloxycarbonyl, 4-ethoxybenzyloxycarbonyl, 2-nitro-4,5-dimethoxybenzyloxycarbonyl, 3,4,5-trimethoxybenzyloxycarbonyl, 1-(p-biphenyl)-1-methylethoxycarbonyl, α,α -dimethyl-3,5-dimethoxybenzyloxycarbonyl, benzhydryloxycarbonyl, t-butyloxycarbonyl (Boc), diisopropylmethoxycarbonyl, isopropylloxycarbonyl, ethoxycarbonyl, methoxycarbonyl, allyloxycarbonyl, 2,2,2-trichloroethoxycarbonyl, phenoxycarbonyl, 4-nitrophenoxy carbonyl, fluorenyl-9-methoxycarbonyl,

cyclopentyloxycarbonyl, adamantyloxycarbonyl, cyclohexyloxycarbonyl, phenylthiocarbonyl and the like; arylalkyl groups such as benzyl, triphenylmethyl, benzyloxymethyl, 9-fluorenylmethyloxycarbonyl (Fmoc) and the like and silyl groups such as trimethylsilyl and the like.

5

(2) Carboxy Protective Groups

As discussed above, the term "carboxy protecting group" refers to a carboxylic acid protecting ester or amide group employed to block or protect the carboxylic acid functionality while the reactions involving other functional sites of the compound are performed. Carboxy protecting groups are disclosed in Greene, "Protective Groups in Organic Synthesis" pp. 152-186 (1981), which is hereby incorporated by reference. Additionally, a carboxy protecting group can be used as a pro-drug whereby the carboxy protecting group can be readily cleaved *in vivo*, for example by enzymatic hydrolysis, to release the biologically active parent.

Such carboxy protecting groups are well known to those skilled in the art, having been extensively used in the protection of carboxyl groups in the penicillin and cephalosporin fields as described in U.S. Pat. Nos. 3,840,556 and 3,719,667, the disclosures of which are hereby incorporated herein by reference. Representative carboxy protecting groups are C₁-C₈ loweralkyl (e.g., methyl, ethyl or t-butyl and the like); arylalkyl such as phenethyl or benzyl and substituted derivatives thereof such as alkoxybenzyl or nitrobenzyl groups and the like; arylalkenyl such as phenylethenyl and the like; aryl and substituted derivatives thereof such as 5-indanyl and the like; dialkylaminoalkyl such as dimethylaminoethyl and the like; alkanoyloxyalkyl groups such as acetoxymethyl, butyryloxymethyl, valeryloxymethyl, isobutyryloxymethyl, isovaleryloxymethyl, 1-(propionyloxy)-1-ethyl, 1-(pivaloyloxy)-1-ethyl, 1-methyl-1-(propionyloxy)-1-ethyl, pivaloyloxymethyl, propionyloxymethyl and the like; cycloalkanoyloxyalkyl groups such as cyclopropylcarbonyloxymethyl, cyclobutylcarbonyloxymethyl, cyclopentylcarbonyloxymethyl, cyclohexylcarbonyloxymethyl and the like; aroyloxyalkyl such as benzoyloxymethyl, benzoyloxyethyl and the like;

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arylalkylcarbonyloxyalkyl such as benzylcarbonyloxymethyl, 2-benzylcarbonyloxyethyl and the like; alkoxycarbonylalkyl or cycloalkyloxycarbonylalkyl such as methoxycarbonylmethyl, cyclohexyloxycarbonylmethyl, 1-methoxycarbonyl-1-ethyl and the like;

5 alkoxycarbonyloxyalkyl or cycloalkyloxycarbonyloxyalkyl such as methoxycarbonyloxymethyl, t-butyloxycarbonyloxymethyl, 1-ethoxycarbonyloxy-1-ethyl, 1-cyclohexyloxycarbonyloxy-1-ethyl and the like;

aryloxy carbonyloxyalkyl such as 2-(phenoxy carbonyloxy)ethyl, 2-(5-indanyloxy carbonyloxy)ethyl and the like; alkoxyalkylcarbonyloxyalkyl such as 2-

10 (1-methoxy-2-methylpropan-2-oyloxy)ethyl and like;

arylalkyloxycarbonyloxyalkyl such as 2-(benzyloxycarbonyloxy)ethyl and the like;

arylalkenyloxycarbonyloxyalkyl such as 2-(3-phenylpropen-2-yloxycarbonyloxy)ethyl and the like; alkoxycarbonylaminoalkyl such as t-

butyloxycarbonylaminomethyl and the like; alkylaminocarbonylaminoalkyl such

15 as methylaminocarbonylaminomethyl and the like; alkanoylaminoalkyl such as acetylaminomethyl and the like; heterocyclic carbonyloxyalkyl such as 4-methylpiperazinylcarbonyloxymethyl and the like; dialkylaminocarbonylalkyl

such as dimethylaminocarbonylmethyl, diethylaminocarbonylmethyl and the like;

(5-(loweralkyl)-2-oxo-1,3-dioxolen-4-yl)alkyl such as (5-t-butyl-2-oxo-1,3-

20 dioxolen-4-yl)methyl and the like; and (5-phenyl-2-oxo-1,3-dioxolen-4-yl)alkyl

such as (5-phenyl-2-oxo-1,3-dioxolen-4-yl)methyl and the like.

Representative amide carboxy protecting groups are aminocarbonyl and loweralkylaminocarbonyl groups.

25 Preferred carboxy-protected compounds of the invention are compounds wherein the protected carboxy group is a loweralkyl, cycloalkyl or arylalkyl ester, for example, methyl ester, ethyl ester, propyl ester, isopropyl ester, butyl ester, sec-butyl ester, isobutyl ester, amyl ester, isoamyl ester, octyl ester, cyclohexyl ester, phenylethyl ester and the like or an alkanoyloxyalkyl, cycloalkanoyloxyalkyl,

30 aroyloxyalkyl or an arylalkylcarbonyloxyalkyl ester. Preferred amide carboxy

protecting groups are loweralkylaminocarbonyl groups. For example, aspartic acid may be protected at the α -C-terminal by an acid labile group (e.g. t-butyl) and protected at the β -C-terminal by a hydrogenation labile group (e.g. benzyl) then deprotected selectively during synthesis.

5

B. Peptide Modification

The manner of producing the modified peptides of the present invention will vary widely, depending upon the nature of the various elements comprising the peptide. The synthetic procedures will be selected so as to be simple, provide for high yields, and allow for a highly purified stable product. Normally, the chemically reactive group will be created at the last stage of the synthesis, for example, with a carboxyl group, esterification to form an active ester. Specific methods for the production of modified peptides of the present invention are described below.

Specifically, the selected peptide is first assayed for anti-viral activity, and then is modified with the linking group only at either the N-terminus, C-terminus or interior of the peptide. The anti-viral activity of this modified peptide-linking group is then assayed. If the anti-viral activity is not reduced dramatically (i.e., reduced less than 10-fold), then the stability of the modified peptide-linking group is measured by its *in vivo* lifetime. If the stability is not improved to a desired level, then the peptide is modified at an alternative site, and the procedure is repeated until a desired level of anti-viral and stability is achieved.

More specifically, each peptide selected to undergo modification with a linker and a reactive entity group will be modified according to the following criteria: if a terminal carboxylic group is available on the peptide and is not critical for the retention of anti-viral activity, and no other sensitive functional group is present on the peptide, then the carboxylic acid will be chosen as attachment point for the linker-reactive group modification. If the terminal carboxylic group is involved in anti-viral activity, or if no carboxylic acids are available, then any other sensitive functional group not critical for the retention of anti-viral activity

will be selected as the attachment point for the linker-reactive entity modification. If several sensitive functional groups are available on a peptide, a combination of protecting groups will be used in such a way that after addition of the linker/reactive entity and deprotection of all the protected sensitive functional groups, retention of anti-viral activity is still obtained. If no sensitive functional groups are available on the peptide, or if a simpler modification route is desired, synthetic efforts will allow for a modification of the original peptide in such a way that retention of anti-viral is maintained. In this case the modification will occur at the opposite end of the peptide

An NHS derivative may be synthesized from a carboxylic acid in absence of other sensitive functional groups in the peptide. Specifically, such a peptide is reacted with N-hydroxysuccinimide in anhydrous CH_2Cl_2 and EDC, and the product is purified by chromatography or recrystallized from the appropriate solvent system to give the NHS derivative.

Alternatively, an NHS derivative may be synthesized from a peptide that contains an amino and/or thiol group and a carboxylic acid. When a free amino or thiol group is present in the molecule, it is preferable to protect these sensitive functional groups prior to perform the addition of the NHS derivative. For instance, if the molecule contains a free amino group, a transformation of the amine into a Fmoc or preferably into a tBoc protected amine is necessary prior to perform the chemistry described above. The amine functionality will not be deprotected after preparation of the NHS derivative. Therefore this method applies only to a compound whose amine group is not required to be freed to induce the desired anti-viral effect. If the amino group needs to be freed to retain the original properties of the molecule, then another type of chemistry described below has to be performed.

In addition, an NHS derivative may be synthesized from a peptide containing an amino or a thiol group and no carboxylic acid. When the selected molecule contains no carboxylic acid, an array of bifunctional linkers can be used to convert the molecule into a reactive NHS derivative. For instance, ethylene

glycol-bis(succinimidyldisuccinate) (EGS) and triethylamine dissolved in DMF and added to the free amino containing molecule (with a ratio of 10:1 in favor of EGS) will produce the mono NHS derivative. To produce an NHS derivative from a thiol derivatized molecule, one can use N-[γ -maleimidobutyryloxy]succinimide ester (GMBS) and triethylamine in DMF. The maleimido group will react with the free thiol and the NHS derivative will be purified from the reaction mixture by chromatography on silica or by HPLC.

An NHS derivative may also be synthesized from a peptide containing multiple sensitive functional groups. Each case will have to be analyzed and solved in a different manner. However, thanks to the large array of protecting groups and bifunctional linkers that are commercially available, this invention is applicable to any peptide with preferably one chemical step only to modify the peptide (as described above) or two steps (as described above involving prior protection of a sensitive group) or three steps (protection, activation and deprotection). Under exceptional circumstances only, would multiple steps (beyond three steps) synthesis be required to transform a peptide into an active NHS or maleimide derivative.

A maleimide derivative may also be synthesized from a peptide containing a free amino group and a free carboxylic acid. To produce a maleimide derivative from a amino derivatized molecule, one can use N-[γ -maleimidobutyryloxy]succinimide ester (GMBS) and triethylamine in DMF. The succinimide ester group will react with the free amino and the maleimide derivative will be purified from the reaction mixture by crystallization or by chromatography on silica or by HPLC.

Finally, a maleimide derivative may be synthesized from a peptide containing multiple other sensitive functional groups and no free carboxylic acids. When the selected molecule contains no carboxylic acid, an array of bifunctional crosslinking reagents can be used to convert the molecule into a reactive NHS derivative. For instance maleimidopropionic acid (MPA) can be coupled to the free amine to produce a maleimide derivative through reaction of the free amine

with the carboxylic group of MPA using HBTU/HOBt/DIEA activation in DMF.

Many other commercially available heterobifunctional crosslinking reagents can alternatively be used when needed. A large number of bifunctional compounds are available for linking to entities. Illustrative reagents include:

5 azidobenzoyl hydrazide, N-[4-(p-azidosalicylamino)butyl]-3'-[2'-pyridyldithio)propionamide), bis-sulfosuccinimidyl suberate, dimethyl adipimide, disuccinimidyl tartrate, N-y-maleimidobutyryloxysuccinimide ester, N-hydroxy sulfosuccinimidyl-4-azidobenzoate, N-succinimidyl [4-azidophenyl]-
10 1,3'-dithiopropionate, N-succinimidyl [4-iodoacetyl]aminobenzoate, glutaraldehyde, and succinimidyl 4-[N-maleimidomethyl]cyclohexane-1-carboxylate.

6. Uses of Modified Anti-Viral Peptides

15 Modified anti-viral peptides of the invention may be used as a therapeutic agent in the treatment of patients who are suffering from viral infection, and can be administered to patients according to the methods described below and other methods known in the art. Effective therapeutic dosages of the modified peptides may be determined through procedures well known by those in the art and will take into consideration any concerns over potential toxicity of the peptide.

20 The modified peptides can also be administered prophylactically to previously uninfected individuals. This can be advantageous in cases where an individual has been subjected to a high risk of exposure to a virus, as can occur when individual has been in contact with an infected individual where there is a high risk of viral transmission. This can be especially advantageous where there is
25 known cure for the virus, such as the HIV virus. As a example, prophylactic administration of a modified anti-HIV peptide would be advantageous in a situation where a health care worker has been exposed to blood from an HIV-infected individual, or in other situations where an individual engaged in high-risk activities that potentially expose that individual to the HIV virus.

7. **Administration of Modified Anti-Viral and Anti-Fusogenic Peptides**

Generally, the modified peptides will be administered in a physiologically acceptable medium, e.g. deionized water, phosphate buffered saline (PBS), saline, aqueous ethanol or other alcohol, plasma, proteinaceous solutions, mannitol, aqueous glucose, alcohol, vegetable oil, or the like. Other additives which may be included include buffers, where the media are generally buffered at a pH in the range of about 5 to 10, where the buffer will generally range in concentration from about 50 to 250 mM, salt, where the concentration of salt will generally range from about 5 to 500 mM, physiologically acceptable stabilizers, and the like. The compositions may be lyophilized for convenient storage and transport.

The subject modified peptides will for the most part be administered parenterally, such as intravenously (IV), intraarterially (IA), intramuscularly (IM), subcutaneously (SC), or the like. Administration may in appropriate situations be by transfusion. In some instances, where reaction of the functional group is relatively slow, administration may be oral, nasal, rectal, transdermal or aerosol, where the nature of the conjugate allows for transfer to the vascular system. Usually a single injection will be employed although more than one injection may be used, if desired. The modified peptides may be administered by any convenient means, including syringe, trocar, catheter, or the like.

The particular manner of administration will vary depending upon the amount to be administered, whether a single bolus or continuous administration, or the like. Preferably, the administration will be intravascularly, where the site of introduction is not critical to this invention, preferably at a site where there is rapid blood flow, e.g., intravenously, peripheral or central vein. Other routes may find use where the administration is coupled with slow release techniques or a protective matrix. The intent is that the modified peptide be effectively distributed in the blood, so as to be able to react with the blood components. The concentration of the conjugate will vary widely, generally ranging from about 1 pg/ml to 50 mg/ml. The total administered intravascularly will generally be in the range of about 0.1 mg/ml to about 10 mg/ml, more usually about 1 mg/ml to about

5 mg/ml.

By bonding to long-lived components of the blood, such as immunoglobulin, serum albumin, red blood cells and platelets, a number of advantages ensue. The activity of the peptide is extended for days to weeks. Only one administration need be given during this period of time. Greater specificity can be achieved, since the active compound will be primarily bound to large molecules, where it is less likely to be taken up intracellularly to interfere with other physiological processes.

8. Monitoring the Presence of Modified Peptides

The blood of the mammalian host may be monitored for the presence of the modified peptide compound one or more times. By taking a portion or sample of the blood of the host, one may determine whether the peptide has become bound to the long-lived blood components in sufficient amount to be therapeutically active and, thereafter, the level of the peptide compound in the blood. If desired, one may also determine to which of the blood components the peptide is bound. This is particularly important when using non-specific modified peptides. For specific maleimide-modified peptides, it is much simpler to calculate the half life of serum albumin and IgG.

A. Immuno Assays

Another aspect of this invention relates to methods for determining the concentration of the anti-viral peptides and/or analogs, or their derivatives and conjugates in biological samples (such as blood) using antibodies specific for the peptides, peptide analogs or their derivatives and conjugates, and to the use of such antibodies as a treatment for toxicity potentially associated with such peptides, analogs, and/or their derivatives or conjugates. This is advantageous because the increased stability and life of the peptides in vivo in the patient might lead to novel problems during treatment, including increased possibility for toxicity.

The use of anti-therapeutic agent antibodies, either monoclonal or

polyclonal, having specificity for a particular peptide, peptide analog or derivative thereof, can assist in mediating any such problem. The antibody may be generated or derived from a host immunized with the particular peptide, analog or derivative thereof, or with an immunogenic fragment of the agent, or a synthesized immunogen corresponding to an antigenic determinant of the agent. Preferred antibodies will have high specificity and affinity for native, modified and conjugated forms of the peptide, peptide analog or derivative. Such antibodies can also be labeled with enzymes, fluorochromes, or radiolables.

Antibodies specific for modified peptides may be produced by using purified peptides for the induction of peptide-specific antibodies. By induction of antibodies, it is intended not only the stimulation of an immune response by injection into animals, but analogous steps in the production of synthetic antibodies or other specific binding molecules such as screening of recombinant immunoglobulin libraries. Both monoclonal and polyclonal antibodies can be produced by procedures well known in the art.

The anti-peptide antibodies may be used to treat toxicity induced by administration of the modified peptide, analog or derivative thereof, and may be used ex vivo or in vivo. Ex vivo methods would include immuno-dialysis treatment for toxicity employing anti-therapeutic agent antibodies fixed to solid supports. In vivo methods include administration of anti-therapeutic agent antibodies in amounts effective to induce clearance of antibody-agent complexes.

The antibodies may be used to remove the modified peptides, analogs or derivatives thereof, and conjugates thereof, from a patient's blood ex vivo by contacting the blood with the antibodies under sterile conditions. For example, the antibodies can be fixed or otherwise immobilized on a column matrix and the patient's blood can be removed from the patient and passed over the matrix. The modified peptide, peptide analogs, derivatives or conjugates will bind to the antibodies and the blood containing a low concentration of peptide, analog, derivative or conjugate, then may be returned to the patient's circulatory system.

The amount of peptide compound removed can be controlled by adjusting the

pressure and flow rate.

Preferential removal of the peptides, analogs, derivatives and conjugates from the plasma component of a patient's blood can be effected, for example, by the use of a semipermeable membrane, or by otherwise first separating the plasma component from the cellular component by ways known in the art prior to passing the plasma component over a matrix containing the anti-therapeutic antibodies. Alternatively the preferential removal of peptide-conjugated blood cells, including red blood cells, can be effected by collecting and concentrating the blood cells in the patient's blood and contacting those cells with fixed anti-therapeutic antibodies to the exclusion of the serum component of the patient's blood.

The anti-therapeutic antibodies can be administered in vivo, parenterally, to a patient that has received the peptide, analogs, derivatives or conjugates for treatment. The antibodies will bind peptide compounds and conjugates. Once bound the peptide activity will be hindered if not completely blocked thereby reducing the biologically effective concentration of peptide compound in the patient's bloodstream and minimizing harmful side effects. In addition, the bound antibody-peptide complex will facilitate clearance of the peptide compounds and conjugates from the patient's blood stream.

The invention having been fully described can be further appreciated and understood with reference to the following non-limiting examples.

Example 1

Preparation of a Modified DP 178 --Synthesis of

YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWFK(MPA)-NH₂

5 In this example, DP178 (SEQ ID NO:1) is synthesized and modified to include a linker and maleimide group according to the following synthesis scheme. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, DP178 is a potent inhibitor of HIV-1, and inhibits both cell-induced syncytia formation between HIV-1 infected and uninfected cells and infection of uninfected cells by cell-free HIV-1 virus.

10 Solid phase peptide synthesis of the modified peptide on a 100 μmole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Phe-OH, Fmoc-Trp(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ala-OH, Fmoc-Trp(Boc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Met-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH; Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-His(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Tyr(tBu)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). At the end of the

synthesis. The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $\text{Pd}(\text{PPh}_3)_4$ dissolved in 5 mL of CHCl_3 :NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

DP-178 C

Fmoc-Rink Amide MBHA Resin

Step 1

SPPS

Boc-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Lys(Aloc)-PS

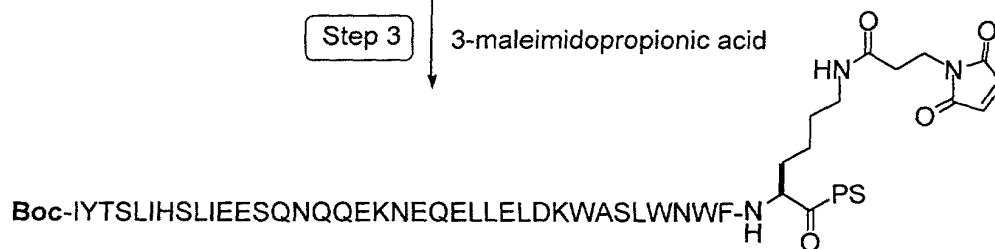
Step 2

$\text{Pd}(\text{PPh}_3)_4/\text{NMM}/\text{HOAc}/\text{CHCl}_3$

Boc-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Lys-PS

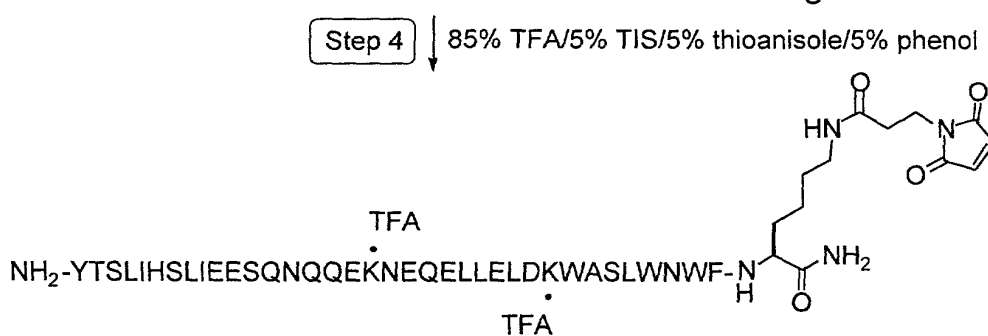
Step 3

3-maleimidopropionic acid



Step 4

85% TFA/5% TIS/5% thioanisole/5% phenol



Example 2

5 Preparation of a Modified DP107--Synthesis of
NNLLRAIEAQQHLLQLTVWQIKQLQARILAVEERYLKDQK(MPA)NH₂

In this example, DP107 (SEQ ID NO:2) is synthesized and modified to

include a linker and maleimide group according to the following synthesis scheme. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, DP107 exhibits potent antiviral activity against HIV.

5 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Leu-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Leu-OH, Fmoc-Ile-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ala-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Val-OH, Fmoc-Thr(tBu)-OH, Fmoc-Leu-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-His(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Asn(Trt)-OH, They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N'*, *N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and

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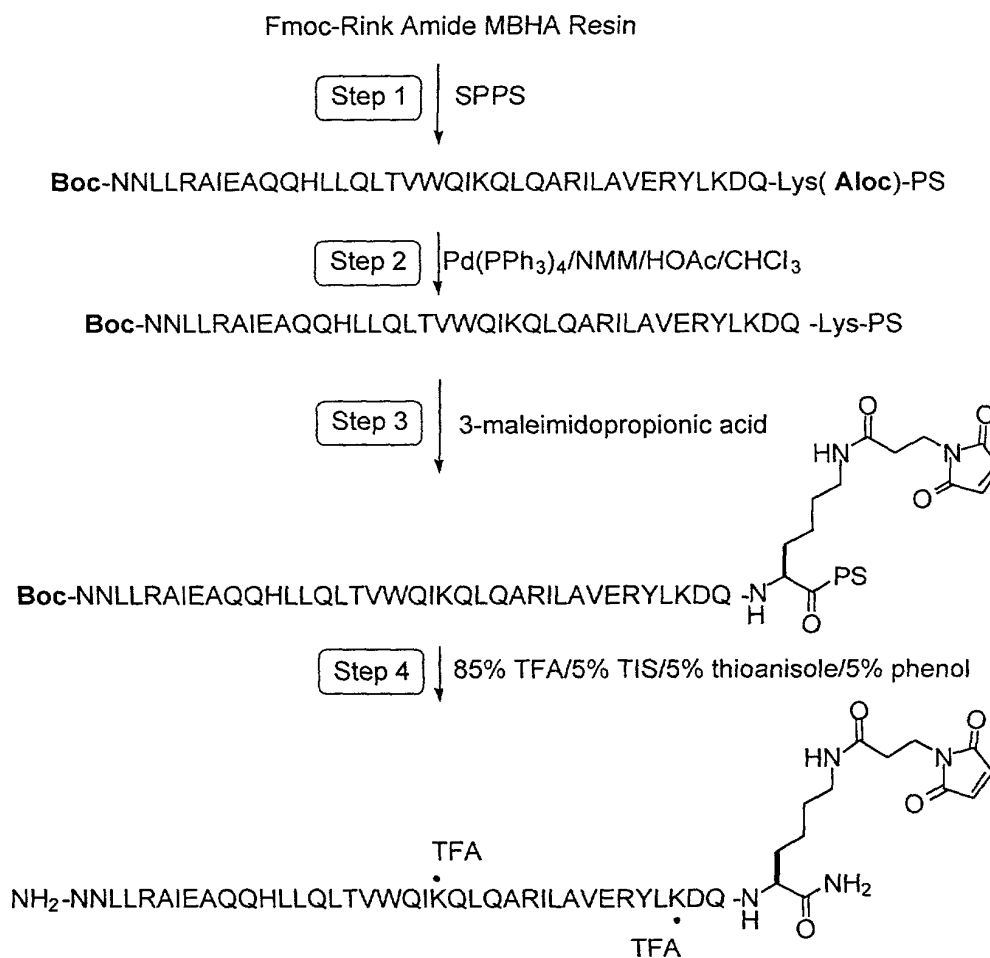
20 Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). At the end of the synthesis. The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $\text{Pd}(\text{PPh}_3)_4$ dissolved in 5 mL of CHCl_3 :NMM:HOAc

25 (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using

30 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by

- 5 dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

DP-107 C



Example 3

Preparation of a Modified anti-RSV peptide (C terminal)

In this example, the peptide

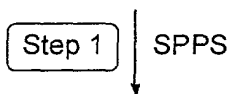
5 VITIELSNIKENKCNGAKVKLIKQELDKYKNAV (SEQ ID NO:16) is modified
to include a linker and maleimide group according to the synthesis scheme set
forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, the native
sequence (SEQ ID NO.) inhibits viral infection of respiratory syncytial virus
(RSV), including inhibiting fusion and syncytia formation between RSV-infected
10 and uninfected Hep-2 cells.

Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale
is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer
and Fmoc protected Rink Amide MBHA. The following protected amino acids are
15 sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Val-OH, Fmoc-Ala-OH,
Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-
OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-
OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH,
Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-
20 Asn(Trt)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH,
Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH,
Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-
Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH. They are dissolved in *N,N*-
dimethylformamide (DMF) and, according to the sequence, activated using *O*-
25 benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU)
and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is
achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide
(DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group
is performed manually and accomplished by treating the resin with a solution of 3
30 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step

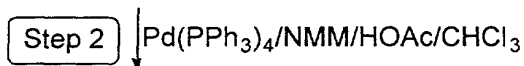
2). The resin is then washed with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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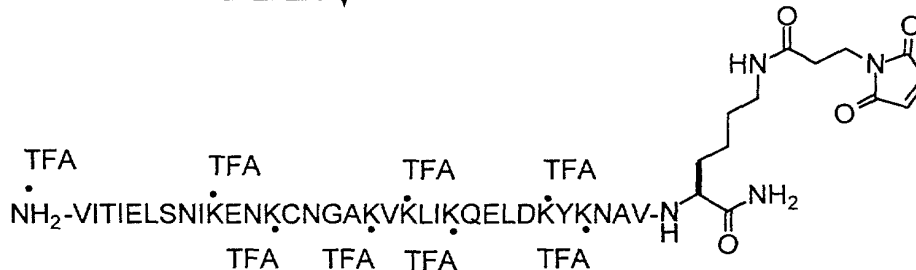
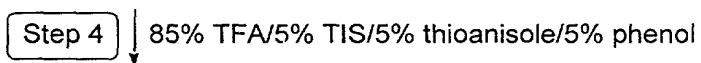
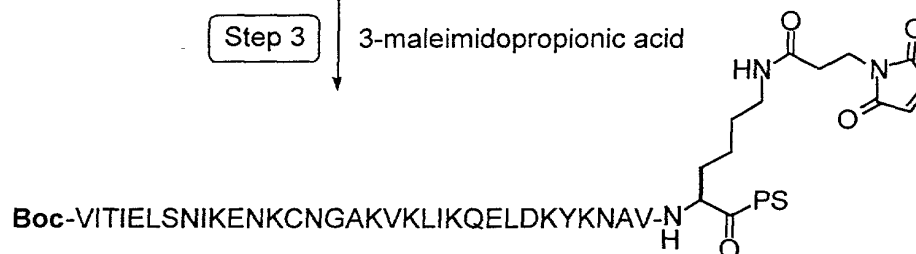
Fmoc-Rink Amide MBHA Resin



Boc-VITIELSNIKENKCNGAKVKLIKQELDKYKNAV-Lys(Aloc)-PS



Boc-VITIELSNIKENKCNGAKVKLIKQELDKYKNAV-Lys -PS



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Example 4

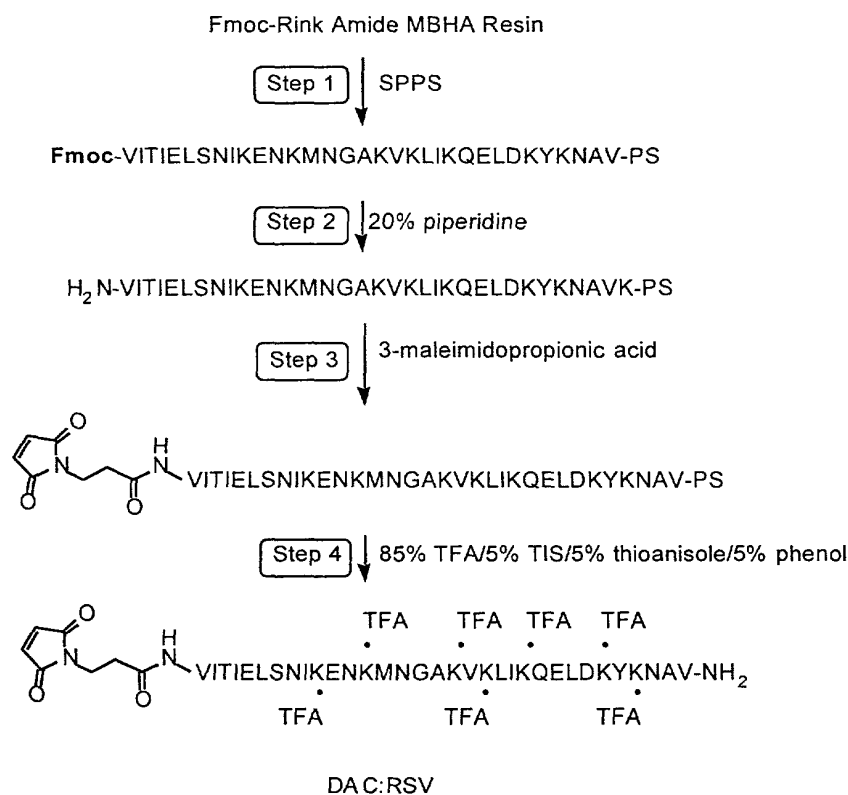
Preparation of a Modified anti-RSV peptide (T-N terminal)

In this example, the peptide

5 VITIELSNIKENKCNCAKVKLIKQELDKYKNAV (SEQ ID NO:17), which
corresponds to the peptide of SEQ ID NO:16 but where a Cysteine (C) has been
substituted for the Methionine (M), residue is synthesized and modified to
include a linker and maleimide group according to the synthesis scheme set forth
below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, the native sequence
10 (SEQ ID NO:16) inhibits viral infection of respiratory syncytial virus (RSV),
including inhibiting fusion and syncytia formation between RSV-infected and
uninfected Hep-2 cells.

15 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale
is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer
and Fmoc protected Rink Amide MBHA. The following protected amino acids are
sequentially added to resin: Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH,
Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-
20 OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-
OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-
Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, **Fmoc-Met-**
OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-
Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-
25 OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH,
Fmoc-Val-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and,
according to the sequence, activated using O-benzotriazol-1-yl-*N,N,N'*-
tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine
(DIEA). Removal of the Fmoc protecting group is achieved using a solution of
30 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).
The synthesis is then re-automated for the addition of the 3-maleimidopropionic

acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

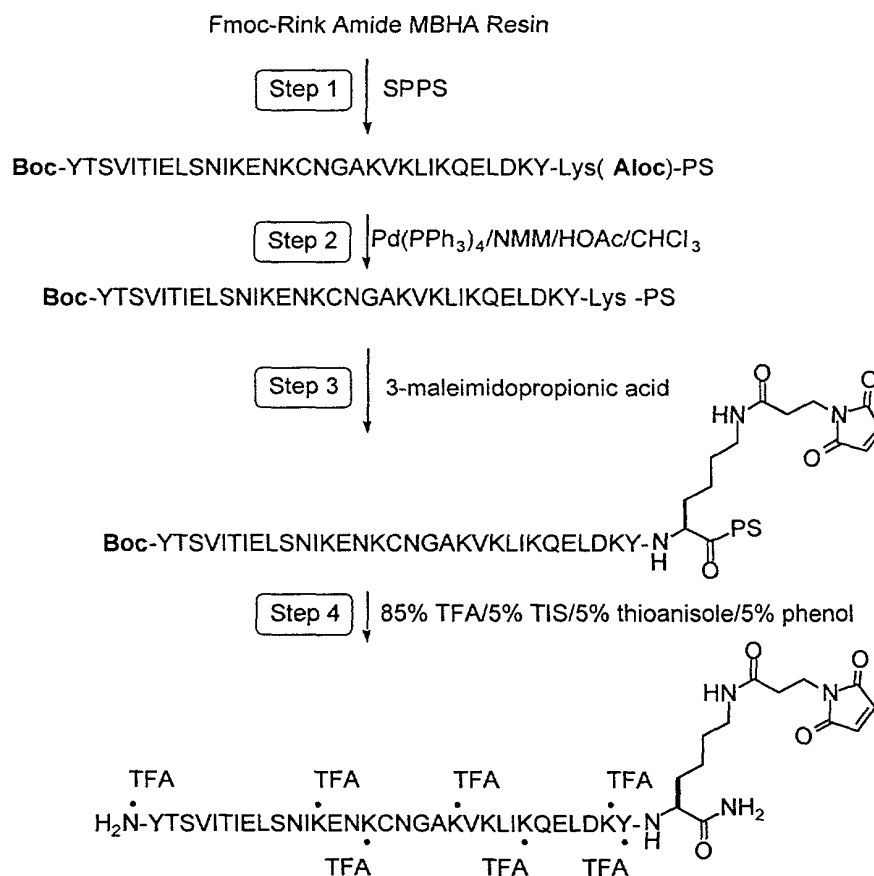


Example 5
Preparation of a Modified anti-RSV peptide

In this example, the peptide SEQ ID NO:14 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:14 inhibits viral infection of respiratory syncytial virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Tyr(tBu)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated

for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



Example 6 (T-143)

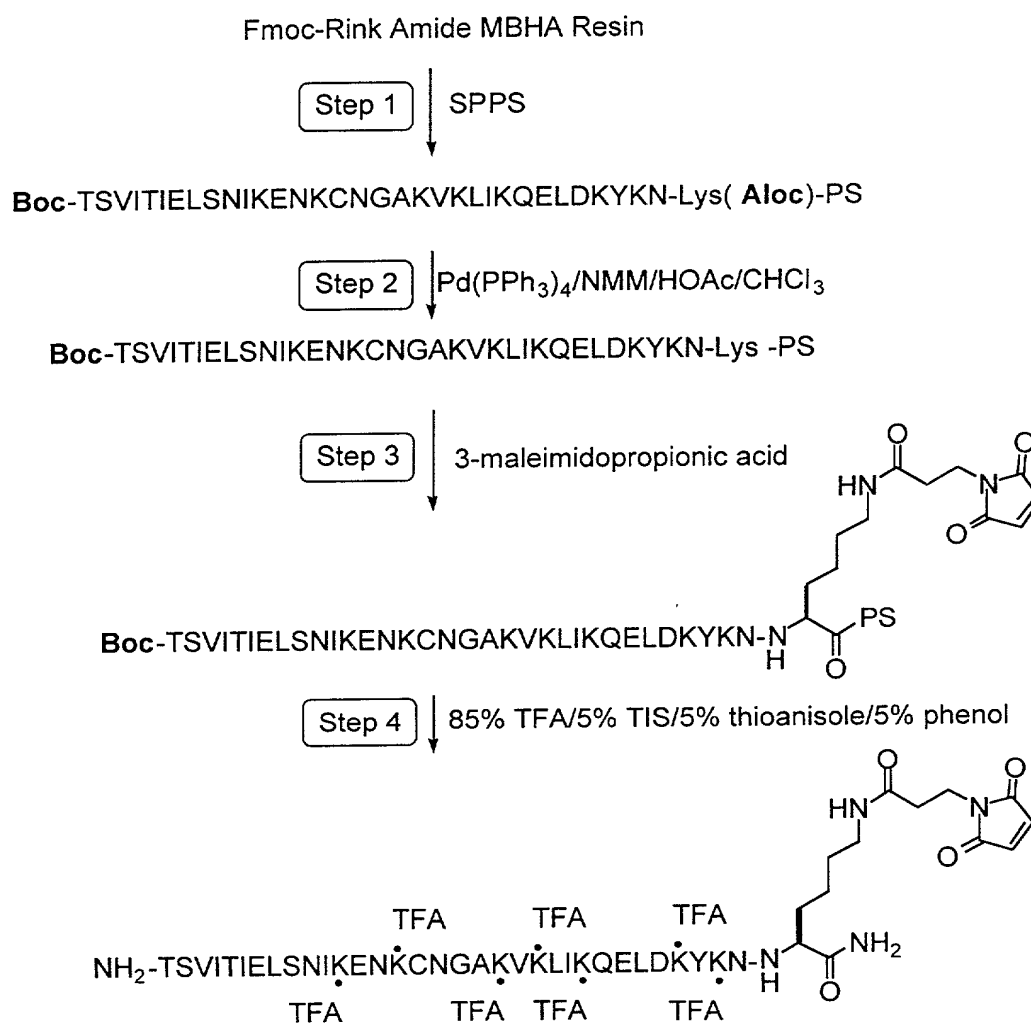
Preparation of a Modified anti-RSV peptide

5 In this example, the peptide SEQ ID NO:15 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:15 inhibits viral infection of respiratory syncytial virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

10 Solid phase peptide synthesis of the modified peptide analog on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-OH, 15 Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, 20 Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyluronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting 25 group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $\text{Pd}(\text{PPh}_3)_4$ dissolved in 5 mL of CHCl_3 :NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with 30 CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6

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x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ₂₁₄ and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.



Example 7

5 Preparation of a Modified anti-RSV peptide (C Terminal)

In this example, the peptide SEQ ID NO:17), which corresponds to SEQ ID NO:16 with a cysteine (C) substituted for the Methionine (M), is synthesized and modified to include a linker and maleimide group according to the synthesis

10 scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, the native sequence SEQ ID NO:16. inhibits viral infection of respiratory syncytial

virus (RSV), including inhibiting fusion and syncytia formation between RSV-infected and uninfected Hep-2 cells.

5 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, **Fmoc-Met-OH**, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Val-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N'*, *N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $\text{Pd}(\text{PPh}_3)_4$ dissolved in 5 mL of CHCl_3 :NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045%

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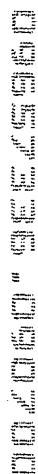
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TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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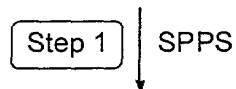
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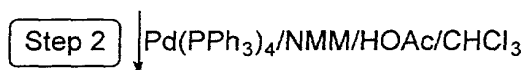
Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Leu-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Val-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Val-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Ile-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide

(i.e., DAC) in >95% purity, as determined by RP-HPLC.

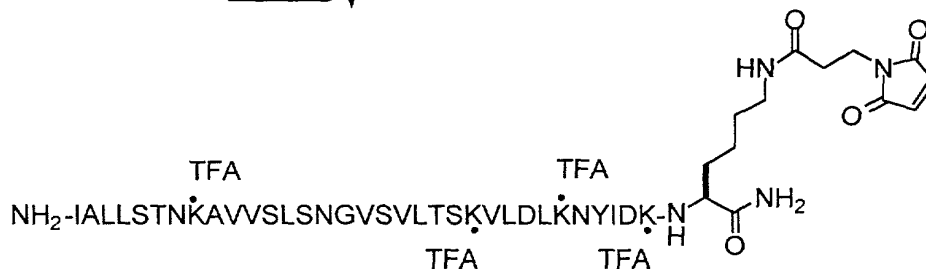
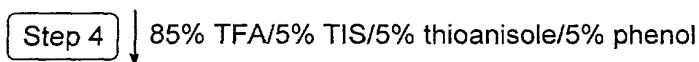
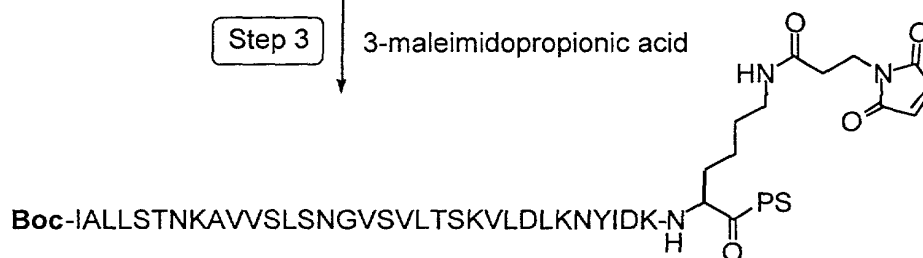
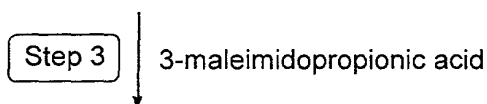
Fmoc-Rink Amide MBHA Resin



Boc-IALLSTNKAVVSLNSGVSVLTskVLDLkNYIDK-Lys(Alloc)-PS



Boc-IALLSTNKAVVSLNSGVSVLTskVLDLkNYIDK-Lys-PS



Example 9 (T-173)

5 Preparation of a Modified anti-HPIV peptide

In this example, the peptide SEQ ID NO:52. is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:52 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and

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uninfected CV-1W cells.

Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ala-OH, Fmoc-Gln(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Val-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIÉA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex

Fmoc-Rink Amide MBHA Resin

SPPS

Step 2

$\text{Pd}(\text{PPh}_3)_4/\text{NMM}/\text{HOAc}/\text{CHCl}_3$

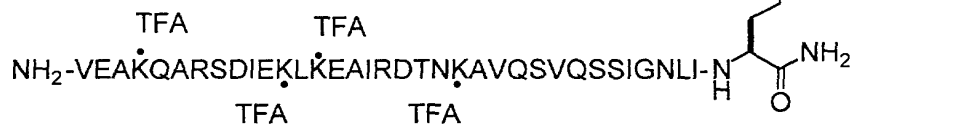
Step 3

3-maleimidopropionic acid



Step 4

85% TFA/5% TIS/5% thioanisole/5% phenol



Example 10

Preparation of a Modified anti-HPIV peptide

In this example, the peptide SEQ ID NO:58 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:58 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Val-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Val-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $\text{Pd}(\text{PPh}_3)_4$ dissolved in 5 mL of CHCl_3 :NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with

- CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The
- 5 peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex
- 10 Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

Fmoc-Rink Amide MBHA Resin

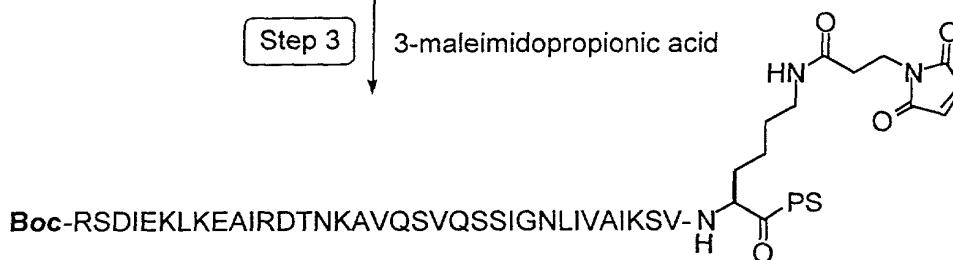
Step 1 ↓ SPPS

Boc-RSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSV-Lys(Aloc)-PS

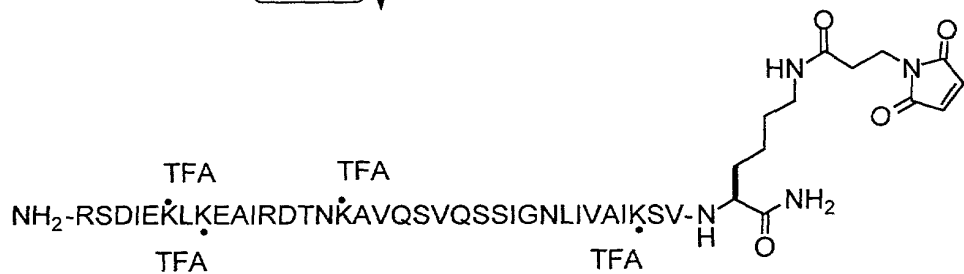
Step 2 ↓ Pd(PPh₃)₄/NMM/HOAc/CHCl₃

Boc-RSDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSV-Lys -PS

Step 3 ↓ 3-maleimidopropionic acid



Step 4 ↓ 85% TFA/5% TIS/5% thioanisole/5% phenol



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Example 11

Preparation of a Modified anti-HPIV peptide

5 In this example, the peptide SEQ ID NO:35 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:35 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Leu-OH, Fmoc-
15 Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-
20 OH, Fmoc-Leu-OH, Fmoc-Ile-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Val-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyluronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3
30 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step

2). The resin is then washed with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3
5 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5
10 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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Fmoc-Rink Amide MBHA Resin

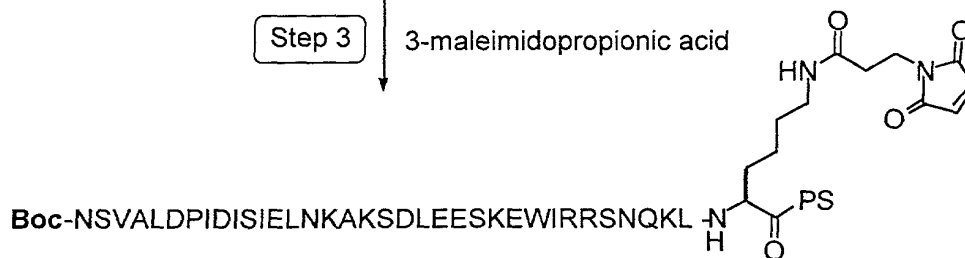
Step 1 ↓ SPPS

Boc-NSVALDPIDISIELNKAKSDLEESKEWIRRSNQKL -Lys(**Aloc**)-PS

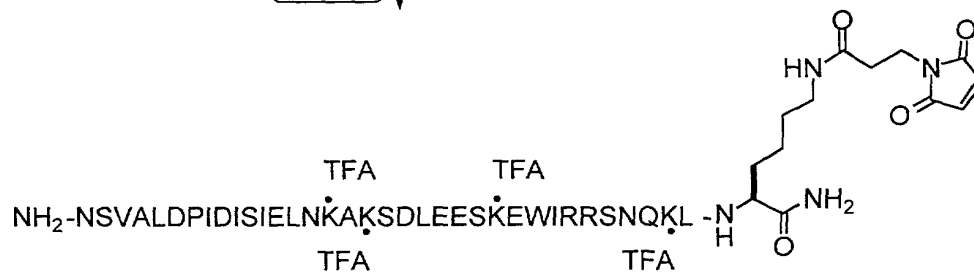
Step 2 ↓ Pd(PPh₃)₄/NMM/HOAc/CHCl₃

Boc-NSVALDPIDISIELNKAKSDLEESKEWIRRSNQKL -Lys-PS

Step 3 ↓ 3-maleimidopropionic acid



Step 4 ↓ 85% TFA/5% TIS/5% thioanisole/5% phenol



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Example 12

Preparation of a Modified anti-HPIV peptide

In this example, the peptide SEQ ID NO:38 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO: 38 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and uninfected CV-1W cells.

Solid phase peptide synthesis of the modified peptide analog on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ala-OH BOC-Lys(Aloc)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of $\text{Pd}(\text{PPh}_3)_4$ dissolved in 5 mL of CHCl_3 :NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed

with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

Fmoc-Rink Amide MBHA Resin

Step 1

SPPS

Boc-Lys(Aloc)-ALDPIDISIELNKA[•]SDLEESKEWIRRSNQKLD[•]SI-PS

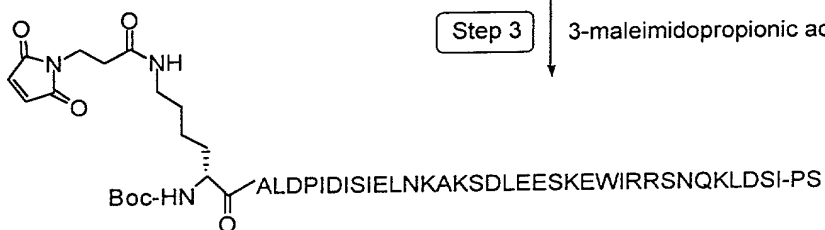
Step 2

$\text{Pd(PPh}_3)_4/\text{NMM/HOAc/CHCl}_3$

Boc-Lys-ALDPIDISIELNKA[•]SDLEESKEWIRRSNQKLD[•]SI-PS

Step 3

3-maleimidopropionic acid



Step 4

85% TFA/5% TIS/5% thioanisole/5% phenol



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Example 13

Preparation of a Modified anti-HPIV peptide

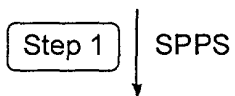
5 In this example, the peptide SEQ ID NO:39 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:39 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and
10 uninfected CV-1W cells.

 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are
15 sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-
20 OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the
25 sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the
30 resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of

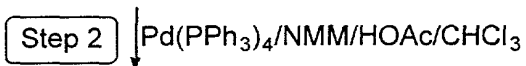
CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3
5 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and
10 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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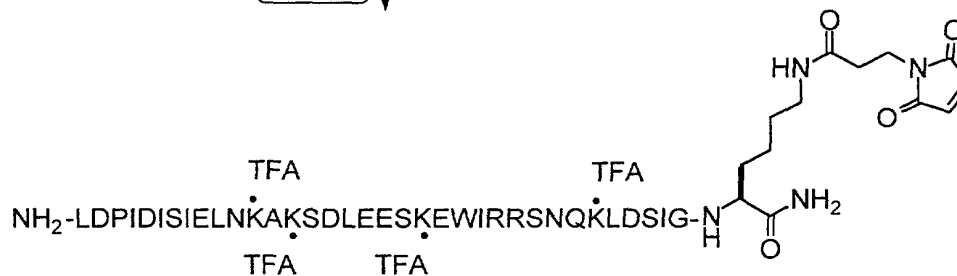
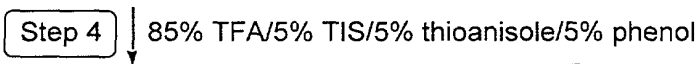
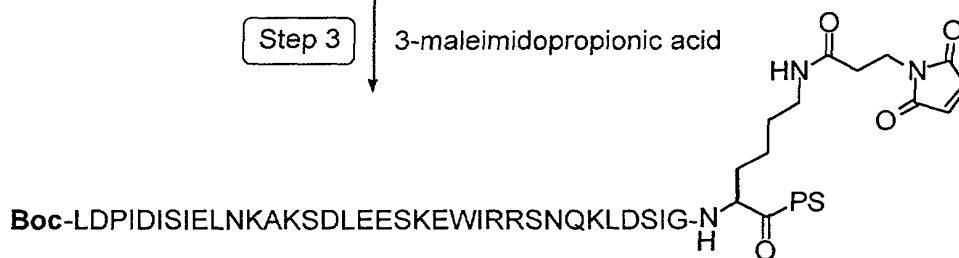
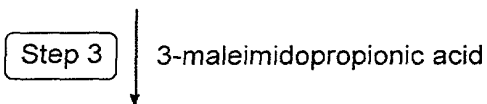
Fmoc-Rink Amide MBHA Resin



Boc-LDPIDISIELNKASDLEESKEWIRRSNQKLDSIG-Lys(**Aloc**)-PS



Boc-LDPIDISIELNKASDLEESKEWIRRSNQKLDSIG-Lys -PS



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Example 14

Preparation of a Modified anti-HPIV peptide

5 In this example, the peptide SEQ ID NO:40 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO. inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and
10 uninfected CV-1W cells.

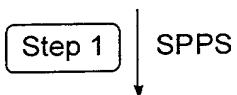
 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected
15 amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH,
20 Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Asp(tBu)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and,
25 according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and
30 accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed

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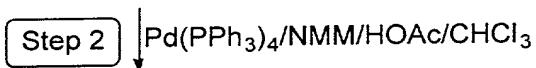
- with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The
- 5 peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex
- 10 Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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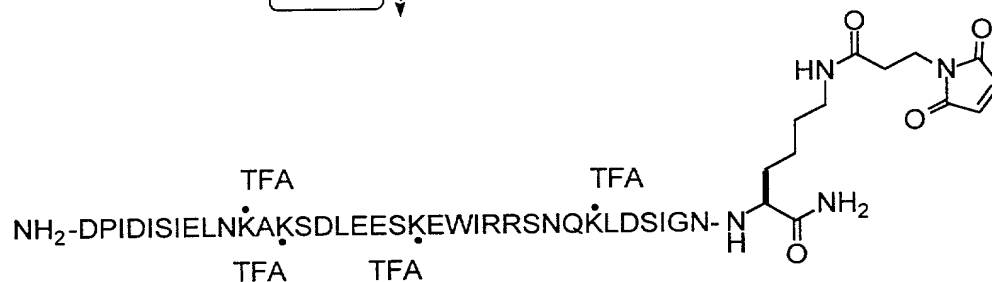
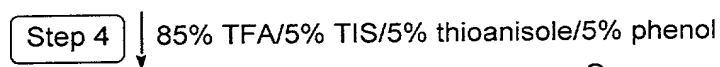
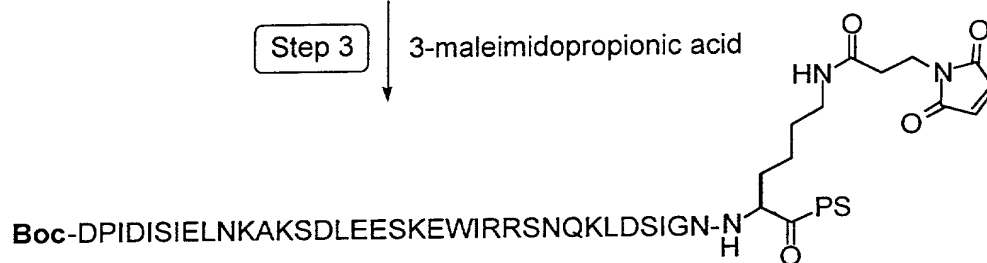
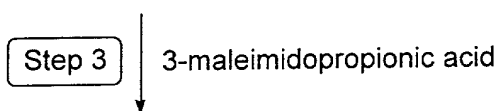
Fmoc-Rink Amide MBHA Resin



Boc-DPIDISIELNKASDLEESKEWIRRSNQKLDSIGN-Lys(**Aloc**)-PS



Boc-DPIDISIELNKASDLEESKEWIRRSNQKLDSIGN-Lys -PS



Example 15

Preparation of a Modified anti-HPIV peptide

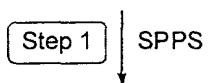
5 In this example, the peptide SEQ ID NO:41 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:41 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and
10 uninfected CV-1W cells.

 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected
15 amino acids are sequentially added to resin: Fmoc-Trp(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH,
20 Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and,
25 according to the sequence, activated using O-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1).The selective deprotection of the Lys (Aloc) group is performed manually and
30 accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed

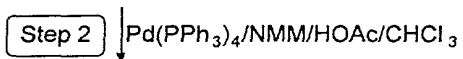
with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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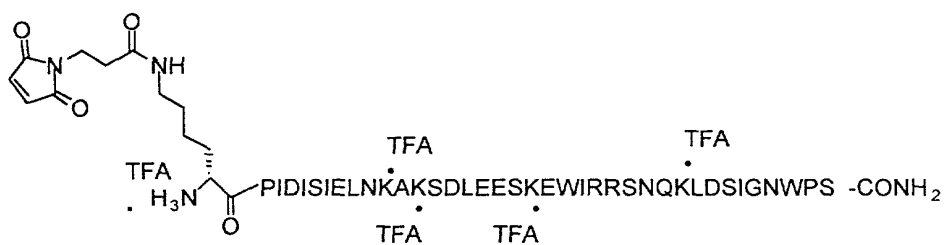
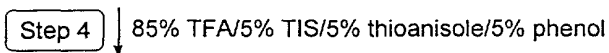
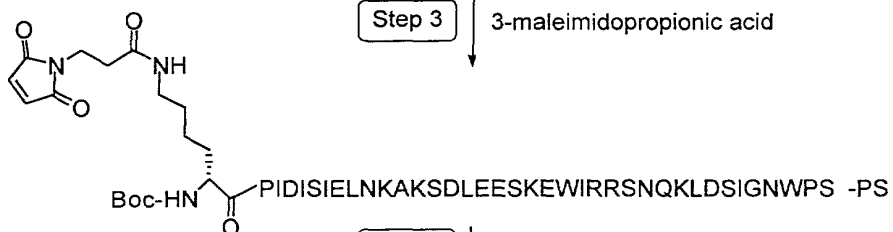
Fmoc-Rink Amide MBHA Resin



Boc-Lys(Aloc)-PIDISIELNKA KSDLEESKEWIRRSNQKLDSIGNW-PS



Boc-Lys-PIDISIELNKA KSDLEESKEWIRRSNQKLDSIGNWPS



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Example 16

Preparation of a Modified anti-HPIV peptide

5 In this example, the peptide SEQ ID NO:42 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:42 inhibits viral infection of human parainfluenza virus 3 (HPIV3), including inhibiting fusion and syncytia formation between HPIV3-infected Hep2 cells and
10 uninfected CV-1W cells.

 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are
15 sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-His(Boc)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gly-OH, Fmoc-Asn(Trt)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Ile-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Ile-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and,
20 according to the sequence, activated using O-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and
25 accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in
30

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5 mL of CHCl_3 :NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC

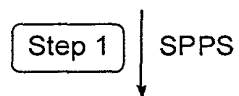
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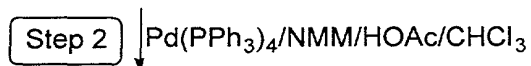
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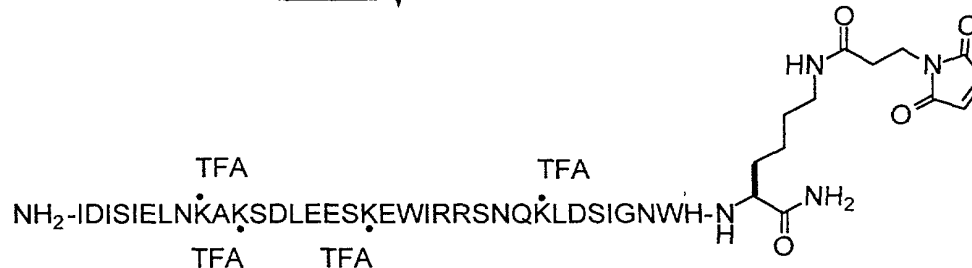
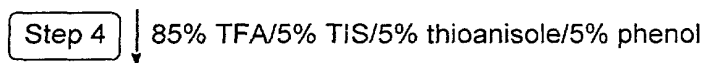
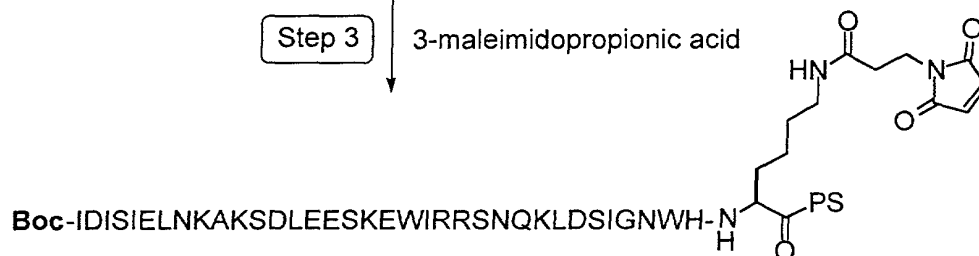
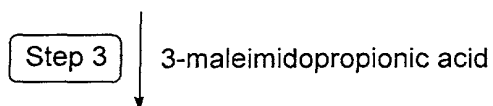
Fmoc-Rink Amide MBHA Resin



Boc-IDISIELNKAKSDLEESKEWIRRSNQKLDSIGNWH-Lys(**Aloc**)-PS



Boc-IDISIELNKAKSDLEESKEWIRRSNQKLDSIGNWH-Lys-PS



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Example 17

Preparation of a Modified anti-MeV peptide

5 In this example, the peptide SEQ ID NO:77. is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:77 inhibits viral infection of measles virus (MeV), including inhibiting fusion and syncytia formation between MeV-infected and uninfected Vero cells.

10

 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, Fmoc-Arg(Pbf)-OH, Fmoc-His(Boc)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and

15

20

25

30

5

10

Fmoc-Rink Amide MBHA Resin

Step 1

SPPS

Boc-HRIDLGPPISLERLDVGTNLGNIAKLEAKELLE-Lys(Alloc)-PS

Step 2

$\text{Pd(PPh}_3)_4$ /NMM/HOAc/ CHCl_3

Boc-HRIDLGPPISLERLDVGTNLGNIAKLEAKELLE-Lys-PS

Step 3

3-maleimidopropionic acid

Boc-HRIDLGPPISLERLDVGTNLGNIAKLEAKELLE-NH-CH(CH₂)₄-NH-CO-CH₂-CH₂-N-maleimide-PS

Step 4

85% TFA/5% TIS/5% thioanisole/5% phenol

NH₂-HRIDLGPPISLERLDVGTNLGNIAKLEAKELLE-NH-CH(CH₂)₄-NH-CO-CH₂-CH₂-N-maleimide

TFA
TFA

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Example 18

Preparation of a Modified anti-MeV peptide

5 In this example, the peptide SEQ ID NO:79 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:79 inhibits viral infection of measles virus (MeV), including inhibiting fusion and syncytia formation between MeV-infected and uninfected Vero cells.

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ile-OH, They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N'*, *N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and

25 Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2).

30 The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL),

DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC

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Fmoc-Rink Amide MBHA Resin

Step 1

SPPS

Boc-IDLGPPISLERLDVGNTNLGNAIAKLEAKELLESS-Lys(Aloc)-PS

Step 2

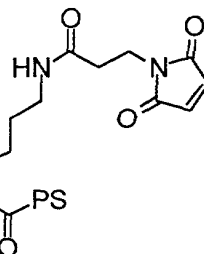
$\text{Pd(PPh}_3)_4/\text{NMM/HOAc/CHCl}_3$

Boc-IDLGPPISLERLDVGNTNLGNAIAKLEAKELLESS-Lys -PS

Step 3

3-maleimidopropionic acid

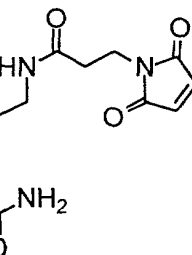
Boc-IDLGPPISLERLDVGNTNLGNAIAKLEAKELLESS-NH



Step 4

85% TFA/5% TIS/5% thioanisole/5% phenol

NH₂-IDLGPPISLERLDVGNTNLGNAIAKLEAKELLESS-NH



Example 19

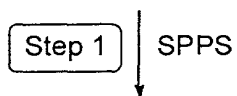
Preparation of a Modified anti-MeV peptide

- 5 In this example, the peptide SEQ ID NO:81 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO: 79 inhibits viral infection of measles virus (MeV), including inhibiting fusion and syncytia formation between MeV-infected and uninfected Vero cells.
- 10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-
- 15 Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Val-OH,
- 20 Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH, Fmoc-Pro-OH, Fmoc-Gly-OH, Fmoc-Leu-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using O-benzotriazol-1-yl-*N,N,N'*, *N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and
- 25 Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2).
- 30 The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the

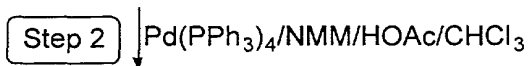
addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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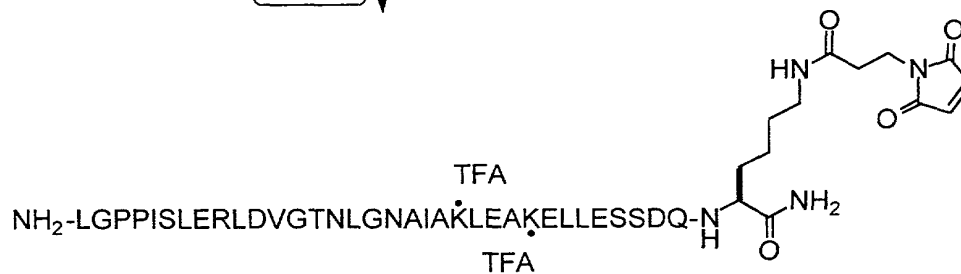
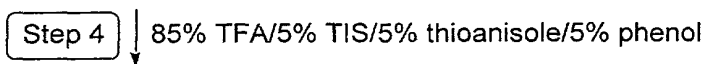
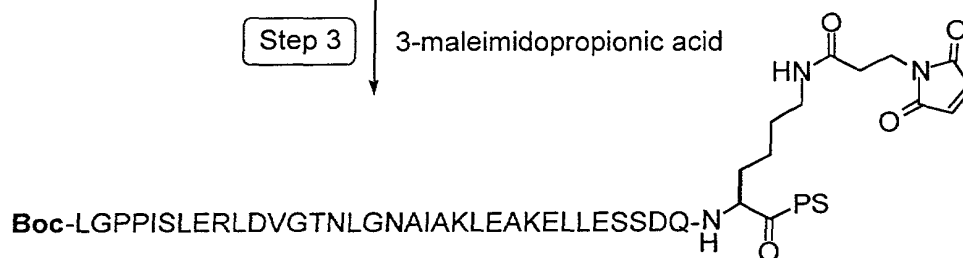
Fmoc-Rink Amide MBHA Resin



Boc-LGPPISLERLDVGTNLGNAIAKLEAKELLESSDQ-Lys(**Aloc**)-PS



Boc-LGPPISLERLDVGTNLGNAIAKLEAKELLESSDQ-Lys-PS



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Example 20

Preparation of a Modified anti-MeV peptide

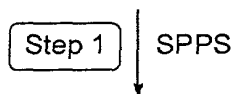
5 In this example, the peptide SEQ ID NO:84 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:84 inhibits viral infection of measles virus (MeV), including inhibiting fusion and syncytia formation between MeV-infected and uninfected Vero cells.

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Leu-
15 OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ala-OH, Fmoc-Ile-OH, Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Leu-OH, Fmoc-Asn(Trt)-OH, Fmoc-
20 Thr(tBu)-OH, Fmoc-Gly-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Leu-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Ser(tBu)-OH, Fmoc-Ile-OH, Fmoc-Pro-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyluronium hexafluorophosphate (HBTU) and
25 Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2).
30 The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the

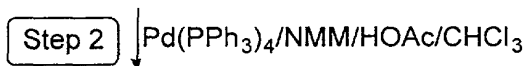
addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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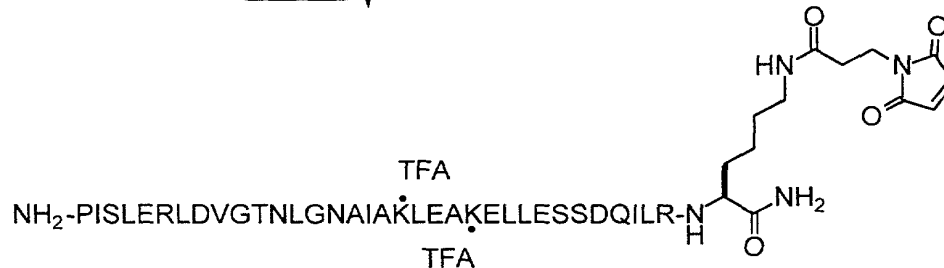
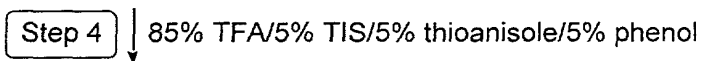
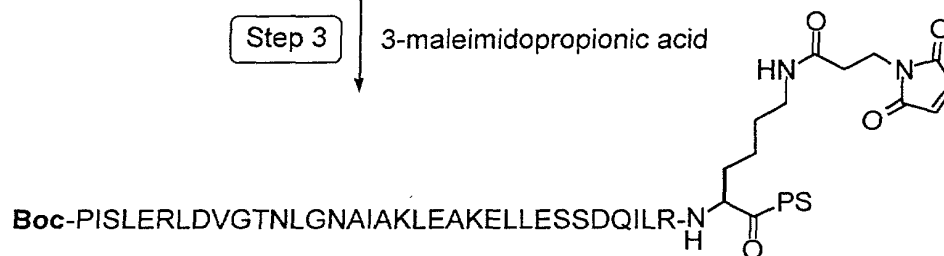
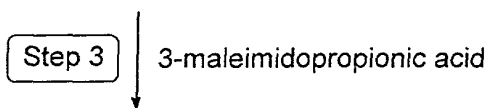
Fmoc-Rink Amide MBHA Resin



Boc-PISLERLDVGTNLGNIAIKLEAKELLESSDQILR-Lys(**Aloc**)-PS



Boc-PISLERLDVGTNLGNIAIKLEAKELLESSDQILR-Lys -PS



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Example 21

Preparation of a Modified anti-SIV peptide

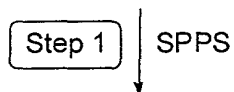
5 In this example, the peptide SEQ ID NO:64 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:64. exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Trp(Boc)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed

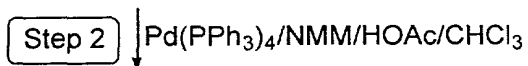
- with CHCl_3 (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The
- 5 peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et_2O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H_2O (A) and 0.045% TFA in CH_3CN (B)) over 180 min at 9.5 mL/min using a Phenomenex
- 10 Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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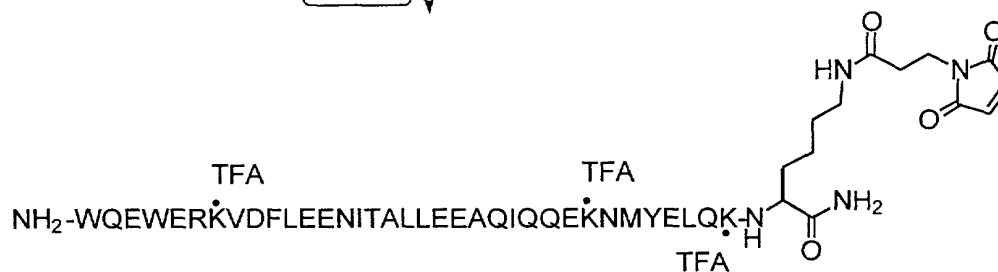
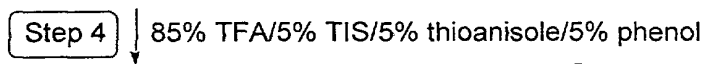
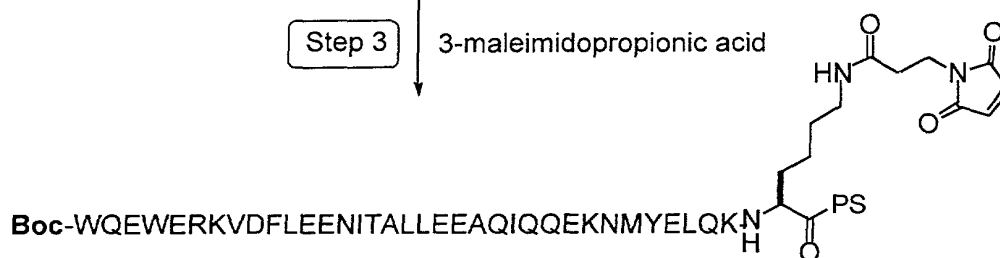
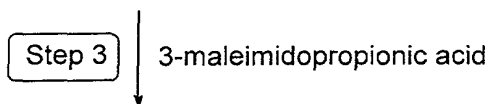
Fmoc-Rink Amide MBHA Resin



Boc-WQEWERKVDLFLEENITALLEEAIQQEKNMYELQK-Lys(Aloc)-PS



Boc-WQEWERKVDLFLEENITALLEEAIQQEKNMYELQK-Lys-PS



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Example 22

Preparation of a Modified anti-SIV peptide

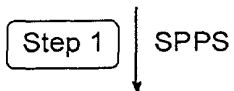
5 In this example, the peptide SEQ ID NO:65 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:65 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Leu-OH, Fmoc-
15 Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH,
20 Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of
25 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in
30 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and

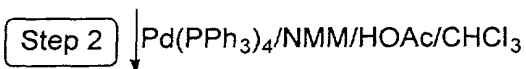
DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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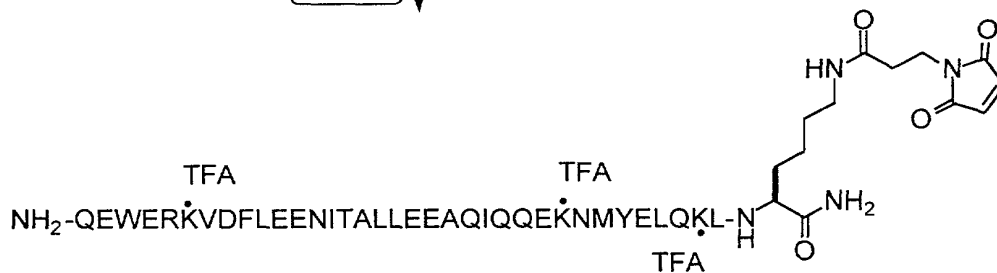
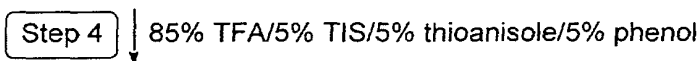
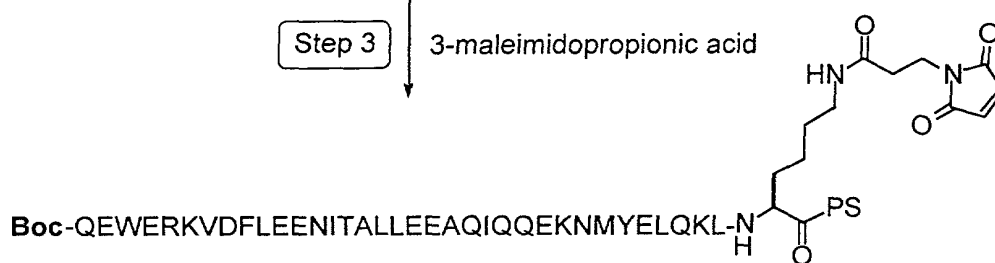
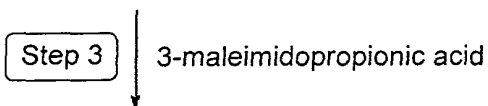
Fmoc-Rink Amide MBHA Resin



Boc-QEWERKVDLFLEENITALLEEAIQQEKNMYELQKL-Lys(Aloc)-PS



Boc-QEWERKVDLFLEENITALLEEAIQQEKNMYELQKL-Lys -PS



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Example 23

Preparation of a Modified anti-SIV peptide

5 In this example, the peptide SEQ ID NO:66 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:66 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of

20 Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Glu(tBu)-OH Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of

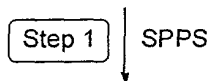
25 hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of

30 CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6

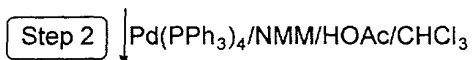
x 5 mL). The synthesis is then re-automated for the addition of the 3-
maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3
times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The
peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5%
5 phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is
purified by preparative reversed phased HPLC using a Varian (Rainin) preparative
binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and
0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex
Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian
10 Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide
(i.e., DAC) in >95% purity, as determined by RP-HPLC.

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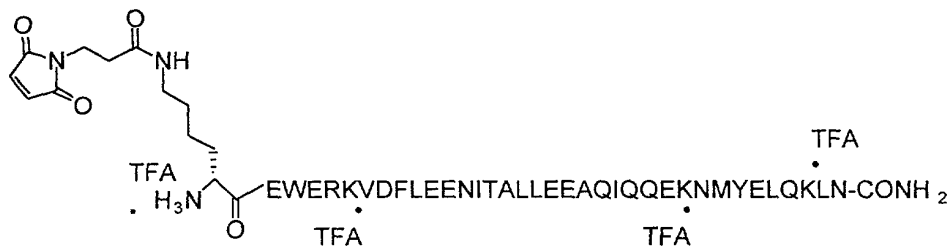
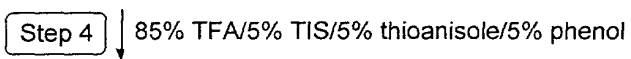
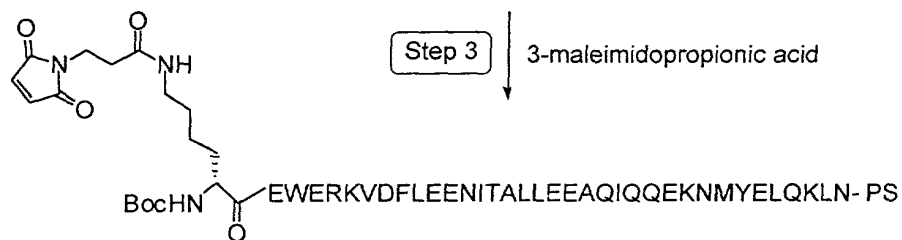
Fmoc-Rink Amide MBHA Resin



Boc-Lys(Aloc)-EWERKVDFLEENITALLEEAIQQEKNMYELQKLN-PS



Boc-Lys-EWERKVDFLEENITALLEEAIQQEKNMYELQKLN-PS



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Example 24

Preparation of a Modified anti-SIV peptide

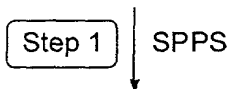
In this example, the peptide SEQ ID NO:67 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:67 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Trp(Boc)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and

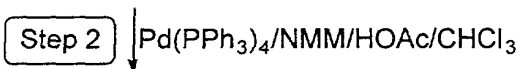
DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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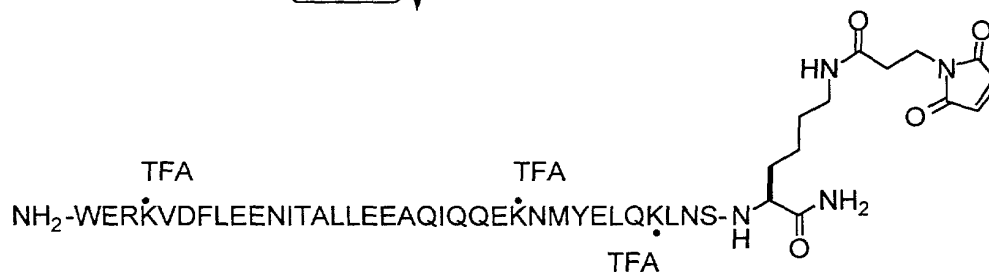
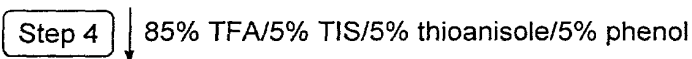
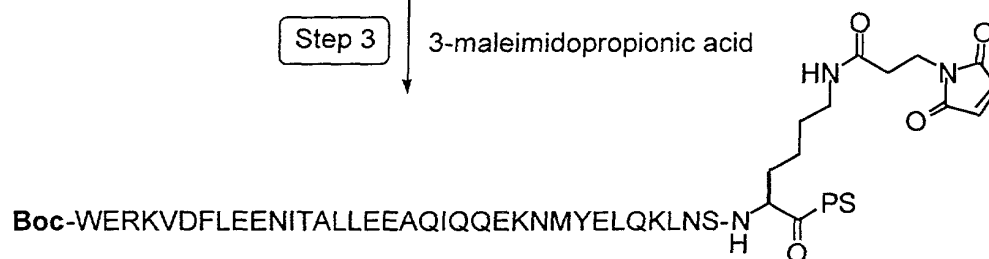
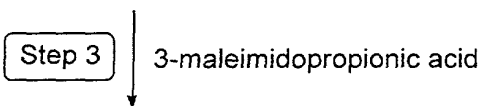
Fmoc-Rink Amide MBHA Resin



Boc-WERKVDLFLEENITALLEEAQIQQEKNMYELQKLNS-Lys(Aloc)-PS



Boc-WERKVDLFLEENITALLEEAQIQQEKNMYELQKLNS-Lys -PS



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Example 25

Preparation of a Modified anti-SIV peptide

5 In this example, the peptide SEQ ID NO:68 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:68 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Glu(tBu)-OH Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and

DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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Fmoc-Rink Amide MBHA Resin

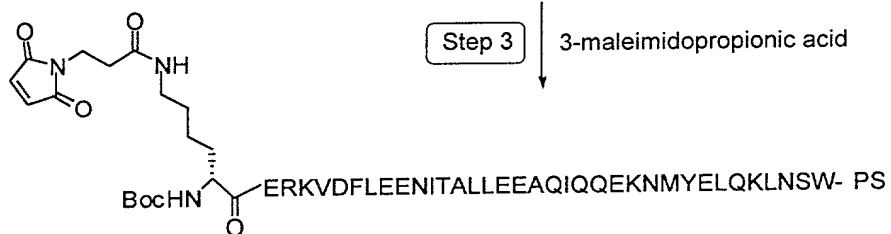
Step 1 ↓ SPPS

Boc-Lys(Aloc)-ERKVDLFLEENITALLEEAIQQEKNMYELQKLNSW -PS

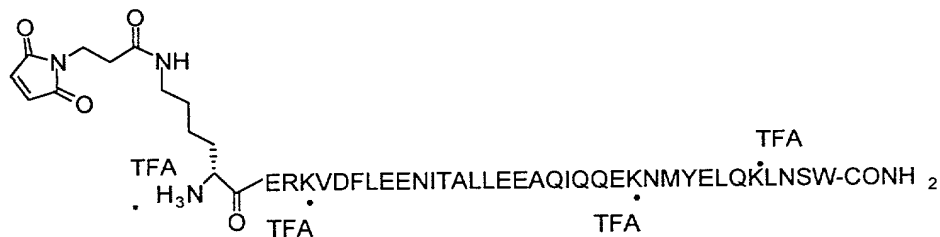
Step 2 ↓ Pd(PPh₃)₄/NMM/HOAc/CHCl₃

Boc-Lys-ERKVDLFLEENITALLEEAIQQEKNMYELQKLNSW- PS

Step 3 ↓ 3-maleimidopropionic acid



Step 4 ↓ 85% TFA/5% TIS/5% thioanisole/5% phenol



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Example 26

Preparation of a Modified anti-SIV peptide

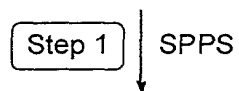
In this example, the peptide SEQ ID NO:69 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:69 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Fmoc-Arg(Pbf)-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and

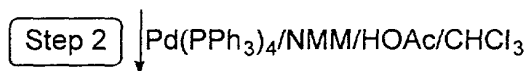
DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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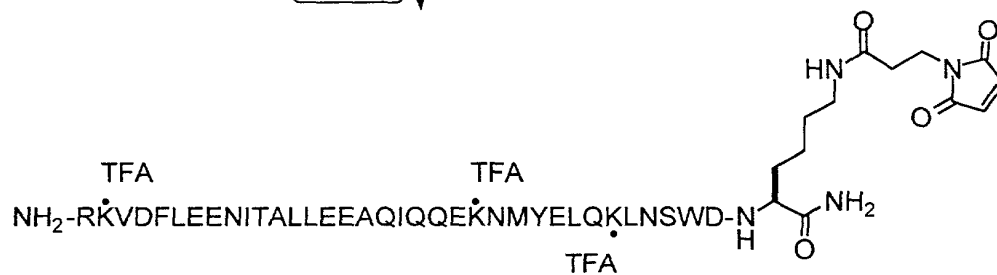
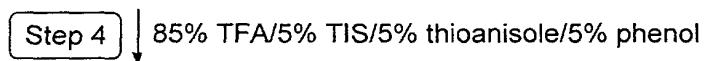
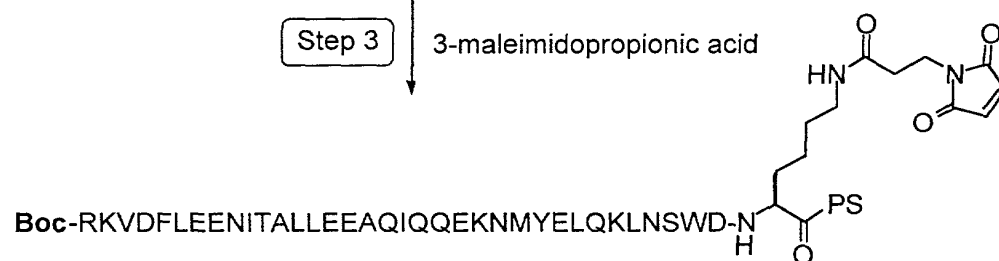
Fmoc-Rink Amide MBHA Resin



Boc-RKVDFLEENITALLEEAIQQEKNMYELQKLNSWD-Lys(Alloc)-PS



Boc-RKVDFLEENITALLEEAIQQEKNMYELQKLNSWD-Lys-PS



Example 27

Preparation of a Modified anti-SIV peptide

5 In this example, the peptide SEQ ID NO:70. is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:70 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

10

15 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, 20 Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH, Fmoc-Lys(Boc)-OH, Boc-Lys(Aloc)-OH,. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, 25 activated using O-benzotriazol-1-yl-*N, N, N', N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the 30 resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of

CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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Fmoc-Rink Amide MBHA Resin

Step 1

SPPS

Boc-Lys(Aloc)-KVDFLEENITALLEEAIQQEKNMYELQKLNSWDV-Lys(Aloc)-PS

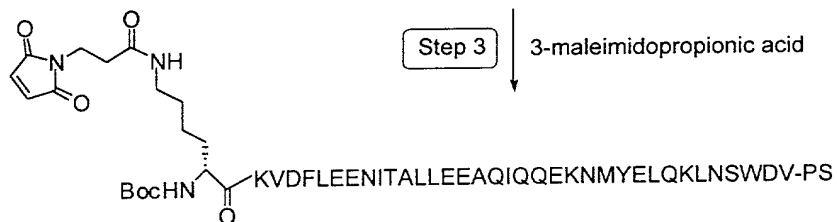
Step 2

$\text{Pd}(\text{PPh}_3)_4/\text{NMM}/\text{HOAc}/\text{CHCl}_3$

Boc-Lys-KVDFLEENITALLEEAIQQEKNMYELQKLNSWDV-PS

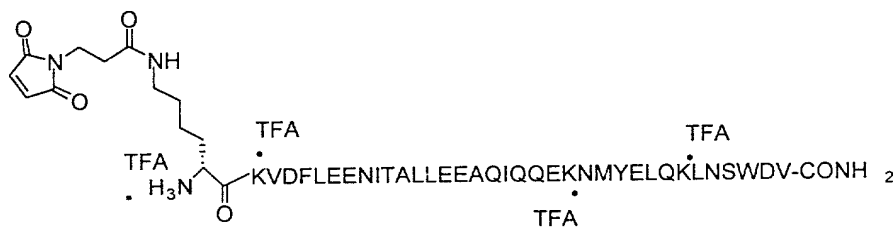
Step 3

3-maleimidopropionic acid



Step 4

85% TFA/5% TIS/5% thioanisole/5% phenol



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Example 28

Preparation of a Modified anti-SIV peptide

5 In this example, the peptide SEQ ID NO:71 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:71 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Phe-OH, Fmoc-Val-OH, 15 Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, 20 Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH, Fmoc-Val-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the 25 Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of 30 CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with

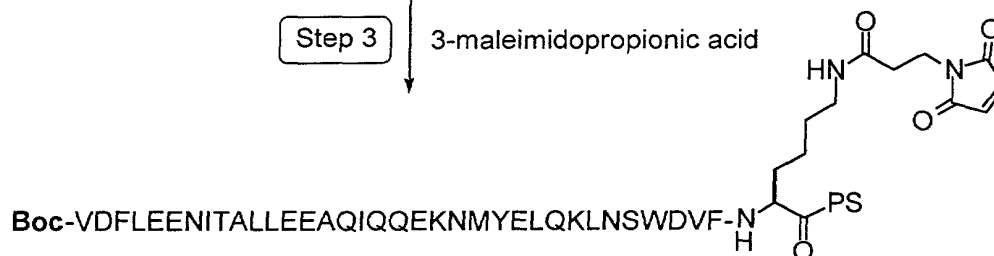
CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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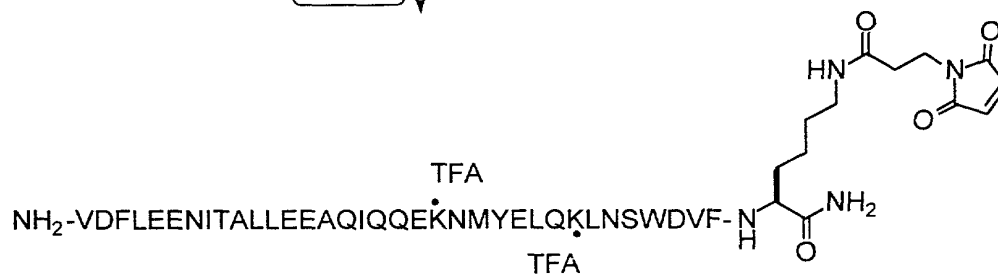
Step 1 SPPS

Step 2 \downarrow Pd(PPh₃)₄/NMM/HOAc/CHCl₃

Step 3 ↓ 3-maleimidopropionic acid



Step 4 ↓ 85% TFA/5% TIS/5% thioanisole/5% phenol



Example 29

Preparation of a Modified anti-SIV peptide

5 In this example, the peptide SEQ ID NO:72 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:72 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Gly-OH, Fmoc-Phe-OH,
15 Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH, Fmoc-Asp(tBu)-OH. They
20 are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N',N'*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of
25 CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with
30

CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6 x 5 mL). The synthesis is then re-automated for the addition of the 3-maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3 times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5% phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is purified by preparative reversed phased HPLC using a Varian (Rainin) preparative binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and 0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide (i.e., DAC) in >95% purity, as determined by RP-HPLC.

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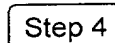
Step 1

Step 2

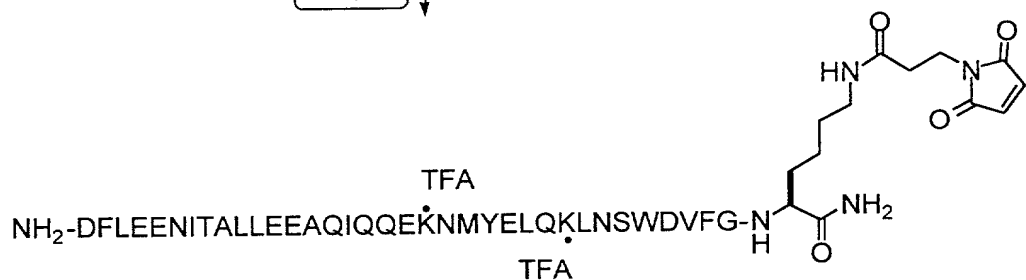
↓

Step 3

—



↓



Example 30

Preparation of a Modified anti-SIV peptide

5 In this example, the peptide SEQ ID NO:73 is synthesized and modified to include a linker and maleimide group according to the synthesis scheme set forth below. As reported in U.S. Pat. Nos. 6,013,236 and 6,020,459, SEQ ID NO:73 exhibits potent antiviral activity as a crude peptide against simian immunodeficiency virus (SIV).

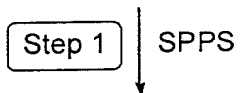
10 Solid phase peptide synthesis of the modified peptide on a 100 μ mole scale is performed using manual solid-phase synthesis, a Symphony Peptide Synthesizer and Fmoc protected Rink Amide MBHA. The following protected amino acids are sequentially added to resin: Fmoc-Lys(Aloc)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gly-OH, Fmoc-Phe-OH, Fmoc-Val-OH, Fmoc-Asp(tBu)-OH, Fmoc-Trp(Boc)-OH, 15 Fmoc-Ser(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Leu-OH, Fmoc-Glu(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Met-OH, Fmoc-Asn(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ile-OH, Fmoc-Gln(Trt)-OH, Fmoc-Ala-OH, Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Thr(tBu)-OH, Fmoc-Ile-OH, Fmoc-Asn(Trt)-OH, 20 Fmoc-Glu(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Leu-OH, Fmoc-Phe-OH. They are dissolved in *N,N*-dimethylformamide (DMF) and, according to the sequence, activated using *O*-benzotriazol-1-yl-*N,N,N,N*-tetramethyl-uronium hexafluorophosphate (HBTU) and Diisopropylethylamine (DIEA). Removal of the 25 Fmoc protecting group is achieved using a solution of 20% (V/V) piperidine in *N,N*-dimethylformamide (DMF) for 20 minutes (step 1). The selective deprotection of the Lys (Aloc) group is performed manually and accomplished by treating the resin with a solution of 3 eq of Pd(PPh₃)₄ dissolved in 5 mL of CHCl₃:NMM:HOAc (18:1:0.5) for 2 h (Step 2). The resin is then washed with 30 CHCl₃ (6 x 5 mL), 20% HOAc in DCM (6 x 5 mL), DCM (6 x 5 mL), and DMF (6

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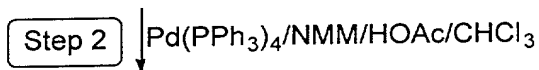
x 5 mL). The synthesis is then re-automated for the addition of the 3-
maleimidopropionic acid (Step 3). Between every coupling, the resin is washed 3
times with *N,N*-dimethylformamide (DMF) and 3 times with isopropanol. The
peptide is cleaved from the resin using 85% TFA/5% TIS/5% thioanisole and 5%
5 phenol, followed by precipitation by dry-ice cold Et₂O (Step 4). The product is
purified by preparative reversed phased HPLC using a Varian (Rainin) preparative
binary HPLC system: gradient elution of 30-55% B (0.045% TFA in H₂O (A) and
0.045% TFA in CH₃CN (B)) over 180 min at 9.5 mL/min using a Phenomenex
Luna 10 μ phenyl-hexyl, 21 mm x 25 cm column and UV detector (Varian
10 Dynamax UVD II) at λ 214 and 254 nm to afford the desired modified peptide
(i.e., DAC) in >95% purity, as determined by RP-HPLC.

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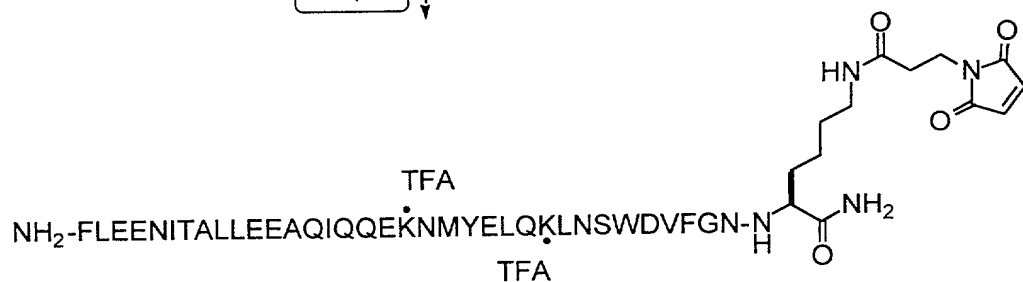
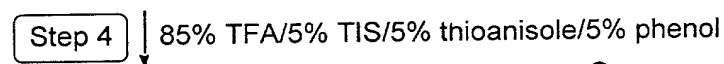
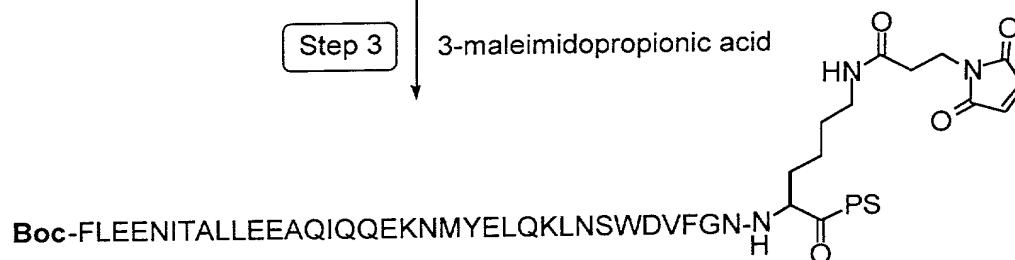
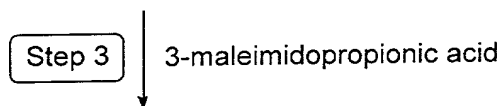
Fmoc-Rink Amide MBHA Resin



Boc-FLEENITALLEEAQIQQEKNMYELQKLNSWDVFGN-Lys(Aloc)-PS



Boc-FLEENITALLEEAQIQQEKNMYELQKLNSWDVFGN-Lys-PS



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1. *Journal of the American Medical Association*, 1997; 277: 1001-1005.

We claim:

1. A modified anti-viral peptide comprising:
a peptide that exhibits anti-viral activity, and
5 a reactive group which is reactive with amino groups, hydroxyl groups, or thiol groups on blood components to form stable covalent bonds.
2. The modified peptide of claim 1 wherein said reactive group is a succinimidyl or a maleimido group.
- 10 3. The modified peptide of claim 1 wherein said reactive group is a maleimido group which is reactive with a thiol group on a blood protein.
4. The modified peptide of claim 1 wherein said peptide is DP178 or
15 DP107 or analogs thereof.
5. The modified peptide of claim 1 wherein said peptide exhibits anti-viral activity against human immunodeficiency virus (HIV).
- 20 6. The modified peptide of claim 5 wherein said peptide is selected from the group consisting of SEQ ID NO:1 to SEQ ID NO:9.
7. The modified peptide of claim 5 wherein said peptide is DP 178 or DP 107.
- 25 8. The modified peptide of claim 1 wherein said peptide exhibits anti-viral activity against human respiratory syncytial virus (RSV).
9. The modified peptide of claim 8 wherein said peptide is selected
30 from the group consisting of SEQ ID NO:10 to SEQ ID NO:30.

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10. The modified peptide of claim 8 wherein said peptide is selected from the group consisting of SEQ ID NO:14 to SEQ ID NO:17 and SEQ ID NO:29.

5

11. The modified peptide of claim 1 wherein said peptide exhibits anti-viral activity against human parainfluenza virus (HPIV).

12. The modified peptide of claim 11 wherein said peptide is selected from the group consisting of SEQ ID NO:31 to SEQ ID NO:62.

10

13. The modified peptide of claim 11 wherein said peptide is selected from the group consisting of SEQ ID NO: 35, SEQ ID NO:38 to SEQ ID NO:42, SEQ ID NO:52 and SEQ ID NO:58.

15

14. The modified peptide of claim 1 wherein said peptide exhibits anti-viral activity against measles virus (MeV).

15. The modified peptide of claim 14 wherein said peptide is selected from the group consisting of SEQ ID NO:74 to SEQ ID NO:86.

20

16. The modified peptide of claim 14 wherein said peptide is selected from the group consisting of SEQ ID NO:77, SEQ ID NO:79, SEQ ID NO:81 and SEQ ID NO:84.

25

17. The modified peptide of claim 1 wherein said peptide exhibits anti-viral activity against simian immunodeficiency virus (SIV).

18. The modified peptide of claim 17 wherein said peptide is selected from the group consisting of SEQ ID NO:63 to SEQ ID NO:73.

30

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19. A composition for use in the prevention and/or treatment of acquired immune deficiency syndrome (AIDS) comprising a peptide that exhibits anti-viral activity against human immunodeficiency virus (HIV), modified with a reactive group which is reactive with amino groups, hydroxyl groups, or thiol groups on blood components to form stable covalent bonds.

20. The composition of claim 19 wherein said reactive group is a maleimido group which is reactive with a thiol group on a blood protein.

21. The composition of claim 20 wherein said peptide is DP178 or DP107 or analogs thereof.

22. A composition for use in the prevention and/or treatment of human respiratory syncytial virus (RSV) infection comprising a peptide that exhibits anti-viral activity against RSV, modified with a reactive group which is reactive with amino groups, hydroxyl groups, or thiol groups on blood components to form stable covalent bonds.

23. The composition of claim 22 wherein said reactive group is a maleimido group which is reactive with a thiol group on a blood protein.

24. The composition of claim 23 wherein said peptide is selected from the group consisting of SEQ ID NO:14 to SEQ ID NO:17 and SEQ ID NO:29.

25. A composition for use in the prevention and/or treatment of human parainfluenza virus (HPIV) infection comprising a peptide that exhibits anti-viral activity against human parainfluenza (HPIV), modified with a reactive group which is reactive with amino groups, hydroxyl groups, or thiol groups on blood components to form stable covalent bonds.

5

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30. The composition of claim 29 wherein said peptide is selected from the group consisting of SEQ ID NO:77, SEQ ID NO:79, SEQ ID NO:81 and SEQ ID NO:84.

ABSTRACT OF THE DISCLOSURE

- 5 Peptides exhibiting anti-viral and anti-fusogenic activity are modified to provide greater stability and improved half-life *in vivo*. The selected peptides include fusion inhibitors DP178 and DP107 and related peptides and analogs thereof. The modified peptides are capable of forming covalent bonds with one or more blood components, preferably a mobile blood component.

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TABLE 2

DP178 CARBOXY TRUNCATIONS	
5	YTS YTSL YTSLI YTSLIH YTSLIHS
10	YTSLIHSL YTSLIHSLI YTSLIHSLIE YTSLIHSLIEE YTSLIHSLIEES
15	YTSLIHSLIEESQ YTSLIHSLIEESQN YTSLIHSLIEESQNNQ YTSLJHSLJEESQNNQ YTSLIHSLIEESQNNQEQ
20	YTSLIHSLIEESQNNQEQEK YTSLIHSLIEESQNNQEQEKN YTSLIHSLIEESQNNQEQEKNE YTSLIHSLIEESQNNQEQEKNEQ YTSLIHSLIEESQNNQEQEKNEQE
25	YTSLIHSLIEESQNNQEQEKNEQEL YTSLIHSLIEESQNNQEQEKNEQELL YTSLIHSLIEESQNNQEQEKNEQELLE YTSLIHSLIEESQNNQEQEKNEQELLEL YTSLIHSLIEESQNNQEQEKNEQELLELD
30	YTSLIHSLIEESQNNQEQEKNEQELLELDK

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YTSLIHSLIEESQNQQEKNEQELLELDKW
YTSLIHSLIEESQNQQEKNEQELLELDKWA
YTSLIHSLIEESQNQQEKNEQELLELDKAS
YTSLIHSLIEESQNQQEKNEQELLELDKWASL
5 YTSLIHSLIEESQNQQEKNEQELLELDKWASLW
YTSLIHSLIEESQNQQEKNEQELLELDKWASLWN
YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNW
YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF

10 The one letter amino acid code of Table 1 is used.

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DP178 AMINO TRUNCATIONS

5	NWF
	WNWF
	LWNWF
	SLWNWF
10	ASLWNWF
	WASLWNWF
	KWASLWNWF
	DKWASLWNWF
	LDKWASLWNWF
15	ELDKWASLWNWF
	LELDKWASLWNWF
	LLELDKWASLWNWF
	ELLELDKWASLWNWF
	QELLELDKWASLWNWF
20	EQELLELDKWASLWNWF
	NEQELLELDKWASLWNWF
	KNEQELLELDKWASLWNWF
	EKNEQELLELDKWASLWNWF
	QEKNEQELLELDKWASLWNWF
25	QQEKNEQELLELDKWASLWNWF
	NQQEKNEQELLELDKWASLWNWF
	QNQQEKNEQELLELDKWASLWNWF
	SQNQQEKNEQELLELDKWASLWNWF
	ESQNQQEKNEQELLELDKWASLWNWF
30	EESQNQQEKNEQELLELDKWASLWNWF

IEESQNQQEKNEQEELLELDKWASLWNWF
LIEESQNQQEKNEQEELLELDKWASLWNWF
SLIEESQNQQEKNEQEELLELDKWASLWNWF
HSLIEESQNQQEKNEQEELLELDKWASLWNWF
5 IHSLIEESQNQQEKNEQEELLELDKWASLWNWF
LIHSLIEESQNQQEKNEQEELLELDKWASLWNWF
SLIHSLIEESQNQQEKNEQEELLELDKWASLWNWF
TSLIHSLIEESQNQQEKNEQEELLELDKWASLWNWF
YTSLIHSLIEESQNQQEKNEQEELLELDKWASLWNWF

10

The one letter amino acid code of Table 1 is used.

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TABLE 4

DP107 CARBOXY TRUNCATIONS	
5	NNL NNLL NNLLR NNLLRA NNLLRAI
10	NNLLRAIE NNLLRAIEA NNLLRAIEAQ NNLLRAIEAQQ NNLLRAIEAQQH
15	NNLLRAIEAQQHL NNLLRAIEAQQHLL NNLLRAIEAQQHLLQ NNLLRAIEAQQHLLQL NNLLRAIEAQQHLLQLT
20	NNLLRAIEAQQHLLQLTV NNLLRAIEAQQHLLQLTVW NNLLRAIEAQQHLLQLTVWQ NNLLRAIEAQQHLLQLTVWQI NNLLRAIEAQQHLLQLTVWQIK
25	NNLLRAIEAQQHLLQLTVWQIKQ NNLLRAIEAQQHLLQLTVWQIKQL NNLLRAIEAQQHLLQLTVWQIKQLQ NNLLRAIEAQQHLLQLTVWQIKQLQA NNLLRAIEAQQHLLQLTVWQIKQLQAR
30	NNLLRAIEAQQHLLQLTVWQIKQLQARI

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NNLLRAIEAQQHLLQLTVWQIKQLQARIL
NNLLRAIEAQQHLLQLTVWQIKQLQARILA
NNLLRAIEAQQHLLQLTVWQIKQLQARILAV
NNLLRAIEAQQHLLQLTVWQIKQLQARILAVE
5 NNLLRAIEAQQHLLQLTVWQIKQLQARILAVER
NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERY
NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYL
NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLK
NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKD
10 NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

The one letter amino acid code of Table 1 is used.

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DP107 AMINO TRUNCATIONS

5	KDQ
	LKDQ
	YLKDQ
	RYLKDQ
	ERYLKDQ
10	VERYLKDQ
	AVERYLKDQ
	LAVERYLKDQ
	ILAVERYLKDQ
	RILAVERYLKDQ
15	ARILAVERYLKDQ
	QARILAVERYLKDQ
	LQARILAVERYLKDQ
	QLQARILAVERYLKDQ
	KQLQARILAVERYLKDQ
20	IKQLQARILAVERYLKDQ
	QIKQLQARILAVERYLKDQ
	WQIKQLQARILAVERYLKDQ
	VWQIKQLQARILAVERYLKDQ
	TVWQIKQLQARILAVERYLKDQ
25	LTVWQIKQLQARILAVERYLKDQ
	QLTVWQIKQLQARILAVERYLKDQ
	LQLTVWQIKQLQARILAVERYLKDQ
	LLQLTVWQIKQLQARILAVERYLKDQ
	HLLQLTVWQIKQLQARILAVERYLKDQ
30	QHLLQLTVWQIKQLQARILAVERYLKDQ

QQHLLQLTVWQIKQLQARILAVERYLKDQ
AQQHLLQLTVWQIKQLQARILAVERYLKDQ
EAQQHLLQLTVWQIKQLQARILAVERYLKDQ
IEAQQHLLQLTVWQIKQLQARILAVERYLKDQ
5 AIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ
RAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ
LRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ
LLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ
NLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ
10 NNLLRAIEAQQHLLQLTVWQIKQLQARILAVERYLKDQ

The one letter amino acid code of Table 1 is used.

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TABLE 6

HIV-2_{NIH} DP178 analog carboxy truncations.

5	LEA
	LEAN
	LEANI
	LEANIS
	LEANISQ
10	LEANISQS
	LEANISQSL
	LEANISQSLE
	LEANISQSLEQ
	LEANISQSLEQA
15	LEANISQSLEQAAQ
	LEANISQSLEQAQI
	LEANISQSLEQAQIQ
	LEANISQSLEQAQIQQ
	LEANISQSLEQAQIQQE
20	LEANISQSLEQAQIQQEK
	LEANISQSLEQAQIQQEKN
	LEANISQSLEQAQIQQEKNM
	LEANISQSLEQAQIQQEKNMY
	LEANISQSLEQAQIQQEKNMYE
25	LEANISQSLEQAQIQQEKNMYEL
	LEANISQSLEQAQIQQEKNMYELQ
	LEANISQSLEQAQIQQEKNMYELQK
	LEANISQSLEQAQIQQEKNMYELQKL
	LEANISQSLEQAQIQQEKNMYELQKLN
30	LEANISQSLEQAQIQQEKNMYELQKLNS

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LEANISQSLEQAQIQQEKNMYELQKLNSW
LEANISQSLEQAQIQQEKNMYELQKLNSWD
LEANISQSLEQAQIQQEKNMYELQKLNSWDV
LEANISQSLEQAQIQQEKNMYELQKLNSWDVF
5 LEANISQSLEQAQIQQEKNMYELQKLNSWDVFT
LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTN
LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNW
LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

10 The one letter amino acid code of Table 1 is used.

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TABLE 7

HIV-2_{NIH} DP178 analog amino truncations.

5	NWL
	TNWL
	FTNWL
	VFTNWL
	DVFTNWL
10	WDVFTNWL
	SWDVFTNWL
	NSWDVFTNWL
	LNSWDVFTNWL
	KLNSWDVFTNWL
15	QKLNSWDVFTNWL
	LQKLNSWDVFTNWL
	ELQKLNSWDVFTNWL
	YELQKLNSWDVFTNWL
	MYELQKLNSWDVFTNWL
20	NMYELQKLNSWDVFTNWL
	KNMYELQKLNSWDVFTNWL
	EKNMYELQKLNSWDVFTNWL
	QEKNMYELQKLNSWDVFTNWL
	QQEKNMYELQKLNSWDVFTNWL
25	IQQEKNMYELQKLNSWDVFTNWL
	QIQQEKNMYELQKLNSWDVFTNWL
	AQIQQEKNMYELQKLNSWDVFTNWL
	QAQIQQEKNMYELQKLNSWDVFTNWL
	EQAQIQQEKNMYELQKLNSWDVFTNWL
30	LEQAQIQQEKNMYELQKLNSWDVFTNWL

SLEQAQIQQEKNMYELQKLNSWDVFTNWL
QSLEQAQIQQEKNMYELQKLNSWDVFTNWL
SQSLEQAQIQQEKNMYELQKLNSWDVFTNWL
ISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL
5 NISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL
ANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL
EANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL
LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL

10 The one letter amino acid code of Table 1 is used.

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TABLE 8

RESPIRATORY SYNCYTIAL VIRUS (RSV) DP107 F2 REGION ANALOG CARBOXY TRUNCATIONS	
5	YTS
	YTSV
	YTSVI
	YTSVIT
10	YTSVITI
	YTSVITIE
	YTSVITIEL
	YTSVITIELS
	YTSVITIELSN
15	YTSVITIELSNI
	YTSVITIELSNIK
	YTSVITIELSNIKE
	YTSVITIELSNIKEN
	YTSVITIELSNIKENK
20	YTSVITIELSNIKENKC
	YTSVITIELSNIKENKCN
	YTSVITIELSNIKENKCNG
	YTSVITIELSNIKENKCNGT
	YTSVITIELSNIKENKCNGTD
25	YTSVITIELSNIKENKCNGTDA
	YTSVITIELSNIKENKCNGTDAK
	YTSVITIELSNIKENKCNGTDAKV
	YTSVITIELSNIKENKCNGTDAKVK
	YTSVITIELSNIKENKCNGTDAKVKL
30	YTSVITIELSNIKENKCNGTDAKVKLI

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YTSVITIELSNIKENKNGTDAKVKLIK
YTSVITIELSNIKENKNGTDAKVKLIKQ
YTSVITIELSNIKENKNGTDAKVKLIKQE
YTSVITIELSNIKENKNGTDAKVKLIKQEL
5 YTSVITIELSNIKENKNGTDAKVKLIKQELD
YTSVITIELSNIKENKNGTDAKVKLIKQELDK
YTSVITIELSNIKENKNGTDAKVKLIKQELDKY
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYK
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKN
10 YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNA
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAV
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAV
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTE
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTEL
15 YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTELQ
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTELQL
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTELQLL
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTELQLLM
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTELQLLMQ
20 YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTELQLLMQS
YTSVITIELSNIKENKNGTDAKVKLIKQELDKYKNAVTELQLLMQST

The one letter amino acid code of Table 1 is used.

TABLE 9

RESPIRATORY SYNCYTIAL VIRUS (RSV) DP107 F2 REGION ANALOG AMINO TRUNCATIONS	
5	QST MQST LMQST LLMQST
10	QLLMQST LQLLMQST ELQLLMQST TELQLLMQST VTELQLLMQST
15	AVTELQLLMQST NAVTELQLLMQST KNAVTELQLLMQST YKNAVTELQLLMQST KYKNAVTELQLLMQST
20	DKYKNAVTELQLLMQST LDKYKNAVTELQLLMQST ELDKYKNAVTELQLLMQST QELDKYKNAVTELQLLMQST KQELDKYKNAVTELQLLMQST
25	IKQELDKYKNAVTELQLLMQST LIKQELDKYKNAVTELQLLMQST KLIKQELDKYKNAVTELQLLMQST VKLIKQELDKYKNAVTELQLLMQST KVKLIKQELDKYKNAVTELQLLMQST
30	AKVKLIKQELDKYKNAVTELQLLMQST

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DAKVKLIKQELDKYKNAVTELQLLMQST
TDAKVKLIKQELDKYKNAVTELQLLMQST
GTDAKVKLIKQELDKYKNAVTELQLLMQST
NGTDAKVKLIKQELDKYKNAVTELQLLMQST
5 CNGTDAKVKLIKQELDKYKNAVTELQLLMQST
KCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
NKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
KENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
IKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
10 NIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
SNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
LSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
ELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
IELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
15 TIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
ITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
VITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
SVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST
TSVITIELSNIKENKCNGTDAKVKLIKQELDKYKNAVTELQLLMQST

20

The one letter amino acid code of Table 1 is used.

TABLE 10

RESPIRATORY SYNCYTIAL VIRUS (RSV) F1 DP178 REGION ANALOG CARBOXY TRUNCATIONS	
5	FYD FYDP FYDPL FYDPLV
10	FYDPLVF FYDPLVFP FYDPLVFPS FYDPLVFPSD FYDPLVFPSDE
15	FYDPLVFPSDEF FYDPLVFPSDEFD FYDPLVFPSDEFDA FYDPLVFPSDEFDAS FYDPLVFPSDEFDASI
20	FYDPLVFPSDEFDASIS FYDPLVFPSDEFDASISQ FYDPLVFPSDEFDASISQV FYDPLVFPSDEFDASISQVN FYDPLVFPSDEFDASISQVNE
25	FYDPLVFPSDEFDASISQVNEK FYDPLVFPSDEFDASISQVNEKI FYDPLVFPSDEFDASISQVNEKIN FYDPLVFPSDEFDASISQVNEKINQ FYDPLVFPSDEFDASISQVNEKINQS
30	FYDPLVFPSDEFDASISQVNEKINQSL

FYDPLVFPSDEFDASISQVNEKINQSLA
FYDPLVFPSDEFDASISQVNEKINQSLAF
FYDPLVFPSDEFDASISQVNEKINQSLAFI
FYDPLVFPSDEFDASISQVNEKINQSLAFIR
5 FYDPLVFPSDEFDASISQVNEKINQSLAFIRK
FYDPLVFPSDEFDASISQVNEKINQSLAFIRKS
FYDPLVFPSDEFDASISQVNEKINQSLAFIRKSD
FYDPLVFPSDEFDASISQVNEKINQSLAFIRKSDE
FYDPLVFPSDEFDASISQVNEKINQSLAFIRKSDEL
10 FYDPLVFPSDEFDASISQVNEKINQSLAFIRKSDELL

The one letter amino acid code of Table 1 is used.

004060" 9E26960

TABLE 11

RESPIRATORY SYNCYTIAL VIRUS (RSV) F1 DP178 REGION ANALOG AMINO TRUNCATIONS	
5	DELL SDELL KSDELL RKSDDELL
10	IRKSDELL FIRKSDELL AFIRKSDELL LAFIRKSDELL SLAFIRKSDELL
15	QSLAFIRKSDELL NQSLAFIRKSDELL INQSLAFIRKSDELL KINQSLAFIRKSDELL EKINQSLAFIRKSDELL
20	NEKINQSLAFIRKSDELL VNEKINQSLAFIRKSDELL QVNEKINQSLAFIRKSDELL SQVNEKINQSLAFIRKSDELL ISQVNEKINQSLAFIRKSDELL
25	SISQVNEKINQSLAFIRKSDELL ASISQVNEKINQSLAFIRKSDELL DASISQVNEKINQSLAFIRKSDELL FDASISQVNEKINQSLAFIRKSDELL EFDASISQVNEKINQSLAFIRKSDELL
30	DEFDASISQVNEKINQSLAFIRKSDELL

004060" 00000000

SDEFDASISQVNEKINQSLAFIRKSDELL
PSDEFDASISQVNEKINQSLAFIRKSDELL
FPSDEFDASISQVNEKINQSLAFIRKSDELL
VFPSDEFDASISQVNEKINQSLAFIRKSDELL
5 LVFPSDEFDASISQVNEKINQSLAFIRKSDELL
PLVFPSDEFDASISQVNEKINQSLAFIRKSDELL
DPLVFPSDEFDASISQVNEKINQSLAFIRKSDELL
YDPLVFPSDEFDASISQVNEKINQSLAFIRKSDELL

10 The one letter amino acid code of Table 1 is used.

002060" 9EE25950

TABLE 12

HUMAN PARAINFLUENZA VIRUS 3 (HPV3) F1 REGION DP178
ANALOG CARBOXY TRUNCATIONS

5	ITL
	ITLN
	ITLNN
	ITLNNS
10	ITLNNSV
	ITLNNSVA
	ITLNNSVAL
	ITLNNSVALD
	ITLNNSVALDP
15	ITLNNSVALDPI
	ITLNNSVALDPID
	ITLNNSVALDPIDI
	ITLNNSVALDPIDIS
	ITLNNSVALDPIDISI
20	ITLNNSVALDPIDISIE
	ITLNNSVALDPIDISIEL
	ITLNNSVALDPIDISIELN
	ITLNNSVALDPIDISIELNK
	ITLNNSVALDPIDISIELNKA
25	ITLNNSVALDPIDISIELNKAK
	ITLNNSVALDPIDISIELNKAKS
	ITLNNSVALDPIDISIELNKAKSD
	ITLNNSVALDPIDISIELNKAKSDL
	ITLNNSVALDPIDISIELNKAKSDLE
30	ITLNNSVALDPIDISIELNKAKSDLEE

ITLNNSVALDPIDISIELNKAUSDLEES
ITLNNSVALDPIDISIELNKAUSDLEESK
ITLNNSVALDPIDISIELNKAUSDLEESKE
ITLNNSVALDPIDISIELNKAUSDLEESKEW
5 ITLNNSVALDPIDISIELNKAUSDLEESKEWI
ITLNNSVALDPIDISIELNKAUSDLEESKEWIR
ITLNNSVALDPIDISIELNKAUSDLEESKEWIRR
ITLNNSVALDPIDISIELNKAUSDLEESKEWIRRS

10 The one letter amino acid code of Table 1 is used.

00/050" 926/5950

TABLE 13

HUMAN PARAINFLUENZA VIRUS 3 (HPV3) F1 REGION DP178	
ANALOG AMINO TRUNCATIONS	
5	RRS
	IRRS
	WIRRS
	EWIRRS
10	KEWIRRS
	SKEWIRRS
	ESKEWIRRS
	EESKEWIRRS
	LEESKEWIRRS
15	DLEESKEWIRRS
	SDLEESKEWIRRS
	KSDLEESKEWIRRS
	AKSDLEESKEWIRRS
	KAKSDLEESKEWIRRS
20	NKAKSDLEESKEWIRRS
	LNKAKSDLEESKEWIRRS
	ELNKAKSDLEESKEWIRRS
	IELNKAKSDLEESKEWIRRS
	SIELNKAKSDLEESKEWIRRS
25	ISIELNKAKSDLEESKEWIRRS
	DISIELNKAKSDLEESKEWIRRS
	IDISIELNKAKSDLEESKEWIRRS
	PIDISIELNKAKSDLEESKEWIRRS
	DPIDISIELNKAKSDLEESKEWIRRS
30	LDPIDISIELNKAKSDLEESKEWIRRS

The one letter amino acid code of Table 1 is used.

TABLE 14

HUMAN PARAINFLUENZA VIRUS 3 (HPV3) F1 REGION	
DP107 ANALOG CARBOXY TRUNCATIONS	
5	ALG
	ALGV
	ALGVA
	ALGVAT
10	ALGVATS
	ALGVATSA
	ALGVATSAQ
	ALGVATSAQI
	ALGVATSAQIT
15	ALGVATSAQITA
	ALGVATSAQITAA
	ALGVATSAQITAAV
	ALGVATSAQITAAVA
	ALGVATSAQITAVAL
20	ALGVATSAQITAVALV
	ALGVATSAQITAVALVE
	ALGVATSAQITAVALVEA
	ALGVATSAQITAVALVEAK
	ALGVATSAQITAVALVEAKQ
25	ALGVATSAQITAVALVEAKQA
	ALGVATSAQITAVALVEAKQAR
	ALGVATSAQITAVALVEAKQARS
	ALGVATSAQITAVALVEAKQARSD
	ALGVATSAQITAVALVEAKQARSDI
30	ALGVATSAQITAVALVEAKQARSDIE

002060" 9225950

ALGVATSAQITA AVALVEAKQARSDIEK
ALGVATSAQITA AVALVEAKQARSDIEKL
ALGVATSAQITA AVALVEAKQARSDIEKLK
ALGVATSAQITA AVALVEAKQARSDIEKLKE
5 ALGVATSAQITA AVALVEAKQARSDIEKLKEA
ALGVATSAQITA AVALVEAKQARSDIEKLKEAI
ALGVATSAQITA AVALVEAKQARSDIEKLKEAIR

The one letter amino acid code of Table 1 is used.

002060" 9EE25950

TABLE 15

HUMAN PARAINFLUENZA VIRUS 3 (HPV3) F1 REGION	
DP107 ANALOG AMINO TRUNCATIONS	
5	IRD
	AIRD
	EAIRD
	KEAIRD
10	LKEAIRD
	KLKEAIRD
	EKLKEAIRD
	IEKLKEAIRD
	DIEKLKEAIRD
15	SDIEKLKEAIRD
	RSDIEKLKEAIRD
	ARSDIEKLKEAIRD
	QARSDIEKLKEAIRD
	KQARSDIEKLKEAIRD
20	AKQARSDIEKLKEAIRD
	EAKQARSDIEKLKEAIRD
	VEAKQARSDIEKLKEAIRD
	LVEAKQARSDIEKLKEAIRD
	ALVEAKQARSDIEKLKEAIRD
25	VALVEAKQARSDIEKLKEAIRD
	AVALVEAKQARSDIEKLKEAIRD
	AAVALVEAKQARSDIEKLKEAIRD
	TAAVALVEAKQARSDIEKLKEAIRD
	ITAVALVEAKQARSDIEKLKEAIRD
30	QITAVALVEAKQARSDIEKLKEAIRD

002060" 92275960

AQITA AVALVEAKQARSDIEKLKEAIRD
SAQITA AVALVEAKQARSDIEKLKEAIRD
TSAQITA AVALVEAKQARSDIEKLKEAIRD
ATSAQITA AVALVEAKQARSDIEKLKEAIRD
5 VATSAQITA AVALVEAKQARSDIEKLKEAIRD
GVATSAQITA AVALVEAKQARSDIEKLKEAIRD
LGVATSAQITA AVALVEAKQARSDIEKLKEAIRD

The one letter amino acid code of Table 1 is used.

090700 "000000" 000000

TABLE 16

ANTI-RESPIRATORY SYNCYTIAL VIRUS (RSV) PEPTIDES

5	TSVITIELSNIKENKCNGTDAKVKLIKQELDKYKN SVITIELSNIKENKCNGTDAKVKLIKQELDKYKNA VITIELSNIKENKCNGTDAKVKLIKQELDKYKNAV VAVSKVLHLEGEVNKIALSTNKAVVSLSNQVSV AVSKVLHLEGEVNKIALSTNKAVVSLSNQVSV
10	VSKVLHLEGEVNKIALSTNKAVVSLSNQVSVL SKVLHLEGEVNKIALSTNKAVVSLSNQVSVLT KVLHLEGEVNKIALSTNKAVVSLSNQVSVLTS LEGEVNKIALSTNKAVVSLSNQVSVLTSKVLD GEVNKIALSTNKAVVSLSNQVSVLTSKVLDLK
15	EVNKIALSTNKAVVSLSNQVSVLTSKVLDLKN VNKIALSTNKAVVSLSNQVSVLTSKVLDLKNY NKIALSTNKAVVSLSNQVSVLTSKVLDLKNYI KIALSTNKAVVSLSNQVSVLTSKVLDLKNYID IALSTNKAVVSLSNQVSVLTSKVLDLKNYIDK
20	ALLSTNKAVVSLSNQVSVLTSKVLDLKNYIDKQ VAVSKVLHLEGEVNKIALSTNKAVVSLSNQVSV AVSKVLHLEGEVNKIALSTNKAVVSLSNQVSV VSKVLHLEGEVNKIALSTNKAVVSLSNQVSVL SKVLHLEGEVNKIALSTNKAVVSLSNQVSVLT
25	KVLHLEGEVNKIALSTNKAVVSLSNQVSVLTS LEGEVNKIALSTNKAVVSLSNQVSVLTSKVLD GEVNKIALSTNKAVVSLSNQVSVLTSKVLDLK EVNKIALSTNKAVVSLSNQVSVLTSKVLDLKN VNKIALSTNKAVVSLSNQVSVLTSKVLDLKNY
30	NKIALSTNKAVVSLSNQVSVLTSKVLDLKNYI

KIALSTNKAVVSLSNGVSVLTSTKVLDLKNYID
IALSTNKAVVSLSNGVSVLTSTKVLDLKNYIDK
ALLSTNKAVVSLSNGVSVLTSTKVLDLKNYIDKQ

-
- 5 The one letter amino acid code of Table 1 is used.

004060" 9884960

TABLE 17

ANTI-HUMAN PARAINFLUENZA VIRUS 3 (HPV3) PEPTIDES

5	TLNNSVALDPIDISIELNKA	SDLEESKEWIRRSN
	LNNSVALDPIDISIELNKA	SDLEESKEWIRRSNQ
	NNSVALDPIDISIELNKA	SDLEESKEWIRRSNQK
	NSVALDPIDISIELNKA	SDLEESKEWIRRSNQKL
	SVALDPIDISIELNKA	SDLEESKEWIRRSNQKLD
10	VALDPIDISIELNKA	SDLEESKEWIRRSNQKLDS
	ALDPIDISIELNKA	SDLEESKEWIRRSNQKLDSI
	LDPIDISIELNKA	SDLEESKEWIRRSNQKLDSIG
	DPIDISIELNKA	SDLEESKEWIRRSNQKLDSIGN
	PIDISIELNKA	SDLEESKEWIRRSNQKLDSIGNW
15	IDISIELNKA	SDLEESKEWIRRSNQKLDSIGNWH
	DISIELNKA	SDLEESKEWIRRSNQKLDSIGNWHQ
	ISIELNKA	SDLEESKEWIRRSNQKLDSIGNWHQS
	SIELNKA	SDLEESKEWIRRSNQKLDSIGNWHQSS
	IELNKA	SDLEESKEWIRRSNQKLDSIGNWHQSST
20	ELNKA	SDLEESKEWIRRSNQKLDSIGNWHQSSTT
	TAAVALVEAKQARSDIEKLKEAIRD	TNKAVQSVQS
	AVALVEAKQARSDIEKLKEAIRD	TNKAVQSVQSSI
	LVEAKQARSDIEKLKEAIRD	TNKAVQSVQSSIGNL
	VEAKQARSDIEKLKEAIRD	TNKAVQSVQSSIGNLI
25	EAKQARSDIEKLKEAIRD	TNKAVQSVQSSIGNLIV
	AKQARSDIEKLKEAIRD	TNKAVQSVQSSIGNLIVA
	KQARSDIEKLKEAIRD	TNKAVQSVQSSIGNLIVAI
	QARSDIEKLKEAIRD	TNKAVQSVQSSIGNLIVAIK
	ARSDIEKLKEAIRD	TNKAVQSVQSSIGNLIVAIKS
30	RS	DIEKLKEAIRD
	TNKAVQSVQSSIGNLIVAIKSV	

096736-090700

SDIEKLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQ
KLKEAIRDTNKAVQSVQSSIGNLIVAIKSVQDYVN
LKEAIRDTNKAVQSVQSSIGNLIVAIKSVQDYVNK
AIRDTNKAVQSVQSSIGNLIVAIKSVQDYVNKEIV

5

The one letter amino acid code of Table 1 is used.

004060"SEE 48960

TABLE 18

ANTI-SIMIAN IMMUNODEFICIENCY VIRUS (SIV) PEPTIDES

5 WQEWERKVD FLEENITALLEEAQIQQEKNMYELQK
 QEWERKVD FLEENITALLEEAQIQQEKNMYELQKL
 EWERKVD FLEENITALLEEAQIQQEKNMYELQKLN
 WERKVD FLEENITALLEEAQIQQEKNMYELQKLNS
 ERKVD FLEENITALLEEAQIQQEKNMYELQKLNSW
10 RKVD FLEENITALLEEAQIQQEKNMYELQKLNSWD
 KVD FLEENITALLEEAQIQQEKNMYELQKLNSWDV
 VDFLEENITALLEEAQIQQEKNMYELQKLNSWDVF
 DFLEENITALLEEAQIQQEKNMYELQKLNSWDVFG
 FLEENITALLEEAQIQQEKNMYELQKLNSWDVFGN

15

The one letter amino acid code of Table 1 is used.

002060" 9EE 25960

TABLE 19

ANTI-MEASLES VIRUS (MEV) PEPTIDES

5	LHRIDLGPISLERLDVGTNLGNAIAKLEAKELL HRIDLGPISLERLDVGTNLGNAIAKLEAKELLE RIDLGPISLERLDVGTNLGNAIAKLEAKELLES IDLGPISLERLDVGTNLGNAIAKLEAKELLESS DLGPISLERLDVGTNLGNAIAKLEAKELLESSD
10	LGPPISLERLDVGTNLGNAIAKLEAKELLESSDQ GPPISLERLDVGTNLGNAIAKLEAKELLESSDQI PPISLERLDVGTNLGNAIAKLEAKELLESSDQIL PISLERLDVGTNLGNAIAKLEAKELLESSDQILR SLERLDVGTNLGNAIAKLEAKELLESSDQILRSM
15	LERLDVGTNLGNAIAKLEAKELLESSDQILRSMK

The one letter amino acid code of Table 1 is used.

096336-090700

COMBINED DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL INFECTION

the specification of which (check one) X is attached hereto or ___ was filed on _____ as Application No. _____ and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information which is material to patentability as defined in 37 CFR § 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)			Priority Claimed	
			Yes	No
Number	Country	Day/Month/Year Filed		
Number	Country	Day/Month/Year Filed		

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States provisional application(s) below.

60/153,406	10 September 1999
Application Number	Filing Date

Application Number	Filing Date
--------------------	-------------

I hereby claim the benefit under 35 U.S.C. § 120 of any United States application(s), or § 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose all information which is material to patentability as defined in 37 CFR § 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Application Number	Filing Date	Status: Patented, Pending, Abandoned
--------------------	-------------	--------------------------------------

Application Number	Filing Date	Status: Patented, Pending, Abandoned
--------------------	-------------	--------------------------------------

I HEREBY APPOINT THE FOLLOWING AS MY ATTORNEYS WITH FULL POWER OF SUBSTITUTION TO PROSECUTE THIS APPLICATION AND TRANSACT ALL BUSINESS IN THE PATENT OFFICE CONNECTED THEREWITH:


Karl A. Limbach	18,689	Alfred A. Equitz	30,922	Mayumi Maeda	40,075
George C. Limbach	19,305	Charles P. Sammut	28,901	Michael R. Ward	38,651
John K. Uilkema	20,282	Mark C. Pickering	36,239	Roger S. Sampson	44,314
Neil A. Smith	25,441	Patricia Coleman James	37,155	Charles L. Hamilton	42,624
Veronica C. Devitt	29,375	Kathleen A. Frost	37,326	Andrew V. Smith	43,132
Ronald L. Yin	27,607	Alan A. Limbach	39,749	Eric N. Hoover	37,355
Gerald T. Sekimura	30,103	Douglas C. Limbach	35,249	Frank J. Mycroft	P-46,946
Michael A. Stallman	29,444	Seong-Kun Oh*		Parisa Jorjani	P-46,813
Philip A. Girard	28,848	Cameron A. King	41,897	Robert M. McConnell	P-46,912
Michael J. Pollock	29,098	Kyla L. Harriel	41,816	J. Thomas McCarthy	22,420
Stephen M. Everett	30,050			Joel G. Ackerman	24,307

* Recognition under 37 CFR 10.9(b)

Send correspondence to Limbach & Limbach L.L.P.
Attn: Michael R. Ward, Esq.
2001 Ferry Building
San Francisco, CA 94111
Telephone: 415/433-4150

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under 18 U.S.C. § 1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole or first inventor DOMINIQUE P. BRIDON

Inventor's signature 

Date

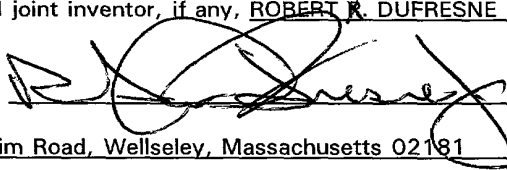
Aug 15, 2000

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Citizenship France

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Inventor's signature 

Date

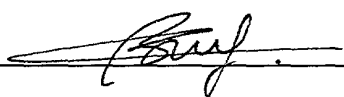
Aug 15, 2000

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Full name of third joint inventor, if any, NISSAB BOUDJELLAB

Inventor's signature 

Date

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H9S 3X1

Full name of fourth joint inventor, if any, MARTIN ROBITAILLE

Inventor's signature Rosanne

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Citizenship Canada

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Full name of fifth joint inventor, if any, PETER G. MILNER

Inventor's signature _____

Date _____

Residence 14690 Manuella Road, Los Altos Hills, California 94022

Citizenship United Kingdom

Post Office Address 14690 Manuella Road, Los Altos Hills, California 94022[illegible]

COMBINED DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

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I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL INFECTION

the specification of which (check one) X is attached hereto or ____ was filed on _____ as Application No. _____ and was amended on _____ (if applicable).

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Prior Foreign Application(s)

Priority Claimed	
Yes	No

Number	Country	Day/Month/Year Filed
--------	---------	----------------------

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60/153,406	10 September 1999
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Application Number	Filing Date	Status: Patented, Pending, Abandoned
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Application Number	Filing Date	Status: Patented, Pending, Abandoned
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I HEREBY APPOINT THE FOLLOWING AS MY ATTORNEYS WITH FULL POWER OF SUBSTITUTION TO PROSECUTE THIS APPLICATION AND TRANSACT ALL BUSINESS IN THE PATENT OFFICE CONNECTED THEREWITH:

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Veronica C. Devitt	29,375	Kathleen A. Frost	37,326	Andrew V. Smith	43,132
Ronald L. Yin	27,607	Alan A. Limbach	39,749	Eric N. Hoover	37,355
Gerald T. Sekimura	30,103	Douglas C. Limbach	35,249	Frank J. Mycroft	P-46,946
Michael A. Stallman	29,444	Seong-Kun Oh*		Parisa Jorjani	P-46,813
Philip A. Girard	28,848	Cameron A. King	41,897	Robert M. McConnell	P-46,912
Michael J. Pollock	29,098	Kyla L. Harriel	41,816	J. Thomas McCarthy	22,420
Stephen M. Everett	30,050			Joel G. Ackerman	24,307

* Recognition under 37 CFR 10.9(b)

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2001 Ferry Building
San Francisco, CA 94111
Telephone: 415/433-4150

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under 18 U.S.C. § 1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole or first inventor DOMINIQUE P. BRIDON

Inventor's signature _____ Date _____
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Citizenship France
Post Office Address 243 Chemin Cote Ste-Catherine, Outremont, Quebec H2V 2B2, Canada

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Inventor's signature _____ Date _____
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Citizenship United States of America
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Full name of third joint inventor, if any, NISSAB BOUDJELLAB

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Inventor's signature _____

Date

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Citizenship Canada

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Full name of fifth joint inventor, if any, PETER G. MILNER

Inventor's signature _____

Peter G. Milner

Date

8/22/2000

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Citizenship United Kingdom

Post Office Address 14690 Manuella Road, Los Altos Hills, California 94022

004060" 92245960

SEQUENCE LISTING

Express Mail: EL254111583US

<110> Conjuchem, Inc.

<120> LONG LASTING FUSION PEPTIDE INHIBITORS OF VIRAL
INFECTION

<130> REDC-1510

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<150> US 60/134,406

<151> 1999-05-17

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<151> 1999-09-10

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<170> PatentIn Ver. 2.1

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<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
peptide

<400> 1

Tyr	Thr	Ser	Leu	Ile	His	Ser	Leu	Ile	Glu	Glu	Ser	Gln	Asn	Gln	Gln
1					5				10					15	

Glu	Lys	Asn	Glu	Gln	Glu	Leu	Leu	Glu	Leu	Asp	Lys	Trp	Ala	Ser	Leu
			20					25					30		

Trp	Asn	Trp	Phe
			35

<210> 2

<211> 38

<212> PRT

<213> Artificial Sequence

<220>

[illegible]

Asn Asn Leu Leu Arg Ala Ile Glu Ala Gln Gln His Leu Leu Gln Leu
1 5 10 15

Thr Val Trp Gln Ile Lys Gln Leu Gln Ala Arg Ile Leu Ala Val Glu
20 25 30

Arg Tyr Leu Lys Asp Gln
35

<211> 36

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic peptide

Tyr Thr Asn Thr Ile Tyr Thr Leu Leu Glu Glu Ser Gln Asn Gln Gln
1 5 10 15

Glu Lys Asn Glu Gln Glu Leu Leu Glu Leu Asp Lys Trp Ala Ser Leu
20 25 30

Trp Asn Trp Phe
35

<211> 36

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic peptide

Tyr Thr Gly Ile Ile Tyr Asn Leu Leu Glu Glu Ser Gln Asn Gln Gln
1 5 10 15

Glu Lys Asn Glu Gln Glu Leu Leu Glu Leu Asp Lys Trp Ala Asn Leu

002060"9EEZ3950

```

                20                25                30

Trp Asn Trp Phe
      35

<210> 5
<211> 36
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
      peptide

<400> 5
Tyr Thr Ser Leu Ile Tyr Ser Leu Leu Glu Lys Ser Gln Thr Gln Gln
  1              5              10              15

Glu Lys Asn Glu Gln Glu Leu Leu Glu Leu Asp Lys Trp Ala Ser Leu
      20              25              30

Trp Asn Trp Phe
      35

<210> 6
<211> 36
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
      peptide

<400> 6
Leu Glu Ala Asn Ile Ser Lys Ser Leu Glu Gln Ala Gln Ile Gln Gln
  1              5              10              15

Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp Asp Ile Phe
      20              25              30

Gly Asn Trp Phe
      35

<210> 7
<211> 36
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<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 7
Leu Glu Ala Asn Ile Ser Gln Ser Leu Glu Gln Ala Gln Ile Gln Gln
1 5 10 15
Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp Asp Val Phe
20 25 30
Thr Asn Trp Leu
35

<210> 8
<211> 41
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 8
Cys Gly Gly Asn Asn Leu Leu Arg Ala Ile Glu Ala Gln Gln His Leu
1 5 10 15
Leu Gln Leu Thr Val Trp Gly Ile Lys Gln Leu Gln Ala Arg Ile Leu
20 25 30
Ala Val Glu Arg Tyr Leu Lys Asp Gln
35 40

<210> 9
<211> 38
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 9

Gln Gln Leu Leu Asp Val Val Lys Arg Gln Gln Glu Met Leu Arg Leu
 1 5 10 15

Thr Val Trp Gly Thr Lys Asn Leu Gln Ala Arg Val Thr Ala Ile Glu
 20 25 30

Lys Tyr Leu Lys Asp Gln
 35

<210> 10

<211> 46

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
 peptide

<400> 10

Tyr Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys
 1 5 10 15

Cys Asn Gly Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys Tyr
 20 25 30

Lys Asn Ala Val Thr Glu Leu Gln Leu Leu Met Gln Ser Thr
 35 40 45

<210> 11

<211> 54

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
 peptide

<400> 11

Ala Ser Gly Val Ala Val Ser Lys Val Leu His Leu Glu Gly Glu Val
 1 5 10 15

Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser
 20 25 30

Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr
 35 40 45

Ile Asp Lys Gln Leu Leu
50

<210> 12
<211> 53
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 12
Gly Glu Pro Ile Ile Asn Phe Tyr Asp Pro Leu Val Phe Pro Ser Asp
1 5 10 15
Glu Phe Asp Ala Ser Ile Ser Gln Val Asn Glu Lys Ile Asn Gln Ser
20 25 30
Leu Ala Phe Ile Arg Lys Ser Asp Glu Leu Leu His Asn Val Asn Ala
35 40 45
Gly Lys Ser Thr Thr
50

<210> 13
<211> 48
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 13
Tyr Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys
1 5 10 15
Cys Asn Gly Thr Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp
20 25 30
Lys Tyr Lys Asn Ala Val Thr Glu Leu Gln Leu Leu Met Gln Ser Thr
35 40 45

<210> 14
 <211> 34
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: synthetic
 peptide

<400> 14
 Tyr Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys
 1 5 10 15

Cys Asn Gly Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys
 20 25 30

Tyr Lys

<210> 15
 <211> 34
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: synthetic
 peptide

<400> 15
 Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys Cys
 1 5 10 15

Asn Gly Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys Tyr
 20 25 30

Lys Asn

<210> 16
 <211> 34
 <212> PRT
 <213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 16

Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys Cys Asn Gly
1 5 10 15

Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys Tyr Lys Asn
20 25 30

Ala Val

<210> 17

<211> 34

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 17

Val Ile Thr Ile Glu Leu Ser Asn Ile Lys Glu Asn Lys Met Asn Gly
1 5 10 15

Asp Ala Lys Val Lys Leu Ile Lys Gln Glu Leu Asp Lys Tyr Lys Asn
20 25 30

Ala Val

<210> 18

<211> 33

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 18

Val Ala Val Ser Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile
1 5 10 15

Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val
 20 25 30

Ser

<210> 19
 <211> 33
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: synthetic
 peptide

<400> 19
 Ala Val Ser Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile Ala
 1 5 10 15

Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val Ser
 20 25 30

Val

<210> 20
 <211> 33
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: synthetic
 peptide

<400> 20
 Val Ser Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile Ala Leu
 1 5 10 15

Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val Ser Val
 20 25 30

Leu

<210> 21

<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 21
Ser Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile Ala Leu Leu
1 5 10 15
Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val Ser Val Leu
20 25 30

Thr

<210> 22
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 22
Lys Val Leu His Leu Glu Gly Glu Val Asn Lys Ile Ala Leu Leu Ser
1 5 10 15
Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val Ser Val Leu Thr
20 25 30

Ser

<210> 23
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 23

Leu Glu Gly Glu Val Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala
1 5 10 15

Val Val Ser Leu Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu
20 25 30

Asp

<210> 24

<211> 33

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 24

Gly Glu Val Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val
1 5 10 15

Ser Leu Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu
20 25 30

Lys

<210> 25

<211> 33

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 25

Glu Val Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser
1 5 10 15

Leu Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys
20 25 30

Asn

<210> 26
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 26
Val Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu
1 5 10 15

Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn
20 25 30

Tyr

<210> 27
<211> 33
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 27
Asn Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser
1 5 10 15

Asn Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr
20 25 30

Ile

<210> 28
<211> 33
<212> PRT
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 28

Lys Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn
1 5 10 15

Gly Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr Ile
20 25 30

Asp

<210> 29

<211> 33

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 29

Ile Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly
1 5 10 15

Val Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr Ile Asp
20 25 30

Lys

<210> 30

<211> 33

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 30

Ala Leu Leu Ser Thr Asn Lys Ala Val Val Ser Leu Ser Asn Gly Val
1 5 10 15

Ser Val Leu Thr Ser Lys Val Leu Asp Leu Lys Asn Tyr Ile Asp Lys
20 25 30

Gln

<210> 31

<211> 70

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 31

Gly Thr Ile Ala Leu Gly Val Ala Thr Ser Ala Gln Ile Thr Ala Ala
1 5 10 15

Val Ala Leu Val Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu
20 25 30

Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser
35 40 45

Ser Ile Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln Asp Tyr Val
50 55 60

Asn Lys Glu Ile Val Pro
65 70

<210> 32

<211> 56

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 32

Tyr Thr Pro Asn Asp Ile Thr Leu Asn Asn Ser Val Ala Leu Asp Pro
1 5 10 15

Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu
20 25 30

<400> 37

Ser Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala
1 5 10 15

Lys Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln
20 25 30

Lys Leu Asp
35

<210> 38

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 38

Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys
1 5 10 15

Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys
20 25 30

Leu Asp Ser
35

<210> 39

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 39

Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser
1 5 10 15

Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu
20 25 30

Asp Ser Ile
35

<210> 40
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 40
Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp
1 5 10 15
Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp
20 25 30

Ser Ile Gly
35

<210> 41
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 41
Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu
1 5 10 15
Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser
20 25 30

Ile Gly Asn
35

<210> 42
<211> 35
<212> PRT
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 42

Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu
1 5 10 15

Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile
20 25 30

Gly Asn Trp
35

<210> 43

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 43

Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu
1 5 10 15

Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly
20 25 30

Asn Trp His
35

<210> 44

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 44

Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser
1 5 10 15

Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly Asn
20 25 30

Trp His Gln
35

<210> 45
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 45
Ile Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys
1 5 10 15

Glu Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly Asn Trp
20 25 30

His Gln Ser
35

<210> 46
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 46
Ser Ile Glu Leu Asn Lys Ala Lys Ser Asp Leu Glu Glu Ser Lys Glu
1 5 10 15

Trp Ile Arg Arg Ser Asn Gln Lys Leu Asp Ser Ile Gly Asn Trp His
20 25 30

Gln Ser Ser
35

[illegible]

Ser Ser Thr
35

<220>

<223> Description of Artificial Sequence: synthetic peptide

Ser Thr Thr
35

<220>
<223> Description of Artificial Sequence: synthetic peptide

<400> 49

Thr Ala Ala Val Ala Leu Val Glu Ala Lys Gln Ala Arg Ser Asp Ile
1 5 10 15

Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser
20 25 30

Val Gln Ser
35

<210> 50

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 50

Ala Val Ala Leu Val Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys
1 5 10 15

Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln
20 25 30

Ser Ser Ile
35

<210> 51

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 51

Leu Val Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu
1 5 10 15

Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile
20 25 30

Gly Asn Leu
35

<210> 52
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 52
Val Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala
1 5 10 15
Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly
20 25 30

Asn Leu Ile
35

<210> 53
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 53
Glu Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile
1 5 10 15
Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn
20 25 30

Leu Ile Val
35

<210> 54
<211> 35
<212> PRT
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 54

Ala Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg
1 5 10 15

Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu
20 25 30

Ile Val Ala
35

<210> 55

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 55

Lys Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp
1 5 10 15

Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile
20 25 30

Val Ala Ile
35

<210> 56

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 56

Gln Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr
1 5 10 15

Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile Val
20 25 30

Ala Ile Lys
35

<210> 57
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 57
Ala Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn
1 5 10 15

Lys Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile Val Ala
20 25 30

Ile Lys Ser
35

<210> 58
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 58
Arg Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys
1 5 10 15

Ala Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile Val Ala Ile
20 25 30

Lys Ser Val
35

<210> 59
<211> 35
<212> PRT
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 59

Ser Asp Ile Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala
1 5 10 15

Val Gln Ser Val Gln Ser Ser Ile Gly Asn Leu Ile Val Ala Ile Lys
20 25 30

Ser Val Gln
35

<210> 60
<211> 35
<212> PRT
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 60

Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val
1 5 10 15

Gln Ser Ser Ile Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln Asp
20 25 30

Tyr Val Asn
35

<210> 61
<211> 35
<212> PRT
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 61

Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln
1 5 10 15

Ser Ser Ile Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln Asp Tyr
20 25 30

Val Asn Lys
35

<210> 62

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 62

Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser Val Gln Ser Ser Ile
1 5 10 15

Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln Asp Tyr Val Asn Lys
20 25 30

Glu Ile Val
35

<210> 63

<211> 47

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 63

Thr Trp Gln Glu Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile
1 5 10 15

Thr Ala Leu Leu Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr
20 25 30

Glu Leu Gln Lys Leu Asn Ser Trp Asp Val Phe Gly Asn Trp Phe
 35 40 45

<210> 64

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
 peptide

<400> 64

Trp Gln Glu Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr
 1 5 10 15

Ala Leu Leu Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu
 20 25 30

Leu Gln Lys
 35

<210> 65

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
 peptide

<400> 65

Gln Glu Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala
 1 5 10 15

Leu Leu Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu
 20 25 30

Gln Lys Leu
 35

<210> 66

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 66

Glu Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu
1 5 10 15

Leu Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln
20 25 30

Lys Leu Asn
35

<210> 67

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 67

Trp Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu
1 5 10 15

Glu Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys
20 25 30

Leu Asn Ser
35

<210> 68

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 68

Glu Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu
1 5 10 15

Glu Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu
20 25 30

Asn Ser Trp
35

<210> 69
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 69
Arg Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu
1 5 10 15

Ala Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn
20 25 30

Ser Trp Asp
35

<210> 70
<211> 35
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 70
Lys Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu Ala
1 5 10 15

Gln Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser
20 25 30

Trp Asp Val
35

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<400> 71
Val Asp Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu Ala Gln
  1                      5                      10                      15

Ile Gln Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp
      20                      25                      30

Asp Val Phe
      35

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<220>

<223> Description of Artificial Sequence: synthetic peptide

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<210> 73
<211> 35
<212> PRT
<213> Artificial Sequence
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31

<400> 73

Phe Leu Glu Glu Asn Ile Thr Ala Leu Leu Glu Glu Ala Gln Ile Gln
1 5 10 15

Gln Glu Lys Asn Met Tyr Glu Leu Gln Lys Leu Asn Ser Trp Asp Val
20 25 30

Phe Gly Asn
35

<210> 74

<211> 35

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 74

Pro Asp Ala Val Tyr Leu His Arg Ile Asp Leu Gly Pro Pro Ile Ser
1 5 10 15

Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys
20 25 30

Leu Glu Asp
35

<210> 75

<211> 34

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 75

Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys
1 5 10 15

Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln Ile Leu Arg Ser
20 25 30

Met Lys

<210> 76

<211> 34

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 76

Leu His Arg Ile Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp
1 5 10 15

Val Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu
20 25 30

Leu Leu

<210> 77

<211> 34

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 77

His Arg Ile Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val
1 5 10 15

Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu
20 25 30

Leu Glu

<210> 78

<211> 34

<212> PRT

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
peptide

<400> 78

Arg Ile Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly
1 5 10 15

Thr Asn Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu
20 25 30

Glu Ser

<210> 79

<211> 34

<212> PRT

<213> Artificial Sequence

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<223> Description of Artificial Sequence: synthetic
peptide

<400> 79

Ile Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr
1 5 10 15

Asn Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu
20 25 30

Ser Ser

<210> 80

<211> 34

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 80

Asp Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn
1 5 10 15

Leu Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser
 20 25 30

Ser Asp

<210> 81
 <211> 34
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: synthetic
 peptide

<400> 81
 Leu Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu
 1 5 10 15

Gly Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser
 20 25 30

Asp Gln

<210> 82
 <211> 34
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: synthetic
 peptide

<400> 82
 Gly Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly
 1 5 10 15

Asn Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp
 20 25 30

Gln Ile

<210> 83
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 83
Pro Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn
1 5 10 15
Ala Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln
20 25 30

Ile Leu

<210> 84
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 84
Pro Ile Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala
1 5 10 15
Ile Ala Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln Ile
20 25 30

Leu Arg

<210> 85
<211> 34
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: synthetic
peptide

<400> 85

Ser Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala Ile Ala
1 5 10 15

Lys Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln Ile Leu Arg
20 25 30

Ser Met

<210> 86

<211> 34

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
peptide

<400> 86

Leu Glu Arg Leu Asp Val Gly Thr Asn Leu Gly Asn Ala Ile Ala Lys
1 5 10 15

Leu Glu Ala Lys Glu Leu Leu Glu Ser Ser Asp Gln Ile Leu Arg Ser
20 25 30

Met Lys